



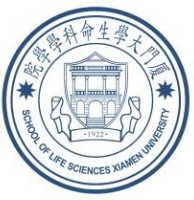
廈門大學
生|命|科|學|學|院
SCHOOL OF LIFE SCIENCES XIAMEN UNIVERSITY

MICROBIOLOGY

微生物學

Lecture 12

Yuan Jing
2017



廈門大學
生|命|科|学|学|院
SCHOOL OF LIFE SCIENCES XIAMEN UNIVERSITY

MICROBIOLOGY

Chapter 17 Recombinant DNA technology

Chapter X Discovery of CRISPR/Cas9

Yuan Jing

Outline

- **Genetic engineering by DNA recombination**
 - **Genomics in Microbiology**
 - **CRISPR/Cas9: History and Future**
-
-

15.1 Key developments in recombinant DNA technology

- ✓ **Restrictions enzymes**
- ✓ **DNA ligase**
- ✓ **PCR**
- ✓ **DNA Recombination**

Restrictions enzymes



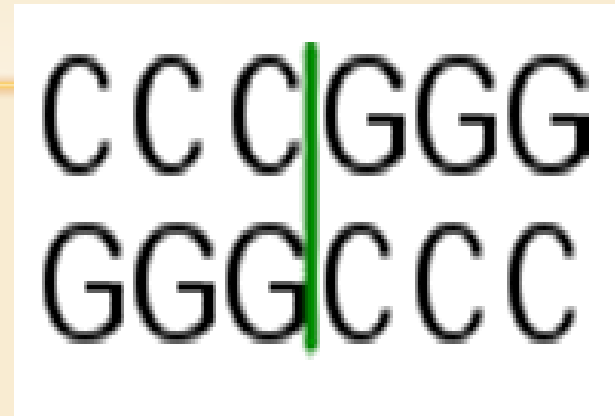
Restrictions enzymes

A restriction enzyme (or restriction endonuclease) is an enzyme that cuts DNA at or near specific recognition nucleotide sequences known as restriction sites.

EcoRI



SmaI



palindromic 回文序列 recognition site

Restrictions enzymes

Enzyme	Source	Recognition Sequence	Cut
<i>EcoRI</i>	<i>Escherichia coli</i>	5' GAATTC 3' CTTAAG	5' ---G AATTC---3' 3' ---CTTAA G---5'
<i>EcoRII</i>	<i>Escherichia coli</i>	5' CCWGG 3' GGWCC	5' --- CCWGG---3' 3' ---GGWCC ---5'
<i>BamHI</i>	<i>Bacillus amyloliquefaciens</i> 芽孢杆菌	5' GGATCC 3' CCTAGG	5' ---G GATCC---3' 3' ---CCTAG G---5'
<i>HindIII</i>	<i>Haemophilus influenzae</i> 流感嗜血杆菌	5' AAGCTT 3' TTCGAA	5' ---A AGCTT---3' 3' ---TTCGA A---5'

The Nobel Prize in Physiology or Medicine 1978

Restrictions enzymes



Werner Arber



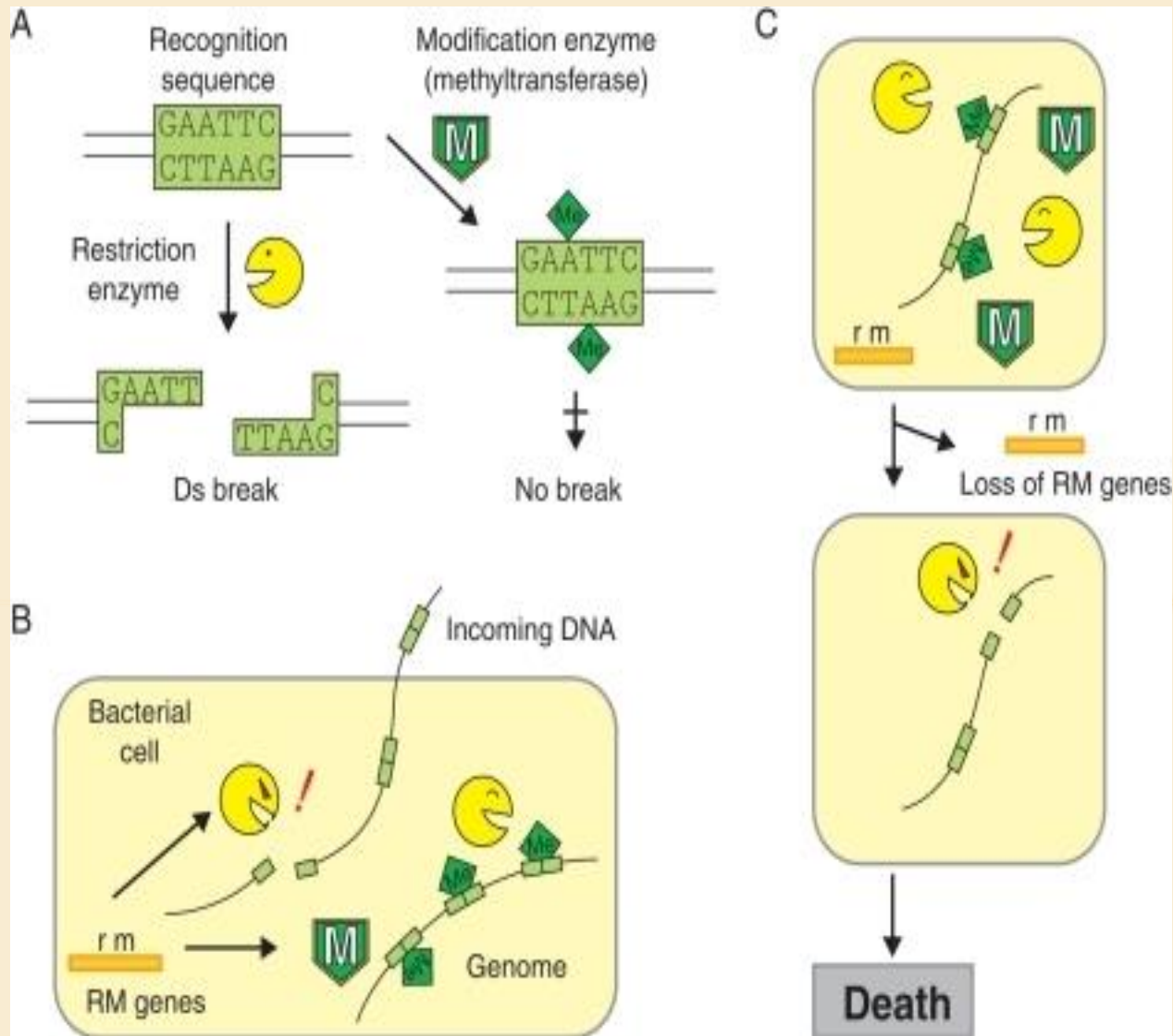
Daniel Nathans



Hamilton O. Smith

The Nobel Prize in Physiology or Medicine 1978 was awarded jointly to Werner Arber, Daniel Nathans and Hamilton O. Smith *"for the discovery of restriction enzymes and their application to problems of molecular genetics"*.

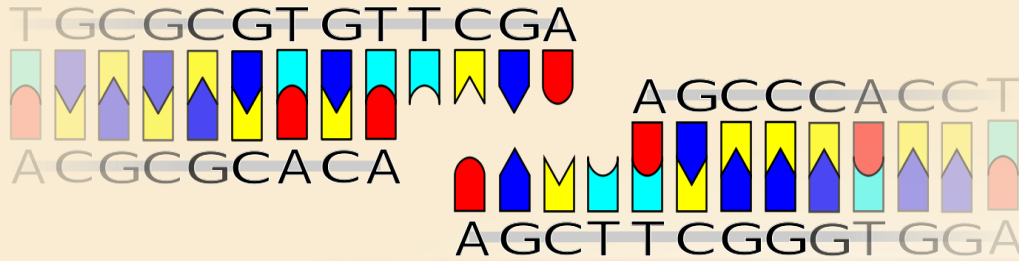
Bacterial Restriction–Modification Systems



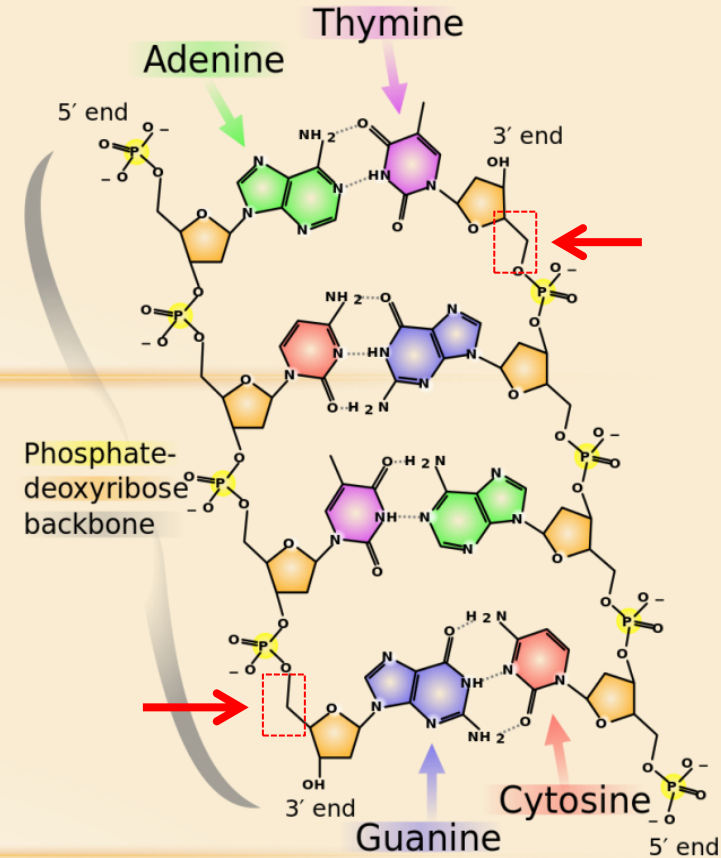
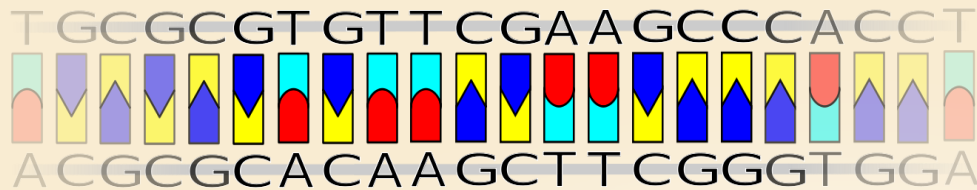
Restriction–modification systems allow bacterial cells to distinguish between their own DNA and any foreign DNA entering the cell, and to destroy the latter. They operate through two enzyme activities: a **restriction endonuclease** that cleaves the foreign DNA, and a modification **methyltransferase** that protects the host DNA.

DNA Ligase DNA 连接酶

DNA ligase is an enzyme that facilitates the joining of DNA strands together by catalyzing the formation of a phosphodiester bond.



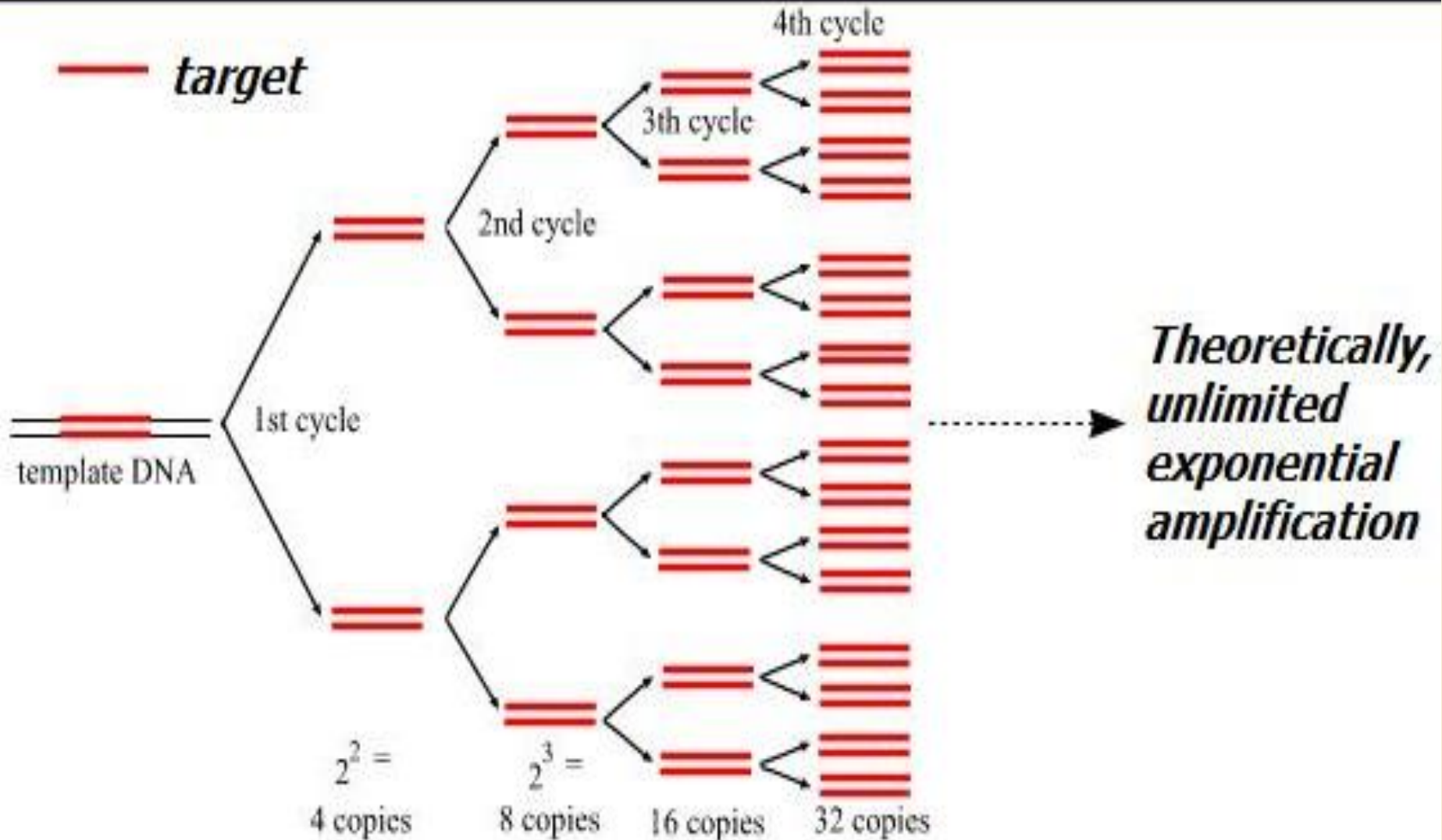
DNA Ligase



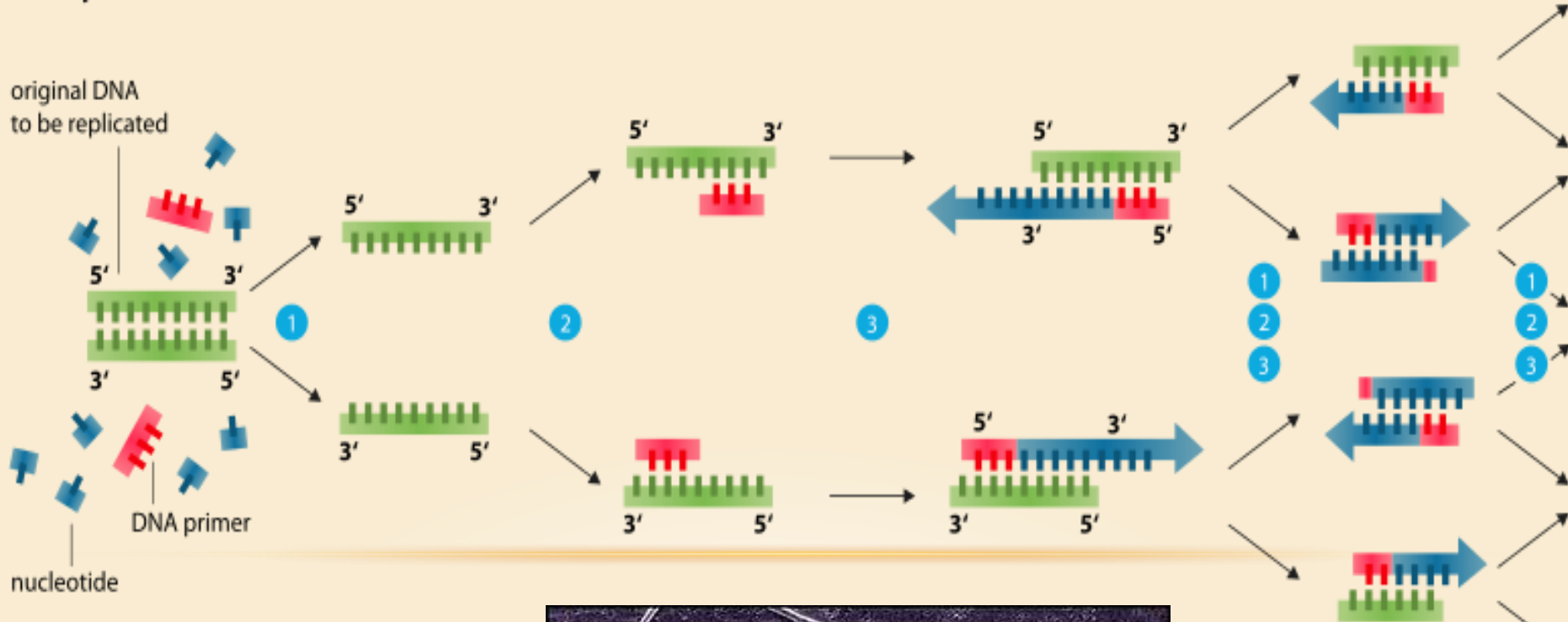


DNA Recombination

Polymerase Chain Reaction (PCR)



Polymerase Chain Reaction (PCR)



- 1 **Denaturation** at 94-96°C
- 2 **Annealing** at ~68°C
- 3 **Elongation** at ca. 72 °C



**DNA polymerase from
*Thermus aquaticus***

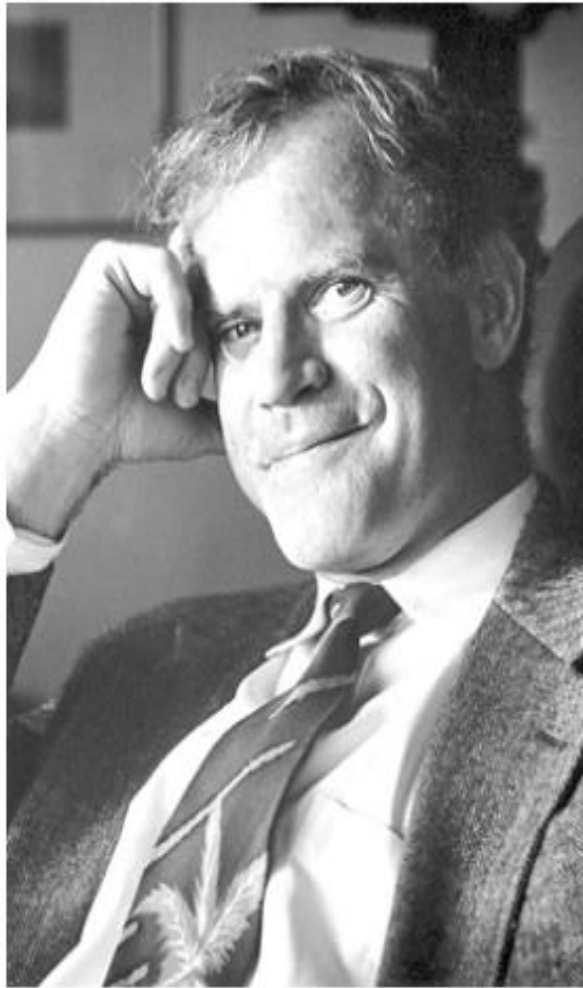
PCR { **Primer**
Polymerase
Buffer
Single Strand



Polymerase Chain Reaction (PCR)

The Nobel Prize in Chemistry 1993

Kary B. Mullis - Facts



Kary B. Mullis

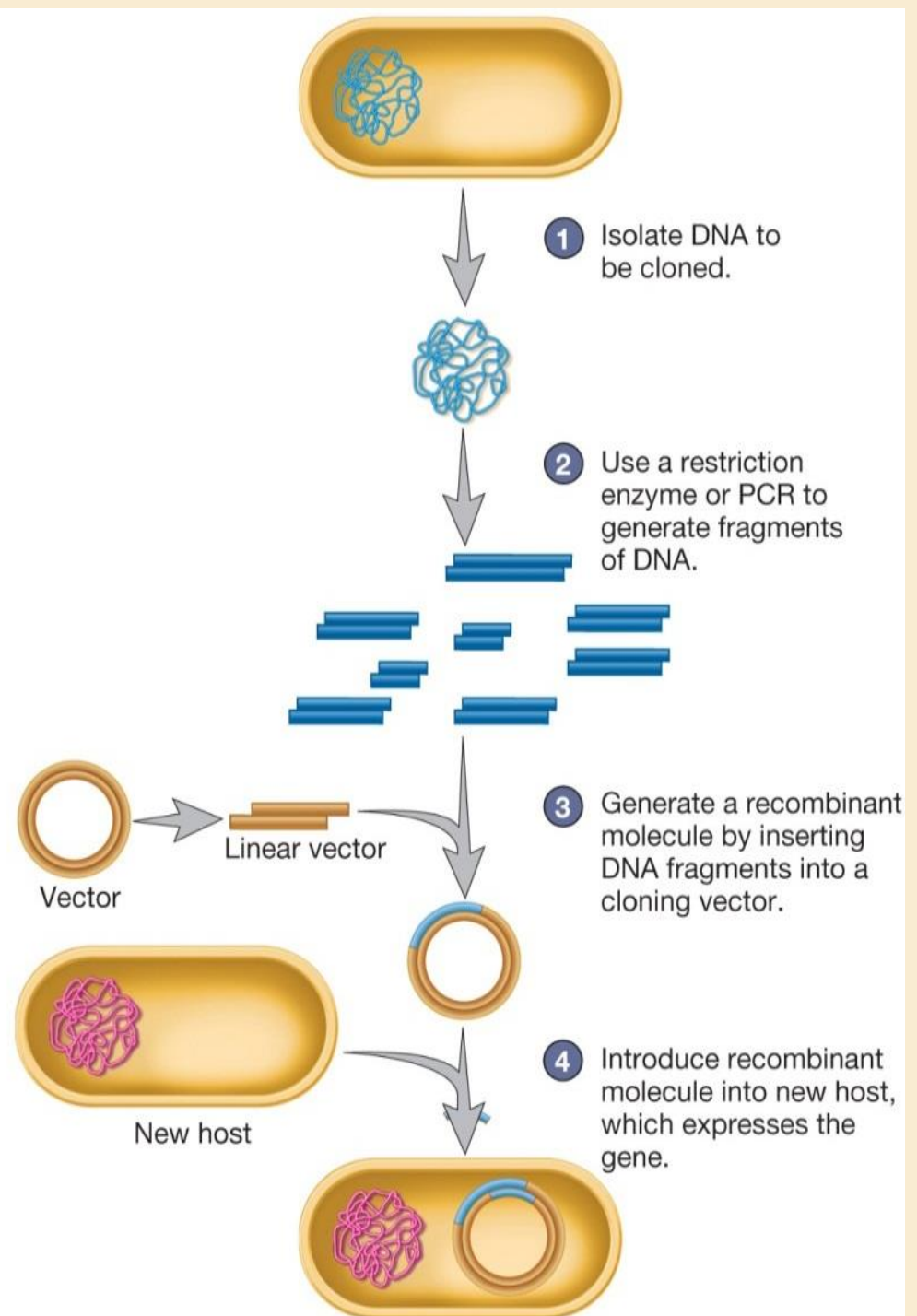
Born: 28 December 1944, Lenoir, NC, USA

Prize motivation: "for his invention of the polymerase chain reaction (PCR) method"

Field: biochemistry

- **Genetic engineering**
 - **deliberate modification of organism's genetic information by directly changing the sequence of nucleic acids in its genome**
 - **Recombinant DNA technology**
 - **procedures used to carry out genetic engineering**
-

Flowchart of Genetic Engineering



Cloning Vectors

Key

- there are four types of cloning vectors
 - Plasmids (most commonly used) 质粒
 - Phages and viruses
 - Cosmids (artificial)
 - Artificial chromosomes

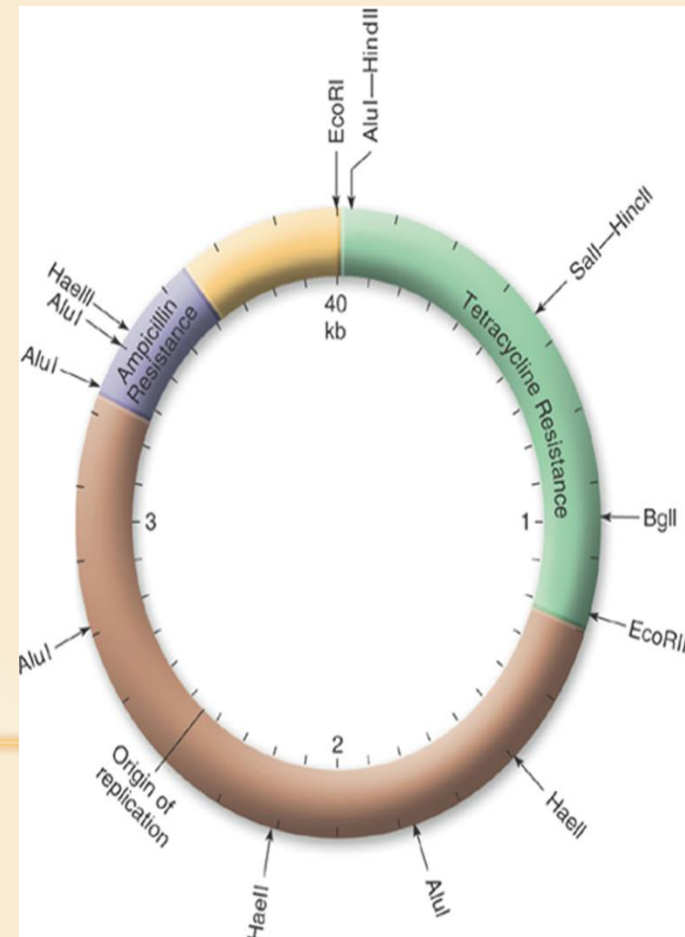


Table 15.3 Recombinant DNA Cloning Vectors

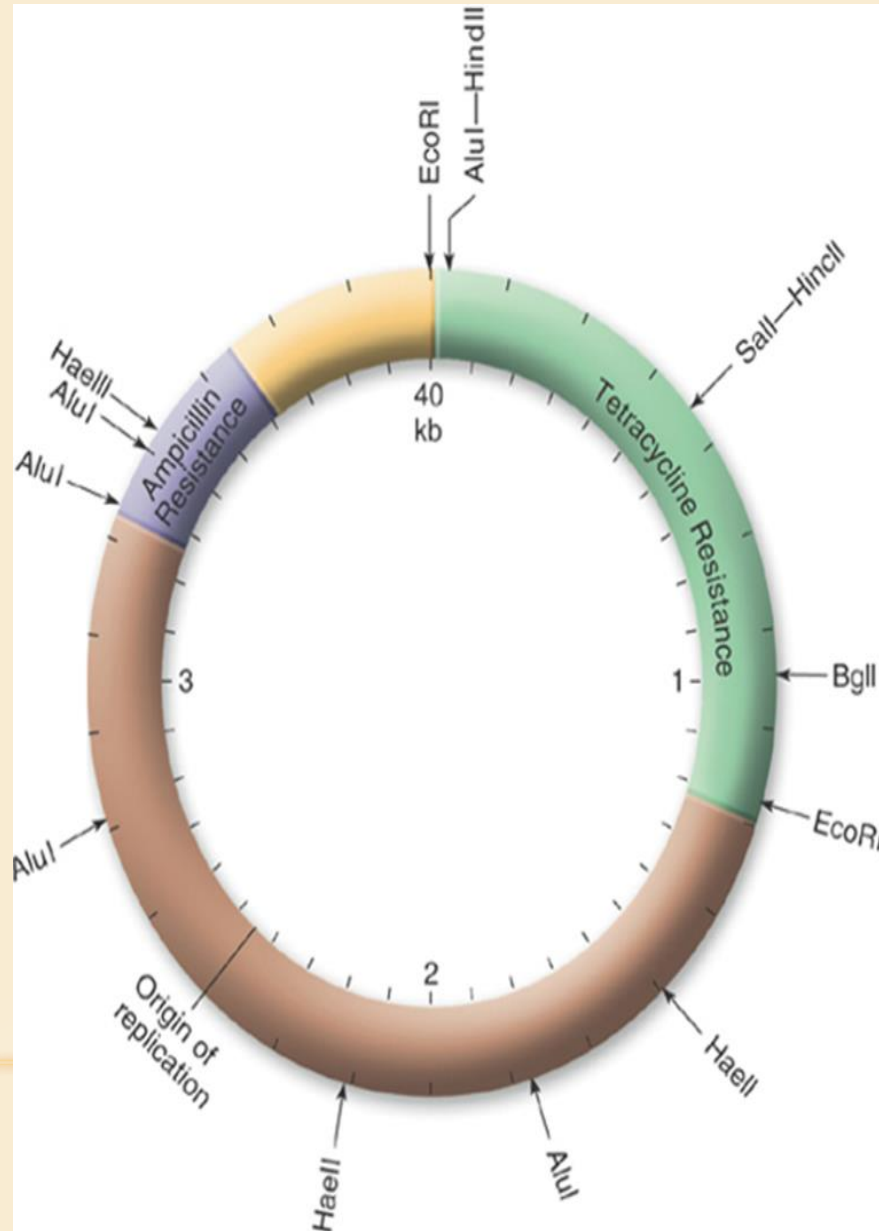
<i>Vector</i>	<i>Insert Size (kb, 1 kb = 1,000 bp)</i>	<i>Example</i>	<i>Features</i>
Plasmid	<20 kb	pBR322, pUC19	Replicates independently of microbial chromosome so many copies may be maintained in a single cell
Bacteriophage	9–25 kb	λ 1059, λ gt11, M13mp18, EMBL3	Packaged into lambda phage particles; single-stranded DNA viruses such as M13 have been modified (e.g., M13mp18) to generate either double- or single-stranded DNA in the host.
Cosmids	30–47 kb	pJC720, pSupercos	Can be packaged into lambda phage particles for efficient introduction into bacteria, then replicates as a plasmid
PACs (P1 artificial chromosomes)	75–100 kb	pPAC	Based on the bacteriophage P1 packaging mechanism
BACs (bacterial artificial chromosomes)	75–300 kb	pBAC108L	Modified F plasmid that can carry large DNA inserts; very stable within the cell
YACs (yeast artificial chromosomes)	100–1,000 kb	pYAC	Can carry largest DNA inserts; replicates in <i>Saccharomyces cerevisiae</i>

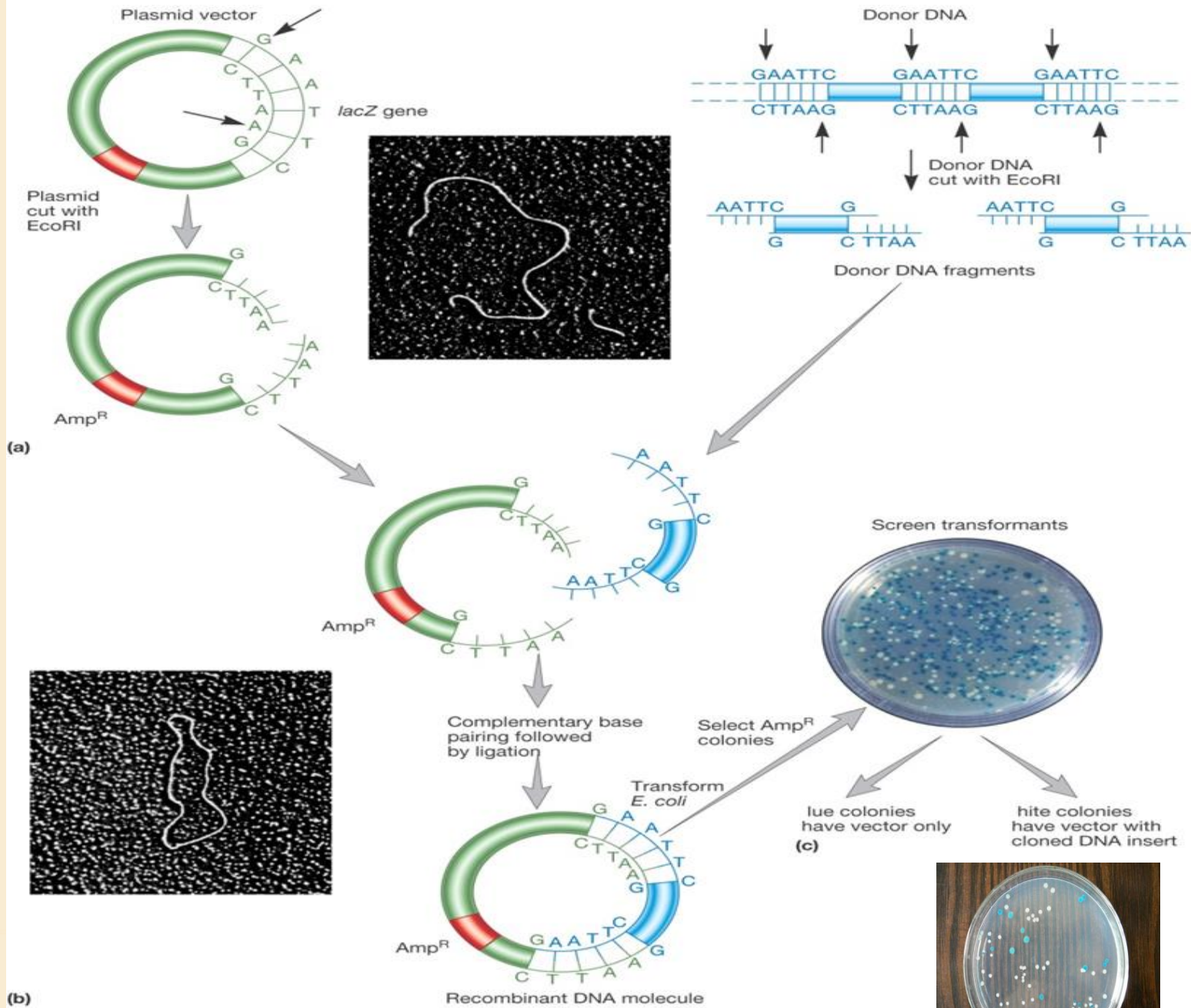
Plasmids

- Replicate autonomously and easy to purify
 - Requirements for vectors
 - an Origin of Replication
 - a Selectable Marker
 - a Multicloning site
 - site that allows DNA to be inserted to the plasmid vector
-

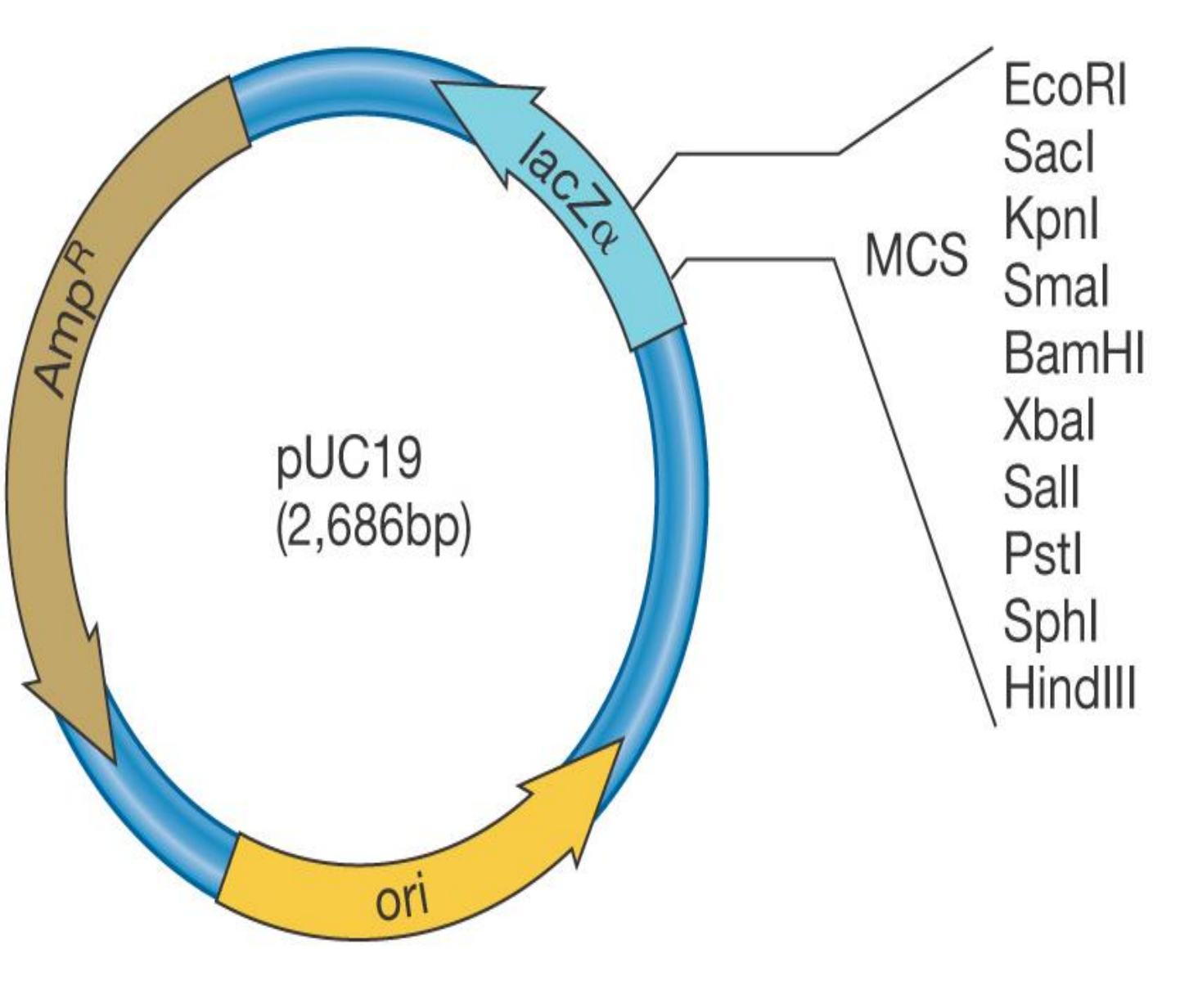
Plasmid as cloning vector

Key





Clone vector for Recombinants Screening

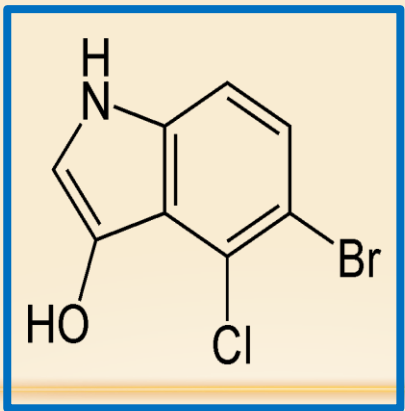
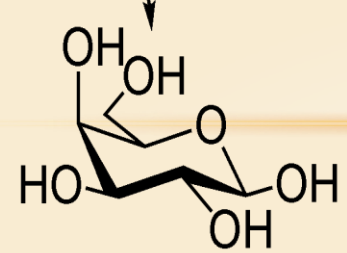
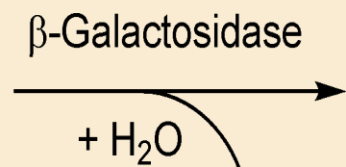


无色化合物(5-溴-4-氯-3-吲哚- β -D-半乳糖苷)

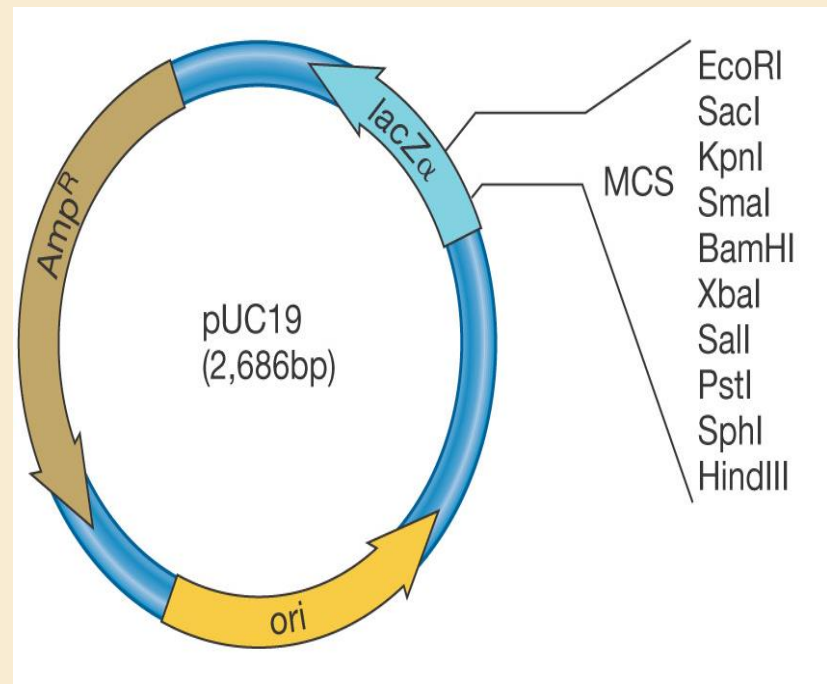
β -半乳糖苷酶

半乳糖 + 深蓝色化合物(5-溴-4-靛蓝)

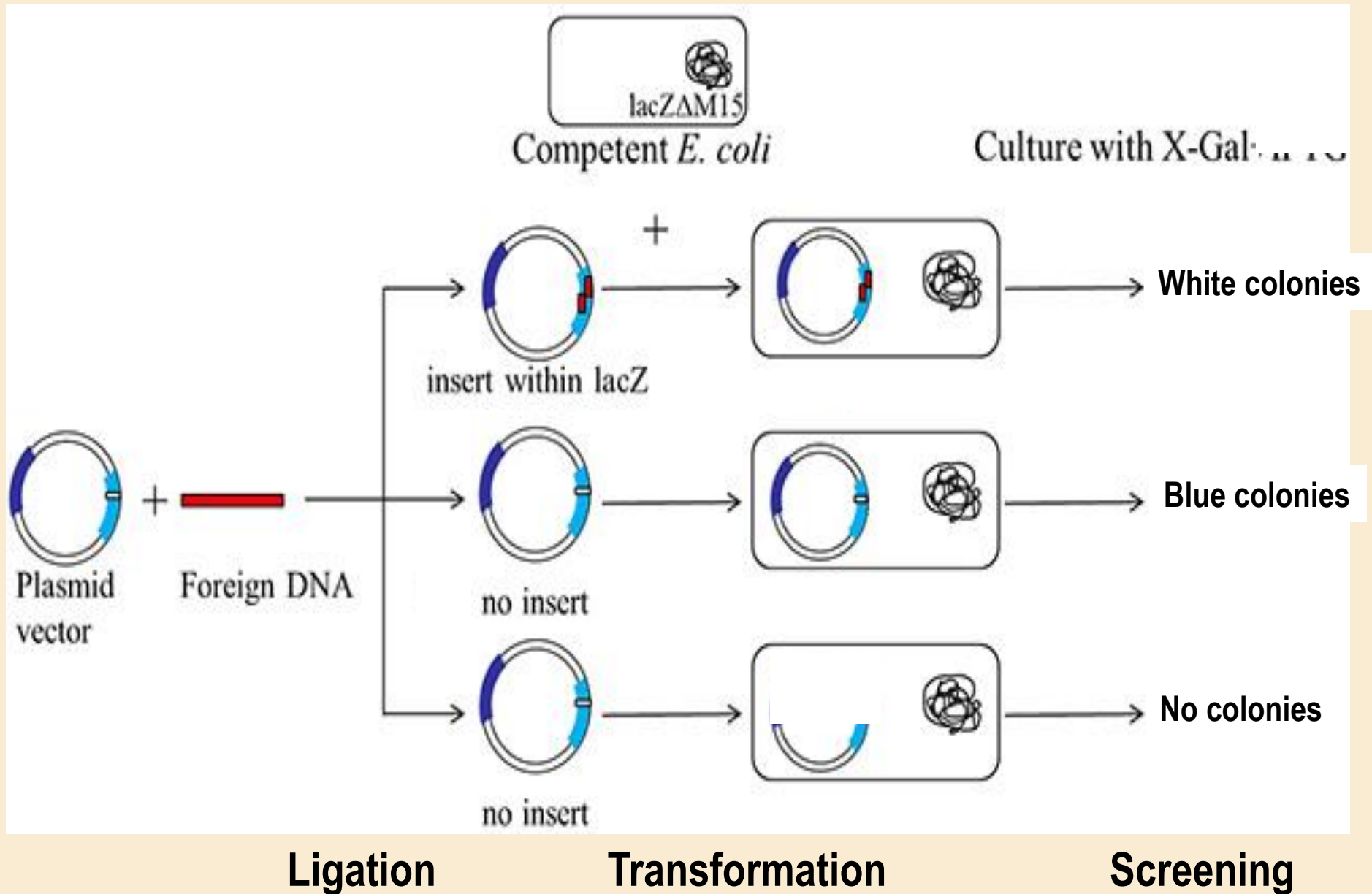
X-Gal



1



Blue White Screening for identifying the Recombinants



- **Most common used host cells**
 - ***E. coli* – bacteria**
 - ***S.cerevisiae* (酵母) – eukaryotic host**
-

Inserting recombinant DNA into eukaryotic host cells

- **DNA introduction into microbes**
 - transformation
 - electroporation 电击转化
 - gene gun 基因枪
 - Ti plasmid of *Agrobacterium tumefaciens* 农杆菌 (used to introduce foreign DNA into plant genomes)
-

Pioneer of Genetic Engineering



Stanley Cohen
Stanford University



Herbert Boyer
UCSF

Attempt to recombine gene from different bacteria into one DNA molecule



Bacterial resistance to antibiotics

Stanley Cohen

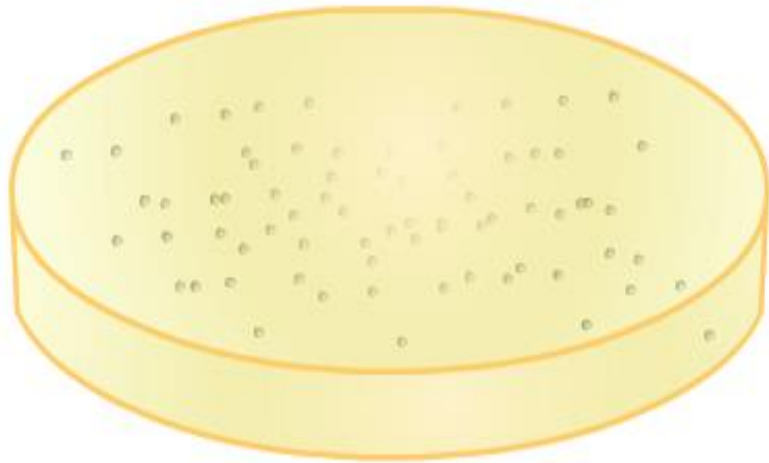


Herbert Boyer

Restriction Enzyme

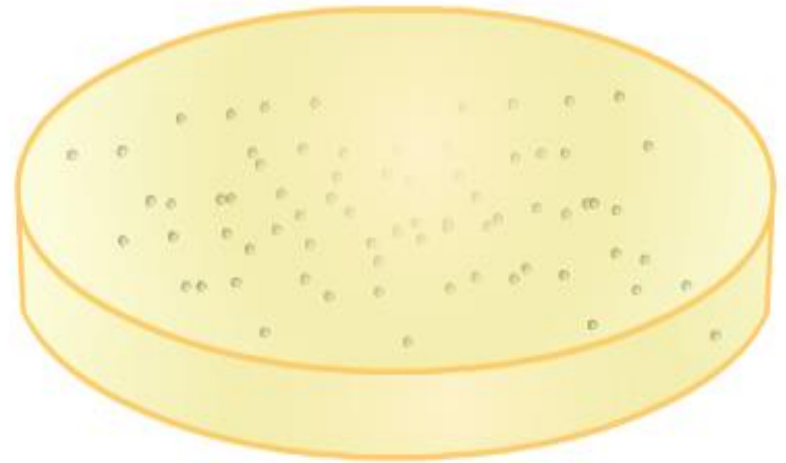
Hawaii, 1972

E.coli
BACTERIA



+TETRACYCLINE

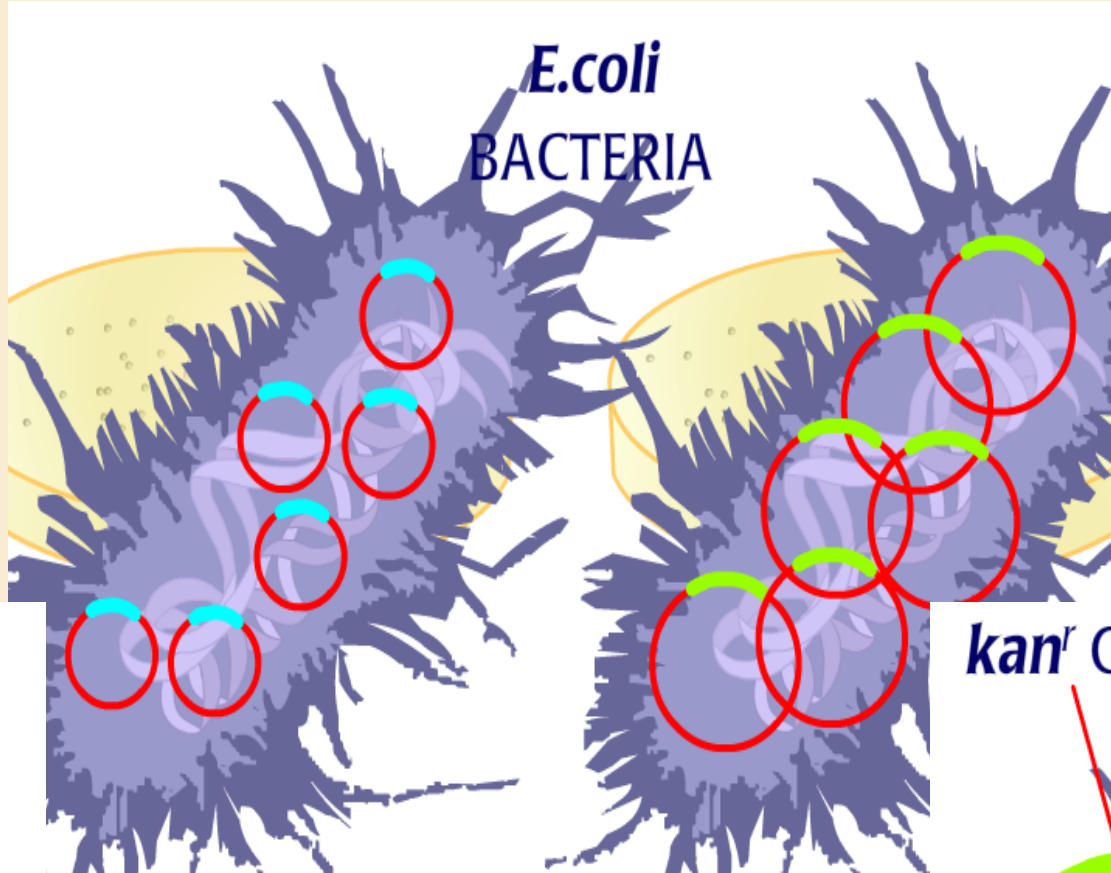
四环素



+KANAMYCIN

卡那霉素

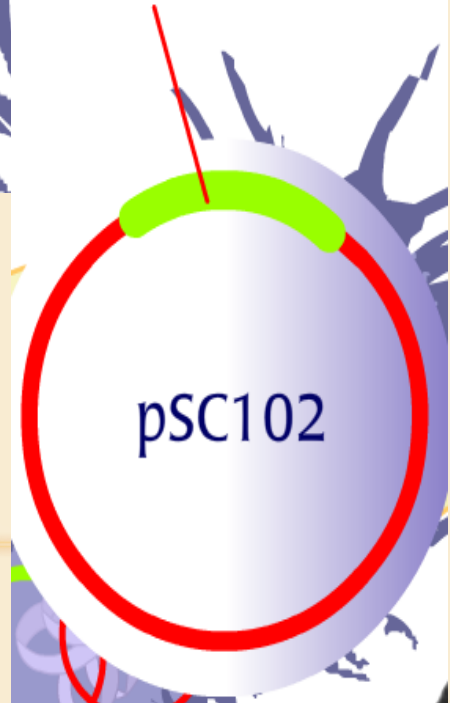
E.coli
BACTERIA



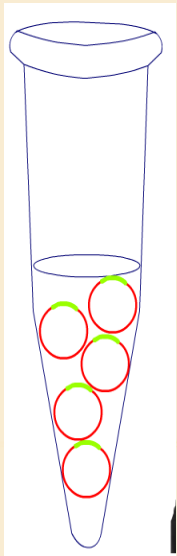
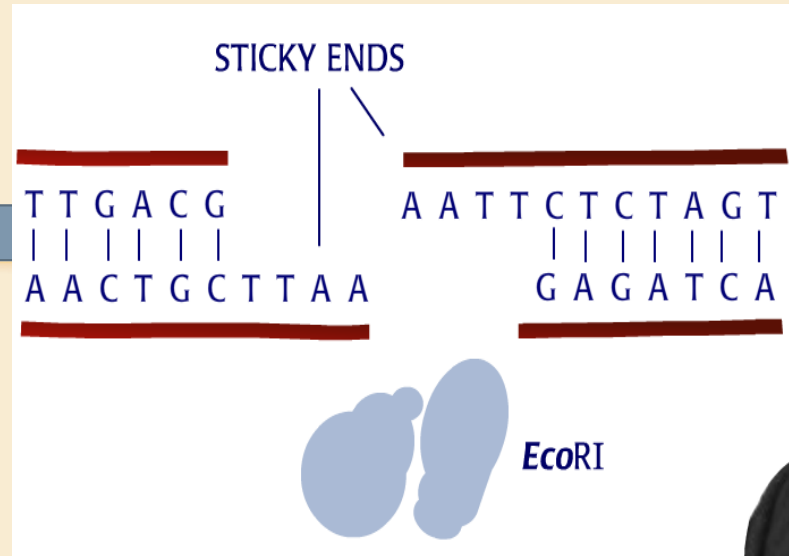
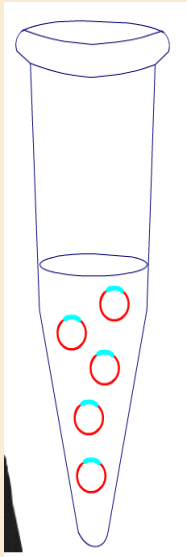
tet^r GENE



kan^r GENE

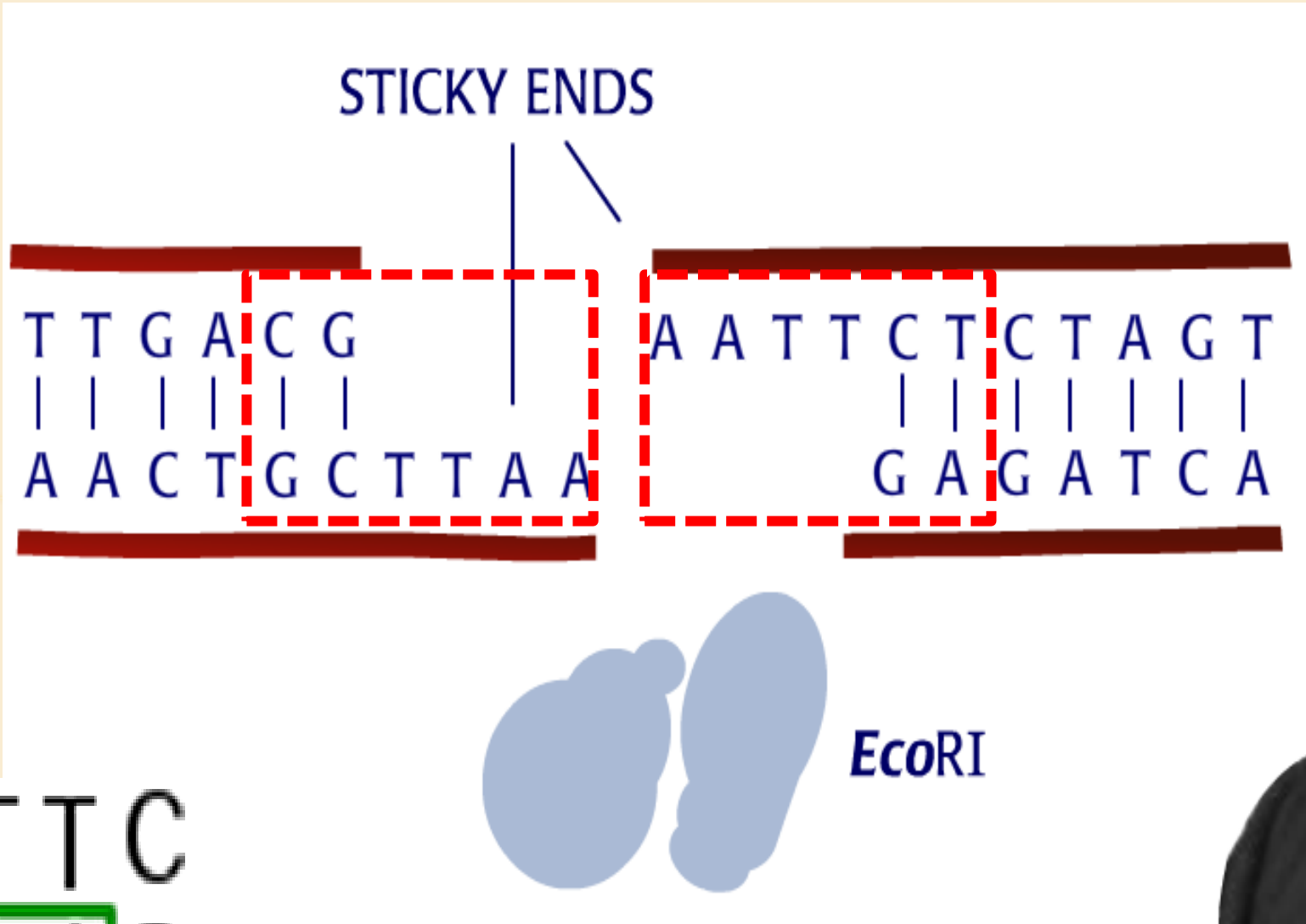


Plasmid digestion by Restriction enzyme 限制性内切酶酶切



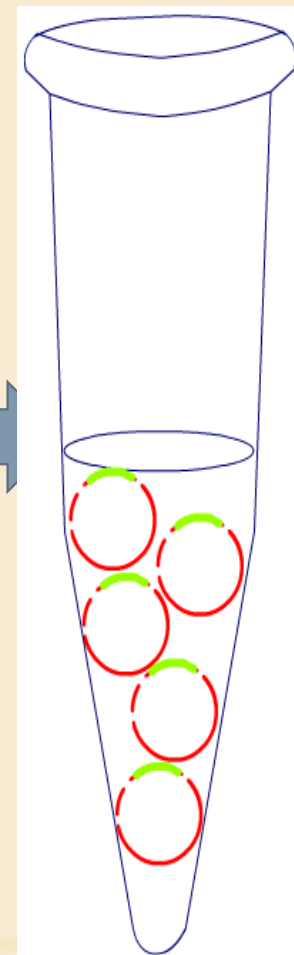
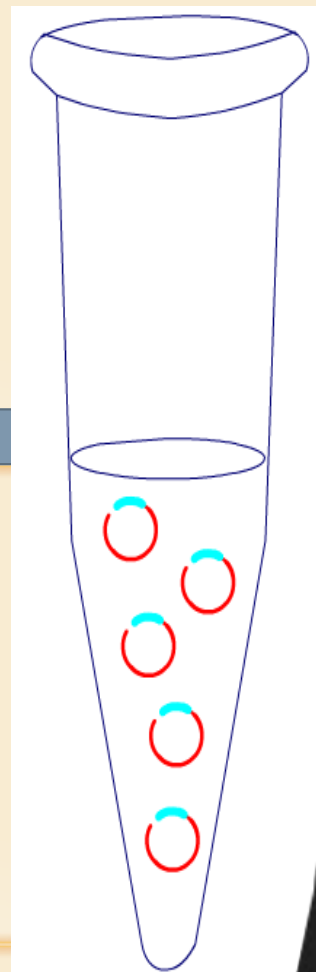
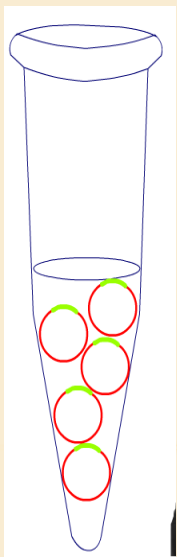
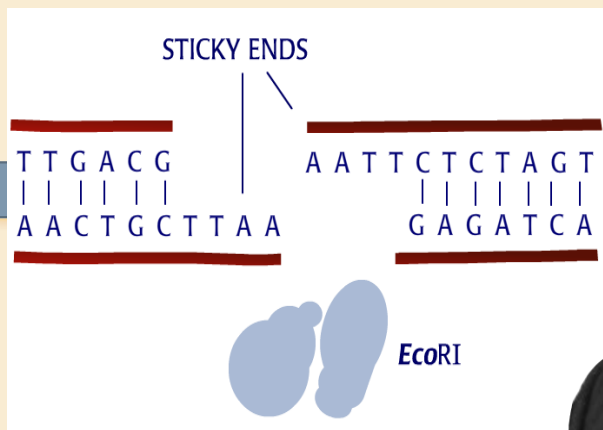
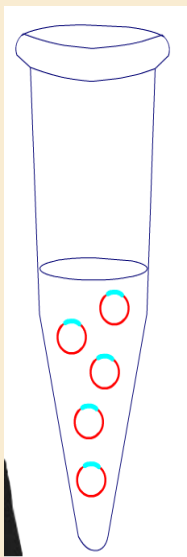
Restriction enzyme

EcoRI

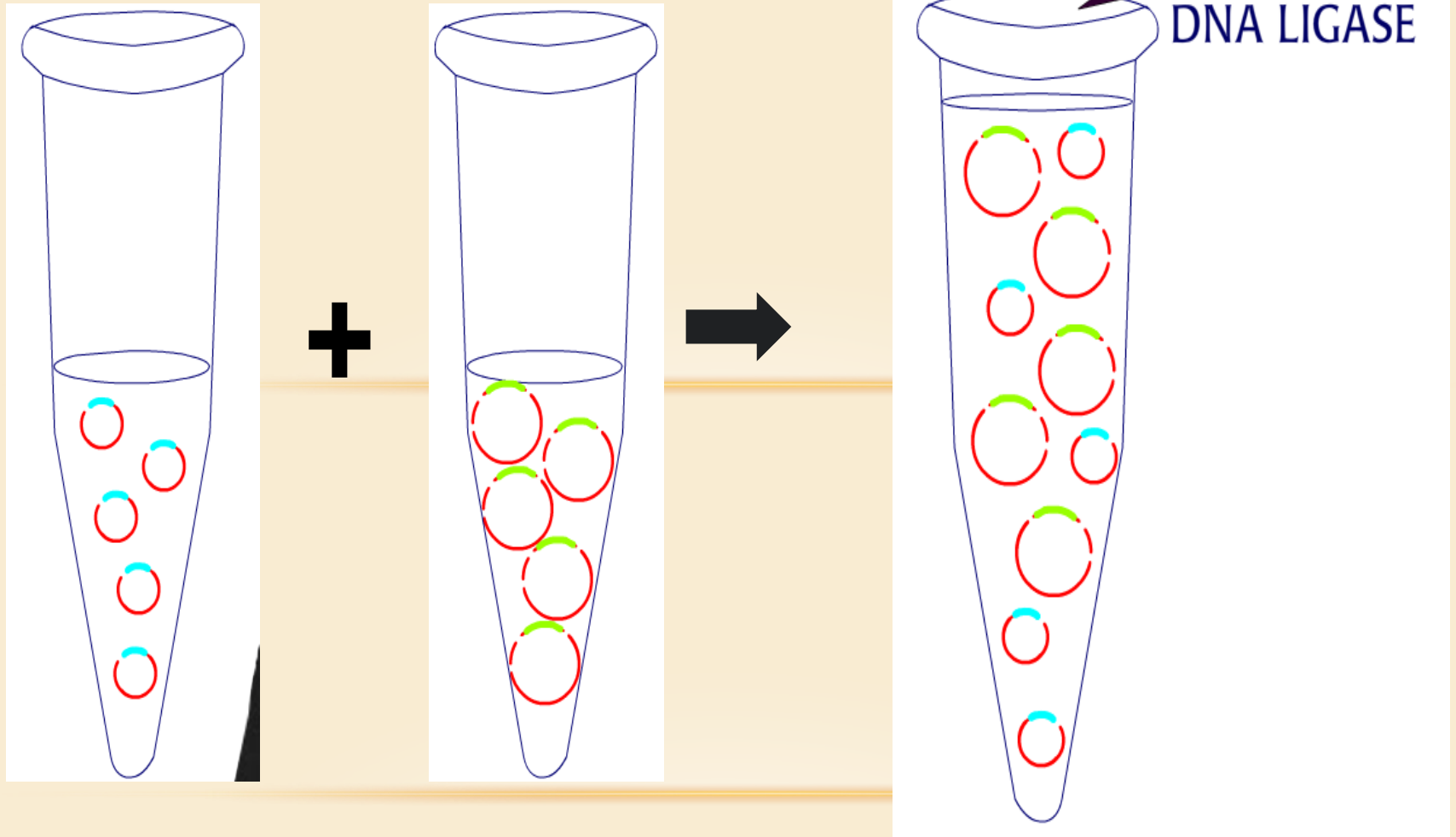


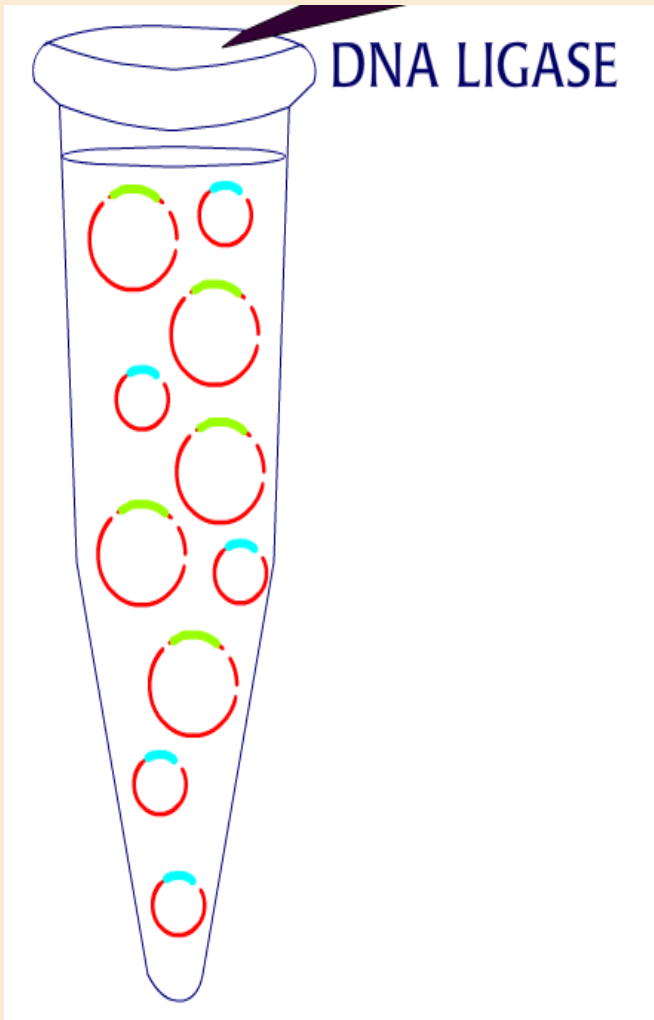
GAATTC
CTTAAG

Plasmid digestion by Restriction enzyme 限制性内切酶酶切



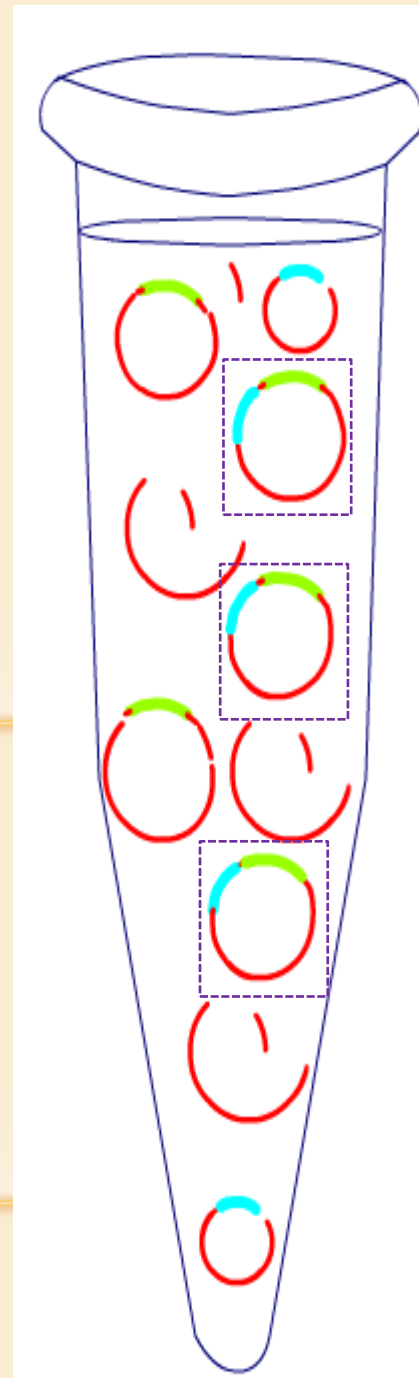
DNA Ligase DNA 连接酶



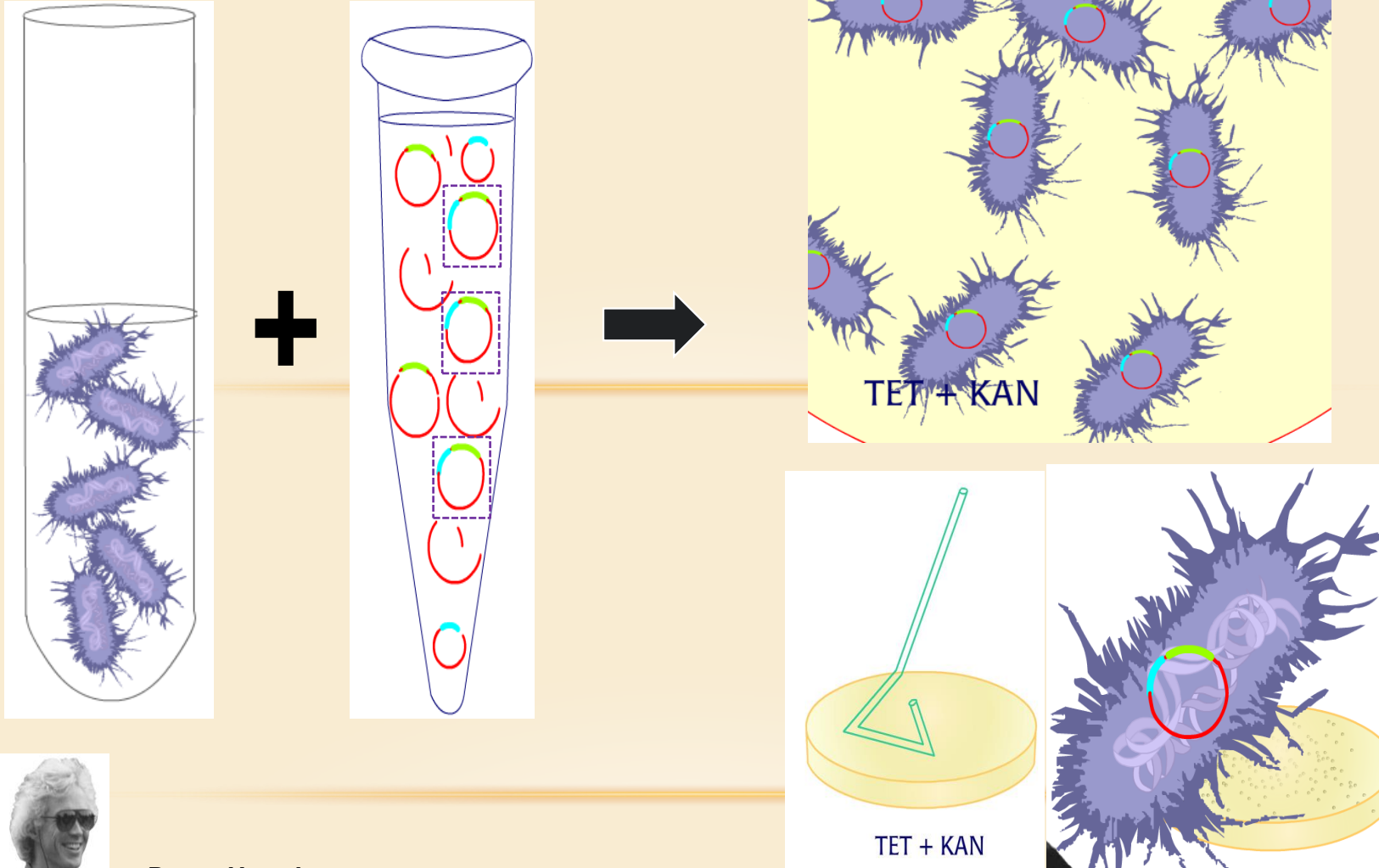


DNA Ligase

A thick black arrow pointing from the left tube to the right tube, indicating the action of DNA Ligase.

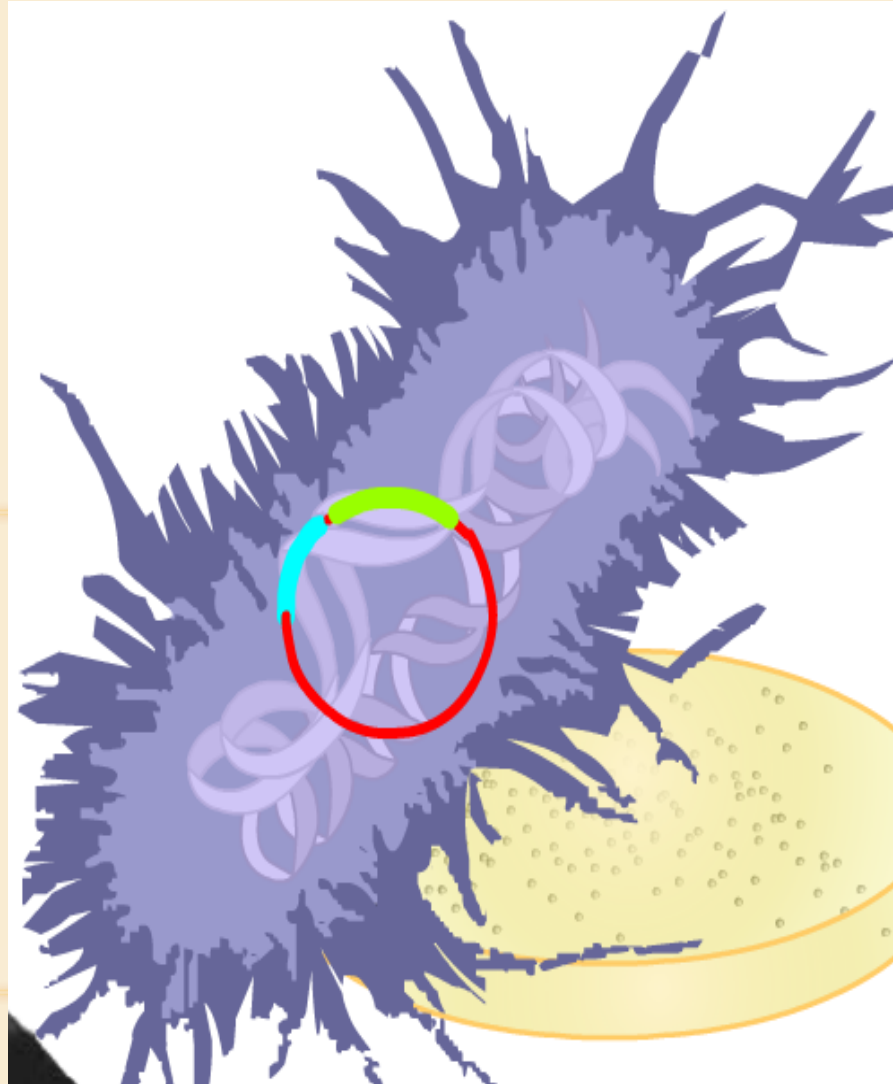


Plasmids transformed to Competent Cell

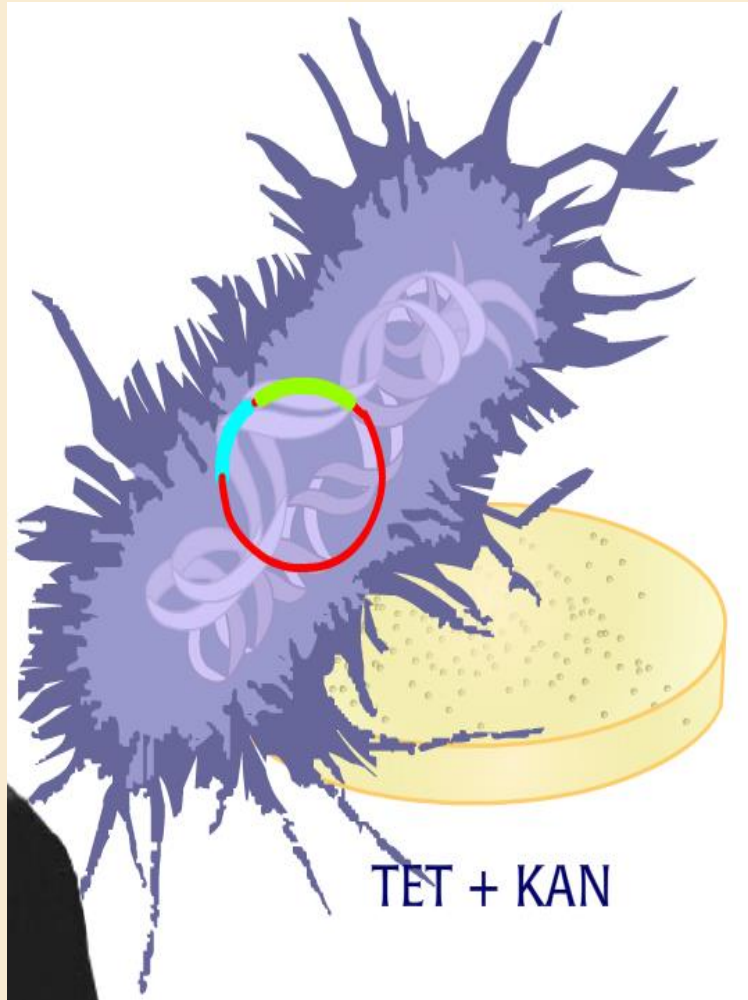


Doug Hanahan

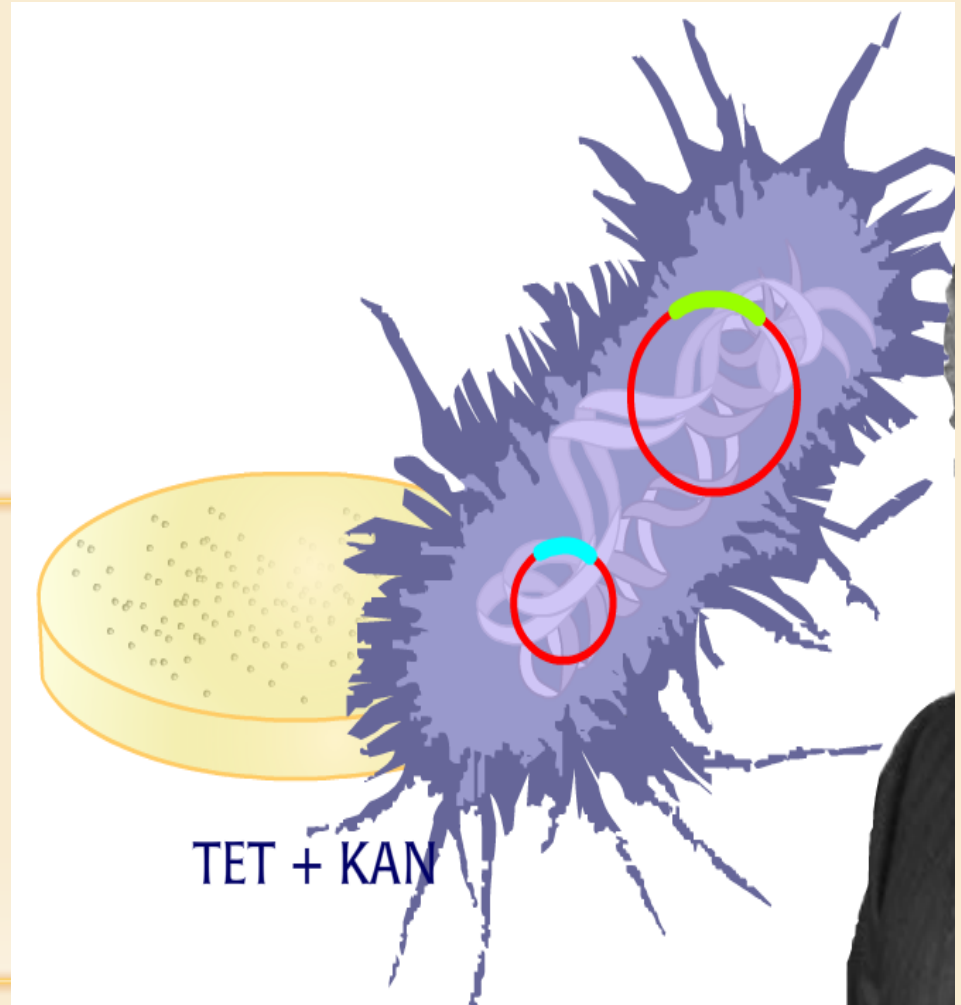
The First Recombinant Plasmid



One Possibility

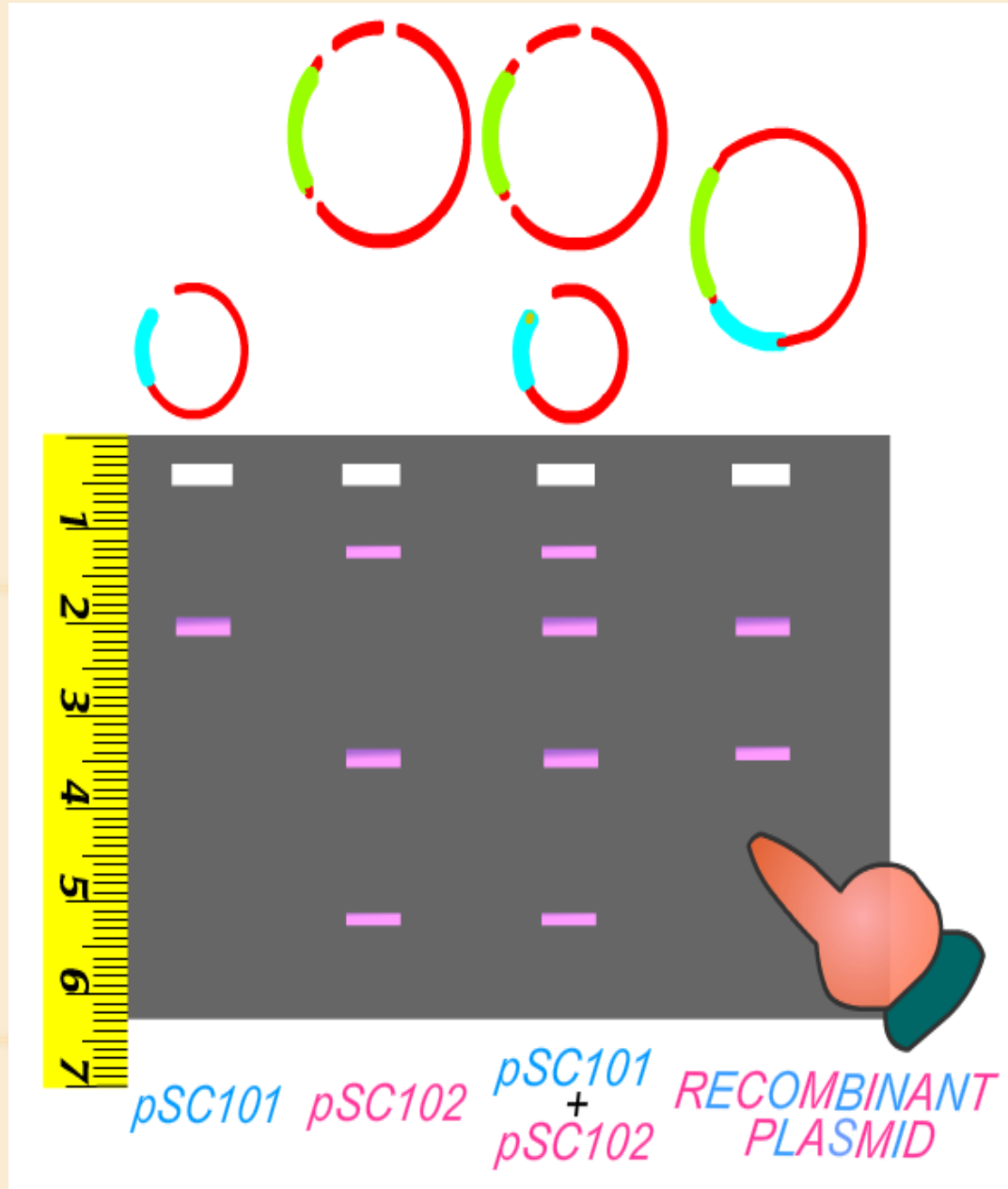


Recombinant Plasmid

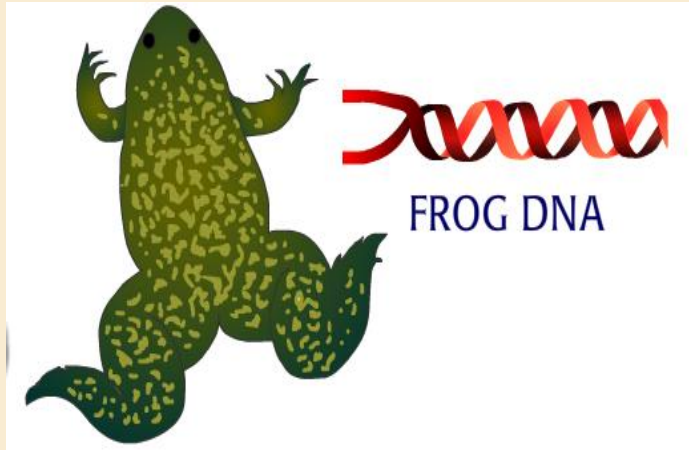


Two re-ligated plasmids

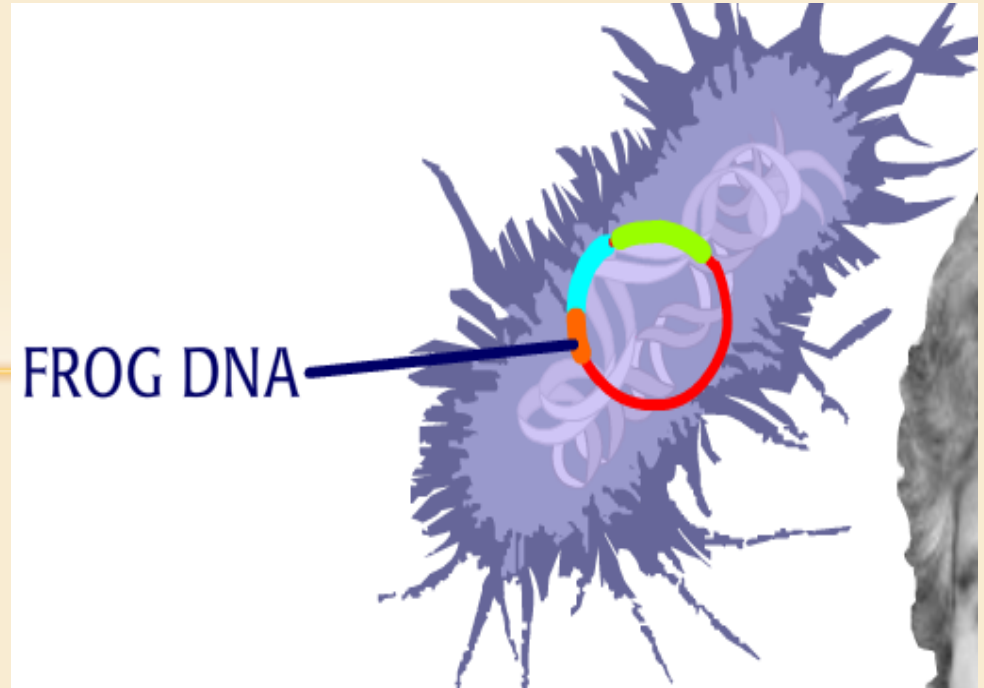
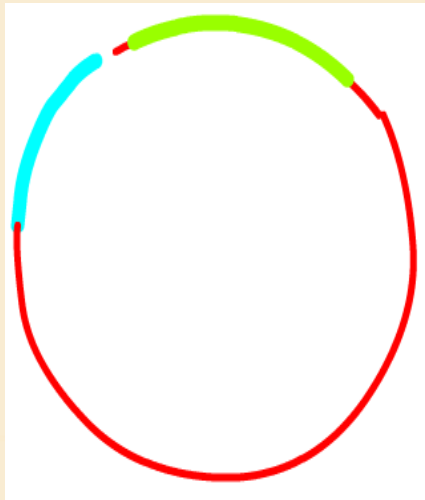
Recombinant Plasmid Characterization by Restriction Enzyme



Frog Gene recombined to the E.coli plasmid



+



Frog rRNA gene

First Time Genetic Engineering

Frog
ribosomal
RNA gene

EcoRI

Escherichia
coli plasmid
pSC101

Ligase

chemical cell
transformation
method

**Bacterial
cells**

Genetic Engineering invented by “Boyer and Cohen” a quick and easy way to make chemicals like:

HGH (human growth hormone),

synthetic insulin,

factor VIII for hemophilia 血友病,

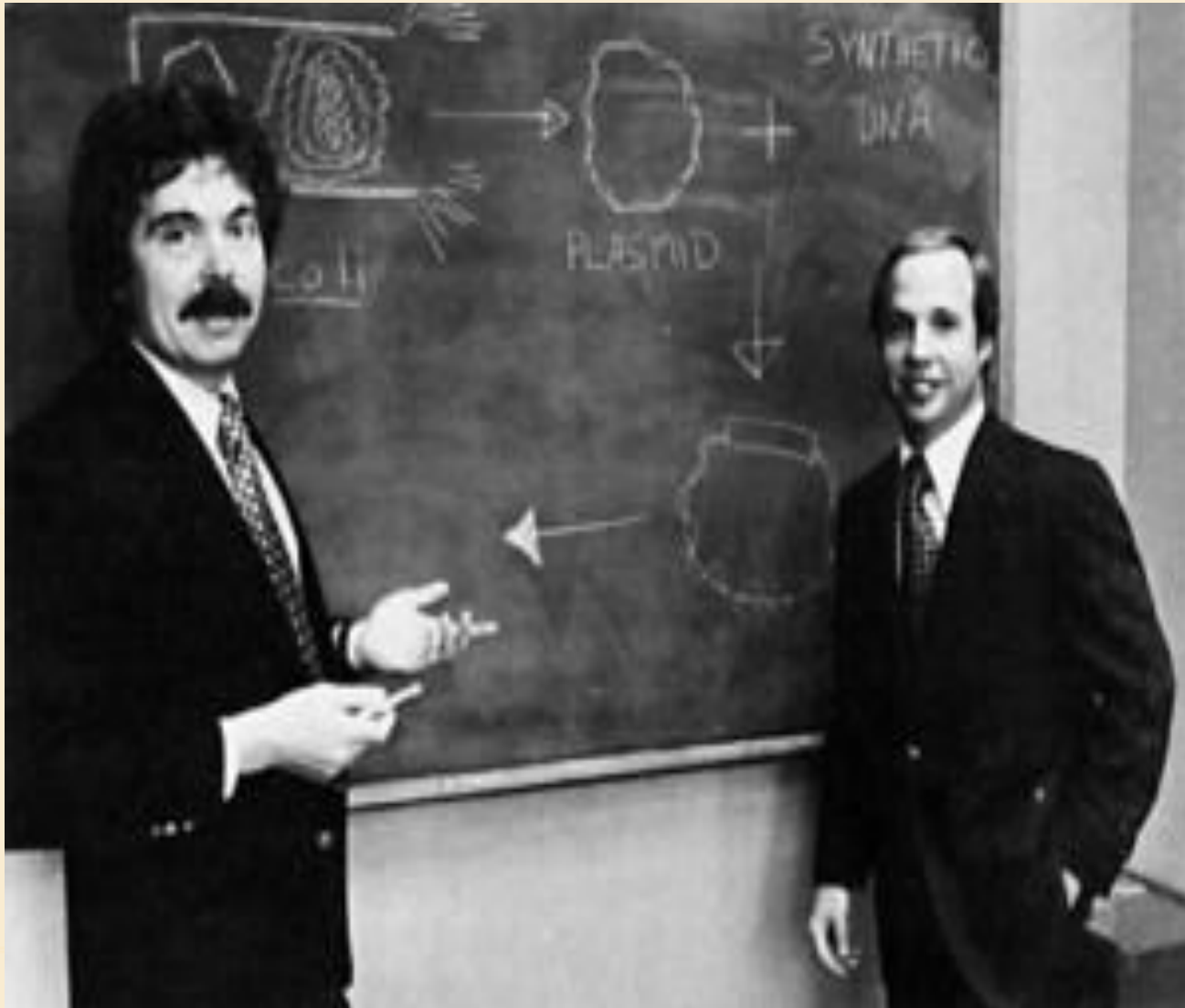
somatostatin 生长抑制素 for acromegaly 肢端肥大症

clot-dissolving agent tissue plasminogen activator (tPA)

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Bio-Science and Technology Make Fortune



Herbert W. Boyer

Robert A. Swanson

Founder in 1976

Genentech
IN BUSINESS FOR LIFE

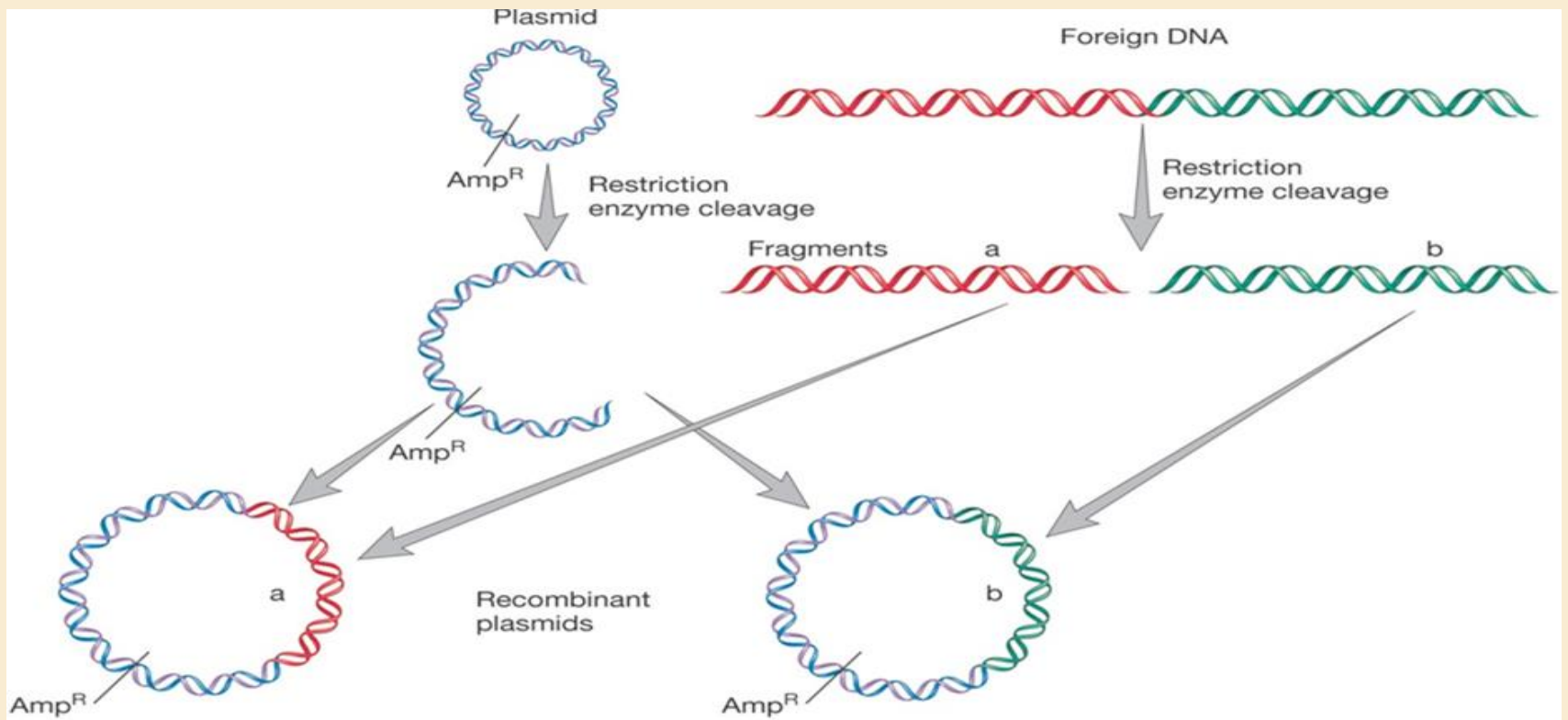
First Recombinant Insulin Protein



1978

Genentech produced the world's first genetically engineered human insulin. They engineered bacteria that produced human insulin whereas previous methods for obtaining insulin involved taking it from animals.

- 1978 genetically cDNA fragment coding for human insulin 胰岛素 recombined into *E. coli* vector
- produced new source of human insulin
- first commercial use for recombinant protein



Transform auxotroph with plasmids.



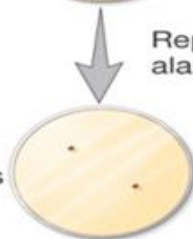
Plate bacteria and grow separate clones.

Select transformants on complete medium containing ampicillin.



Master plate

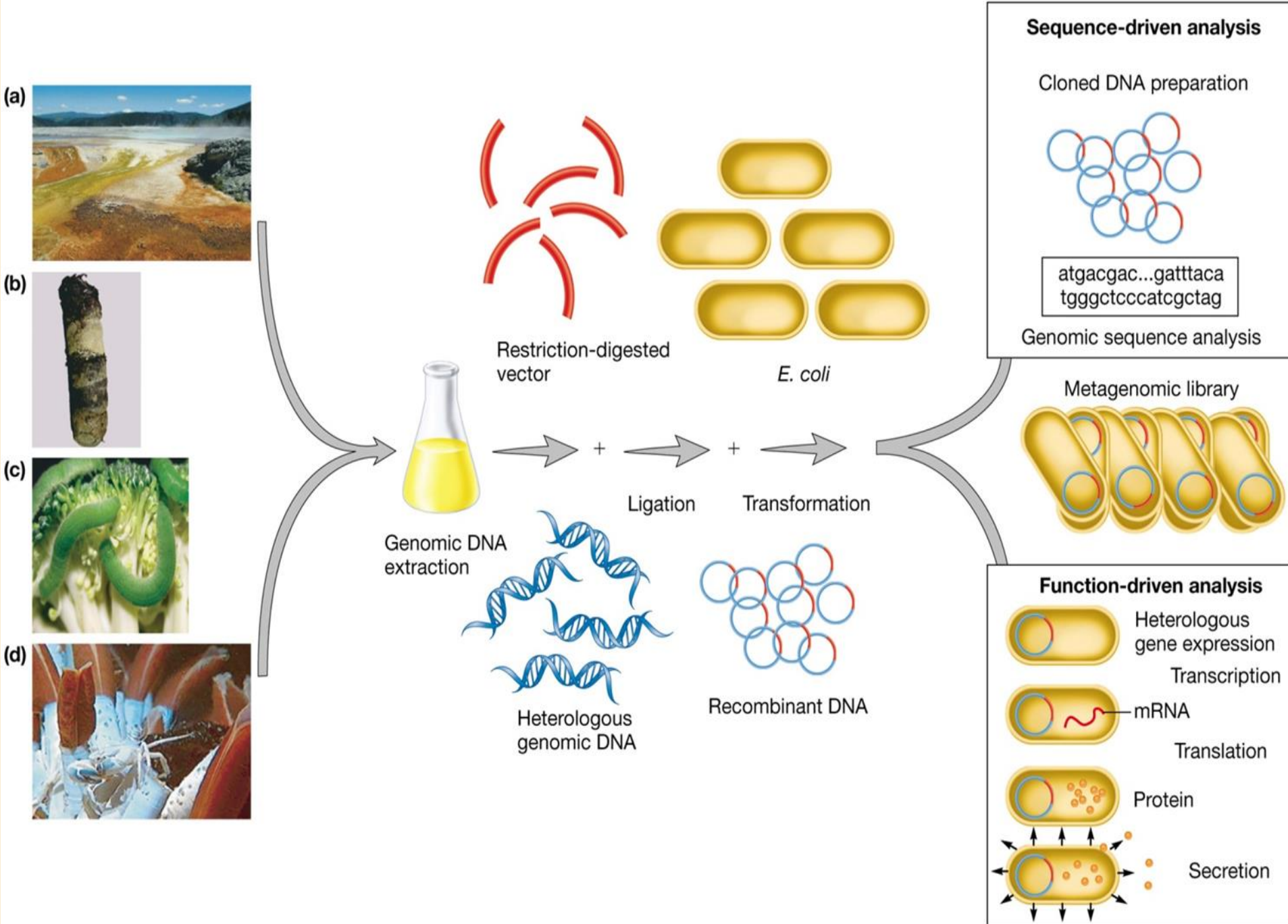
Replica plate onto medium lacking alanine but containing ampicillin



Screen genomic library transformants for colonies with alanine biosynthetic gene.

Only transformants containing the cloned alanine biosynthetic gene grow.

丙氨酸



Outline

- **DNA/RNA are the genetic materials**
- **Transcription regulation of Operon**
- **DNA mutation**
- **DNA repair**
- **Horizontal Gene Transfer & Transposition**
- **Genetic engineering by DNA recombination**
- **Genomics in Microbiology**

Gene



Genetic

Genome



Genomics

Genomic 基因组学 in Microbiology

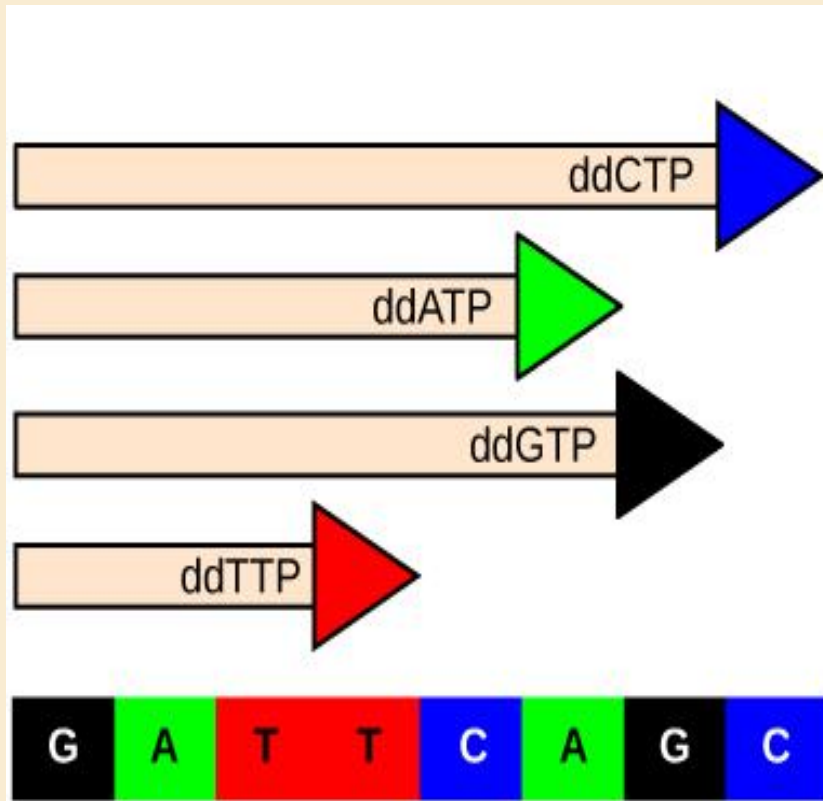
DNA sequencing methods:

Sanger DNA sequencing

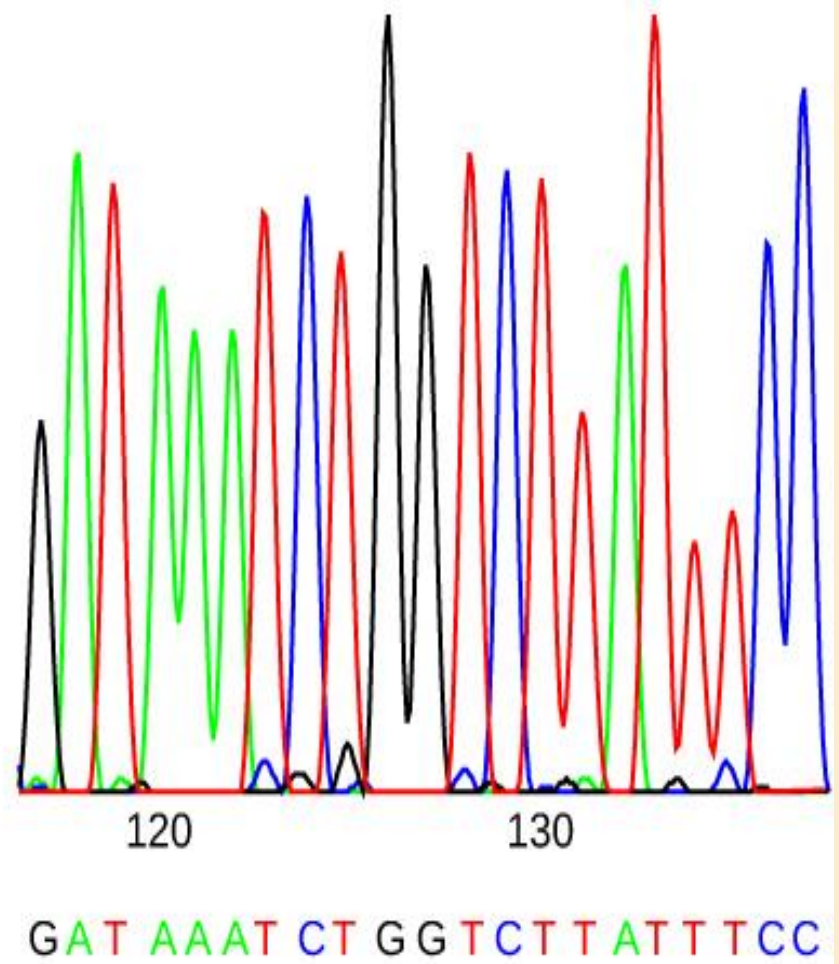
Genome DNA sequencing

- Sanger Method
 - Second generation and next-generation sequencing
-

Sanger Method for DNA sequencing



Dye-labeled dideoxynucleotides are used to generate DNA fragments of different lengths



The Nobel Prize in Chemistry 1980



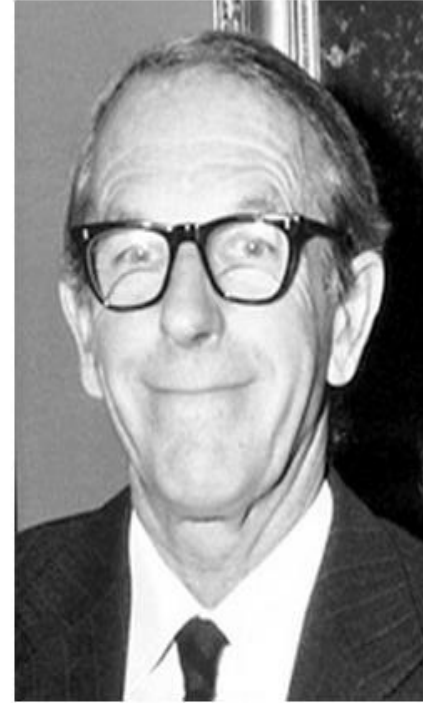
Paul Berg

Prize share:



Walter Gilbert

Prize share:



Frederick Sanger

Prize share:

"for their contributions concerning the determination of base sequences in nucleic acids"

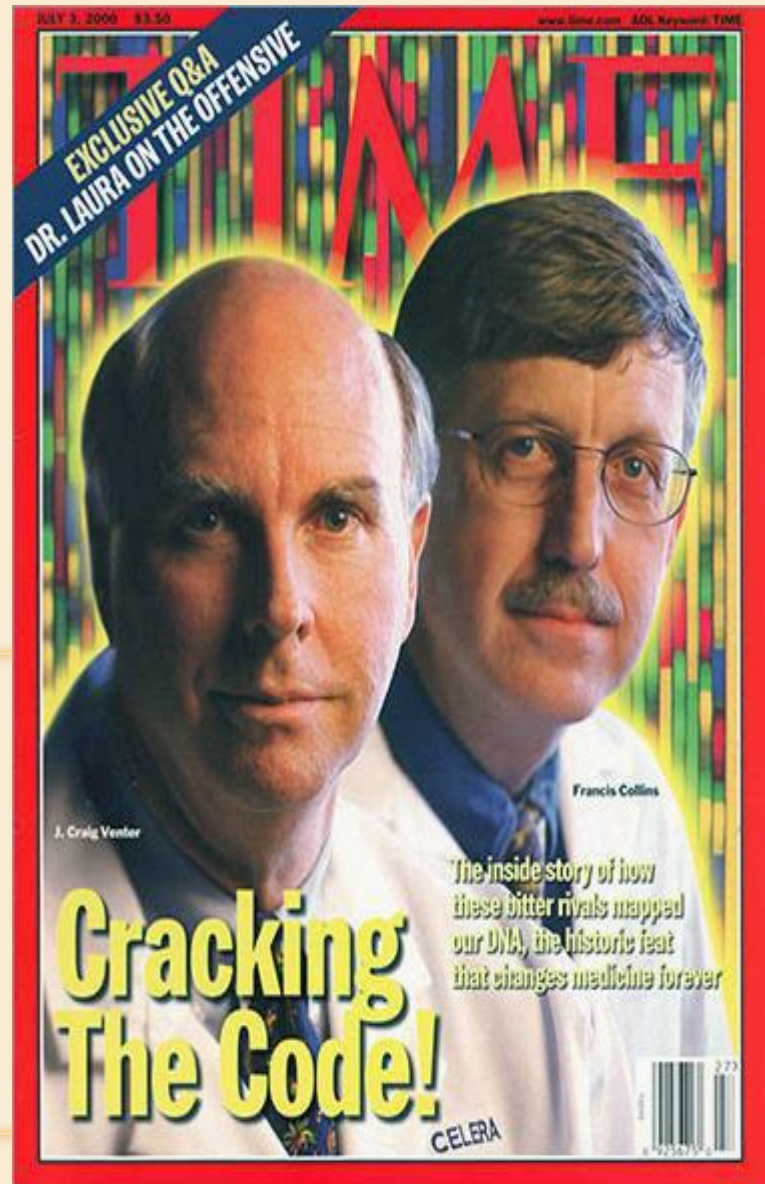


2000, Human Genome Project/HGP



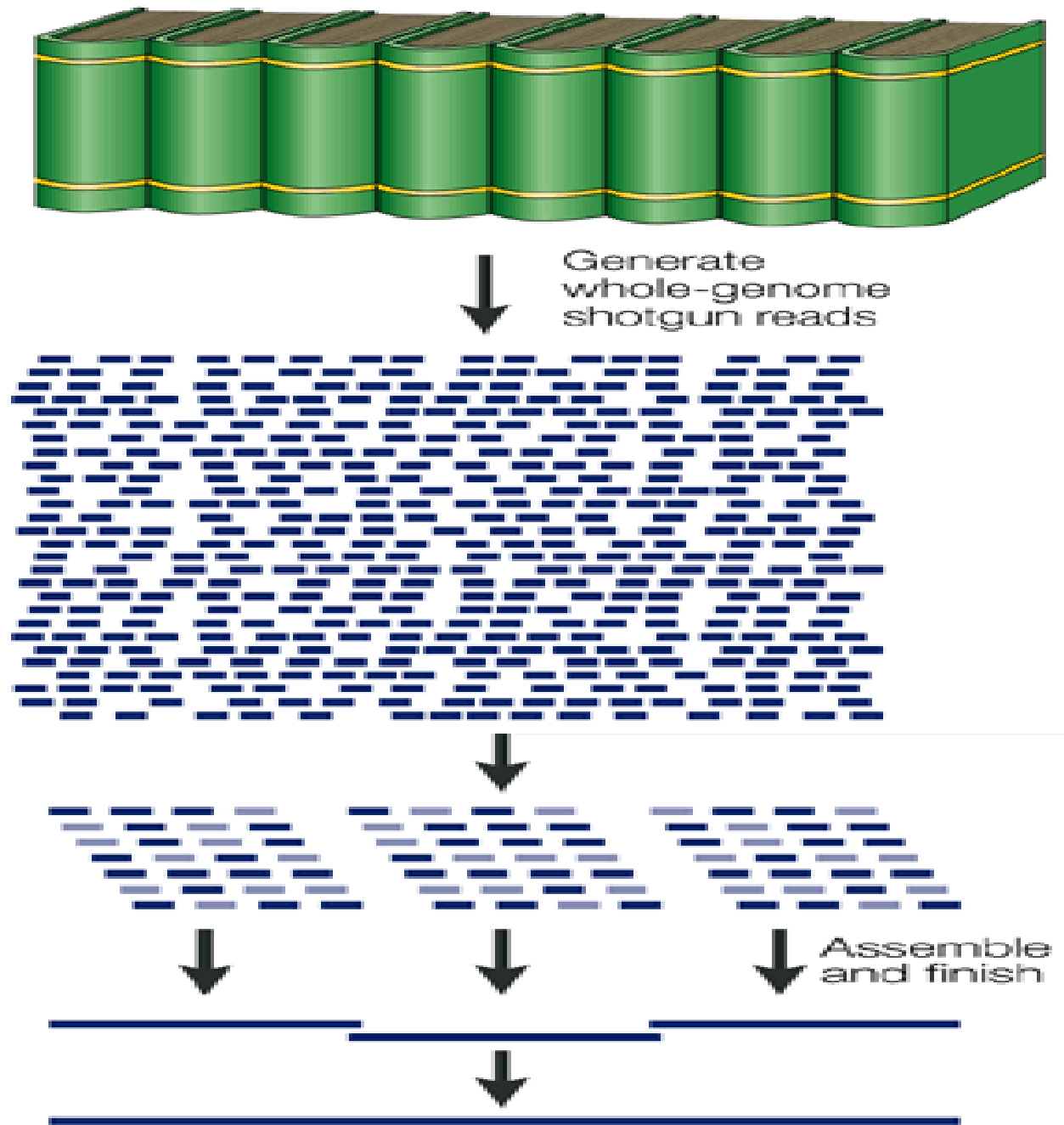
Craig Venter and Francis Collins

Craig Venter

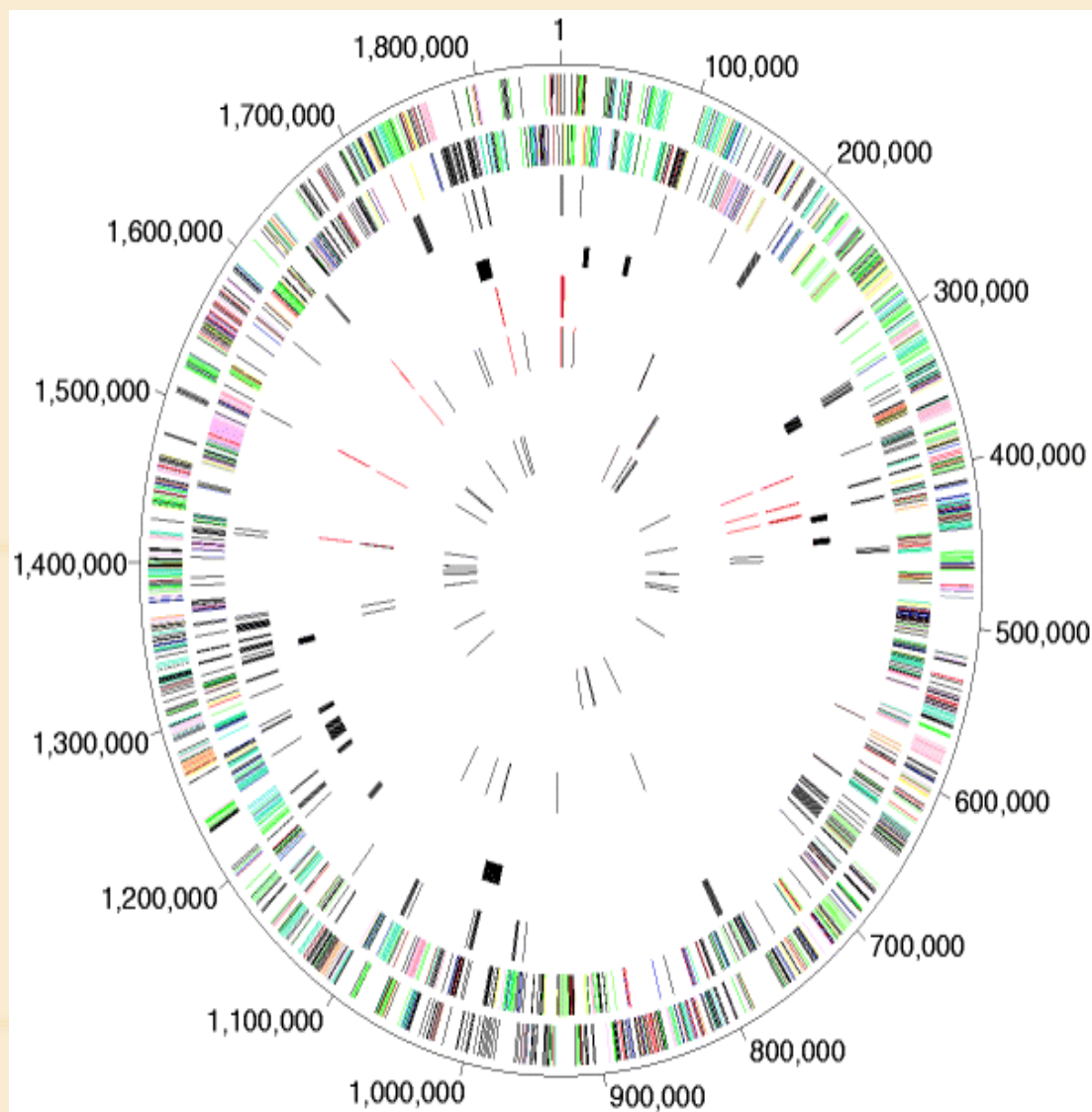


Francis Collins

Whole- genome shotgun sequencing

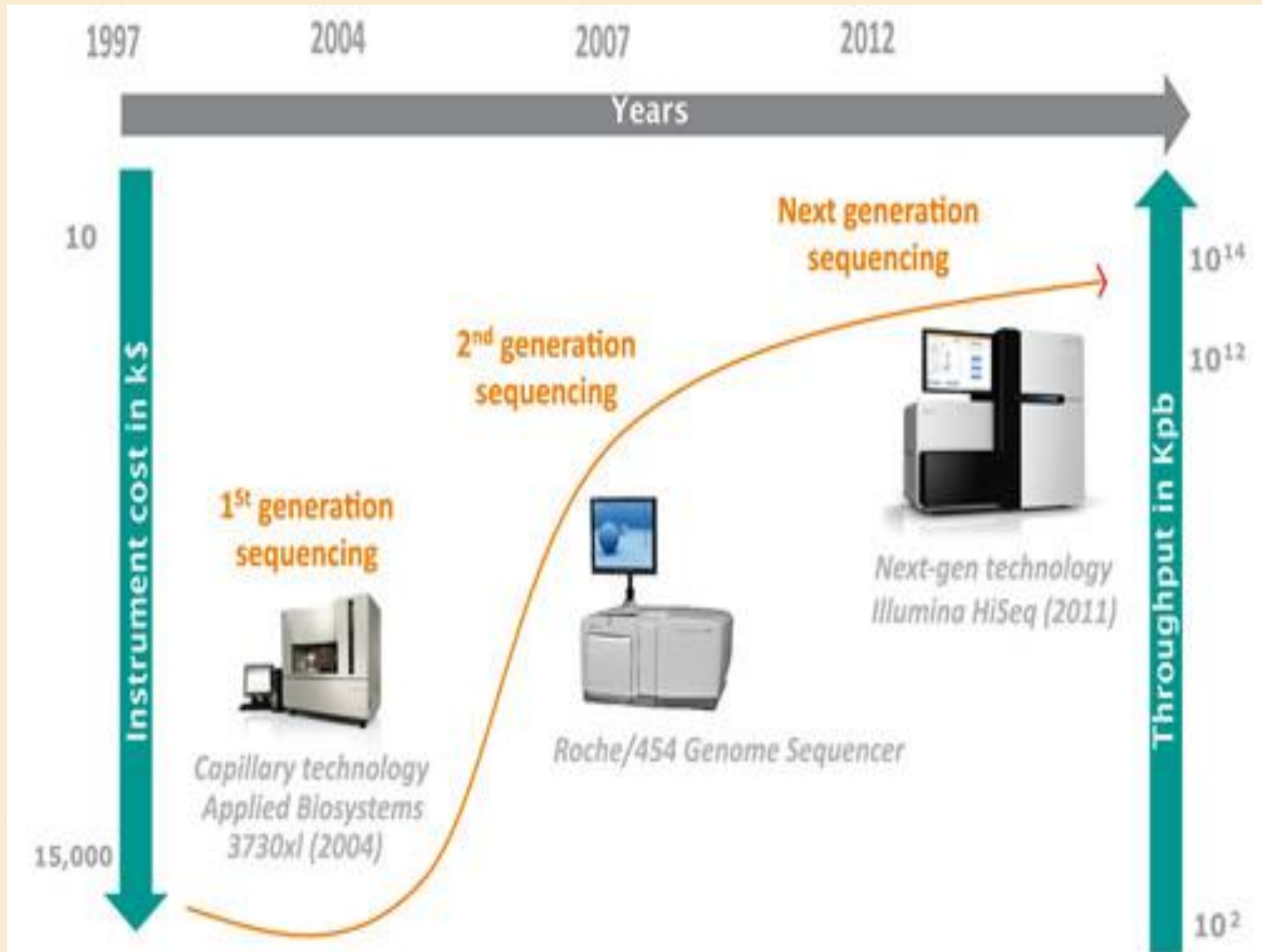


Haemophilus influenzae 流感嗜血杆菌 genome sequencing finished at 1995 using Whole-genome **shotgun sequencing**

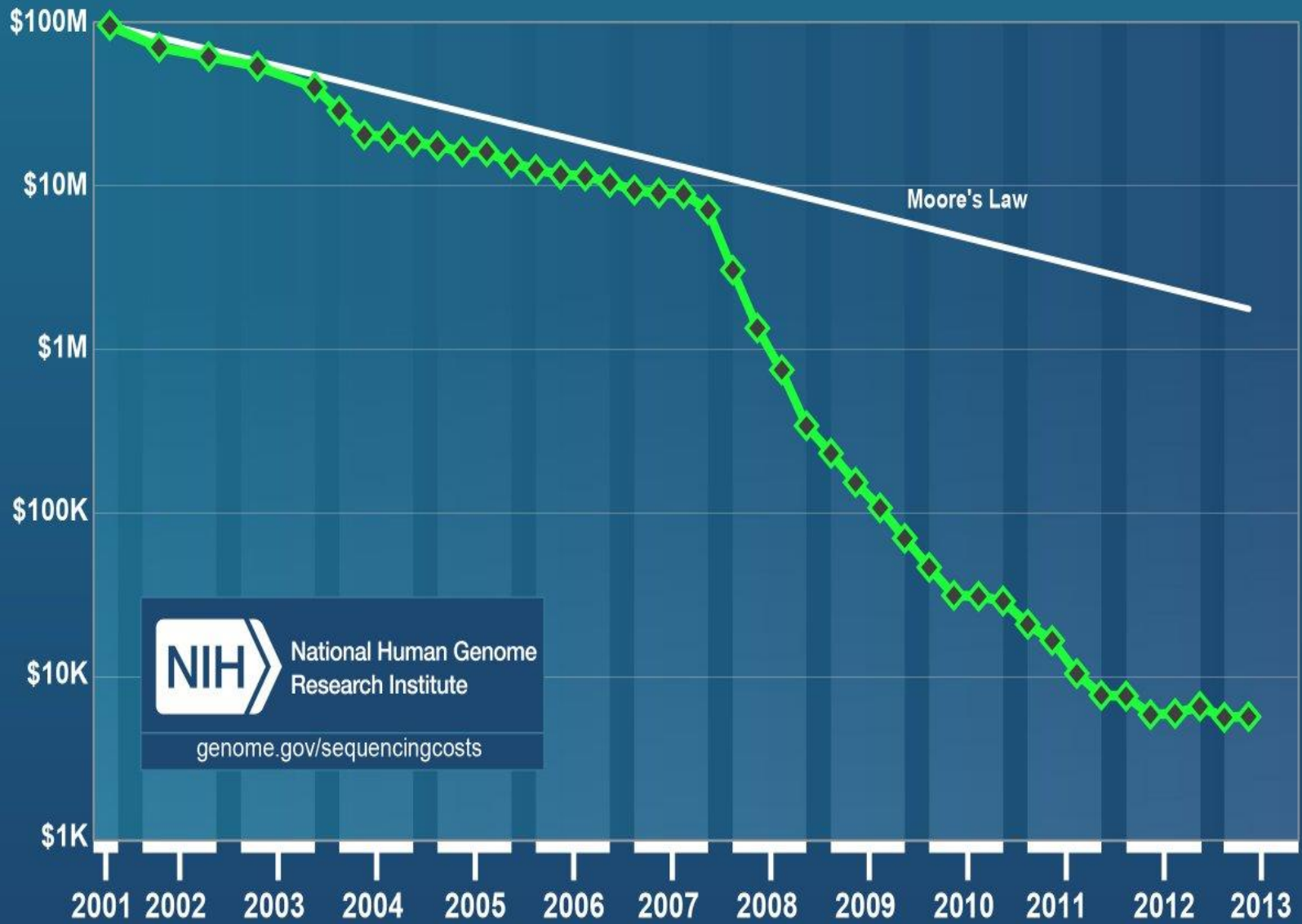


1.8 million bp, 1700 genes estimated

Evolution of sequencing instruments over time



Cost per Genome



NIH National Human Genome Research Institute
genome.gov/sequencingcosts

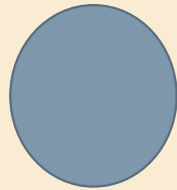
Metagenomics 宏基因组学



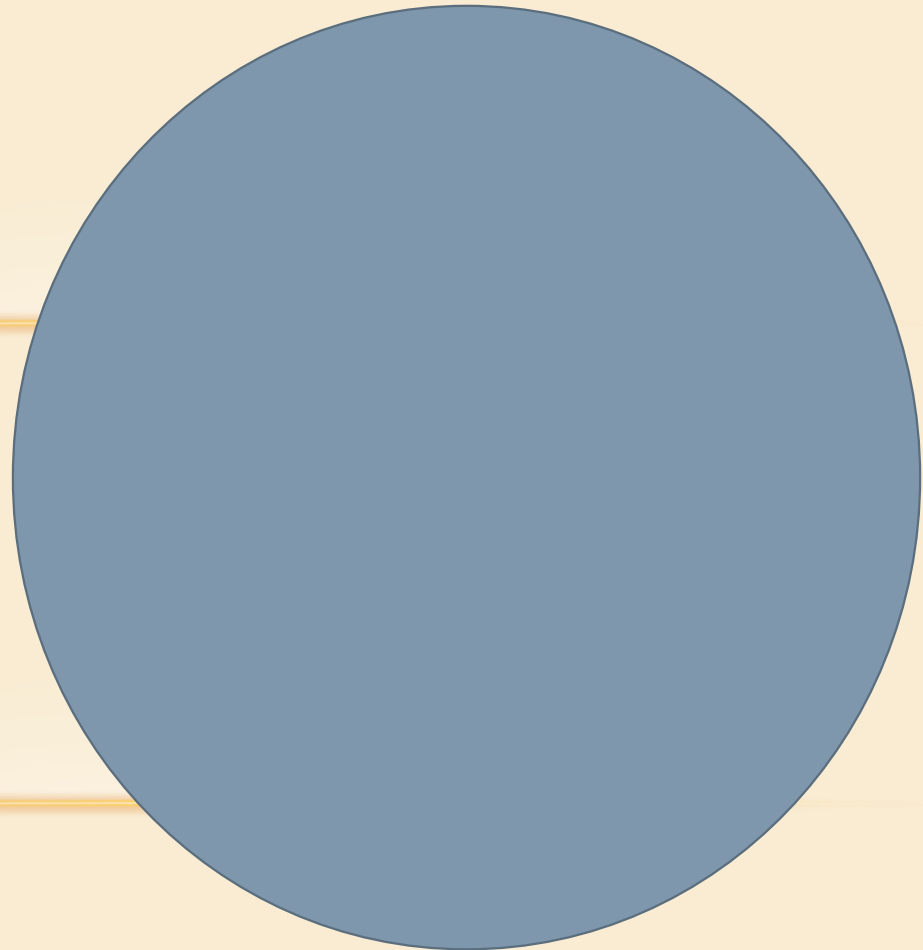
Metagenomics is the study of genetic material recovered directly from environmental samples

Why Metagenomics?

Microorganism that are not culturable



Microorganism that
are culturable

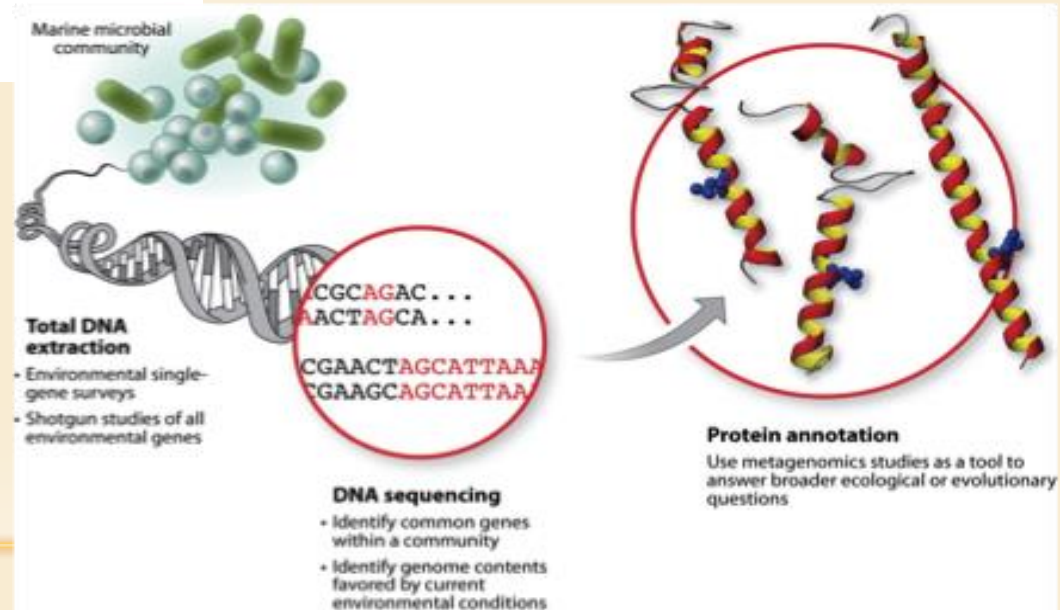
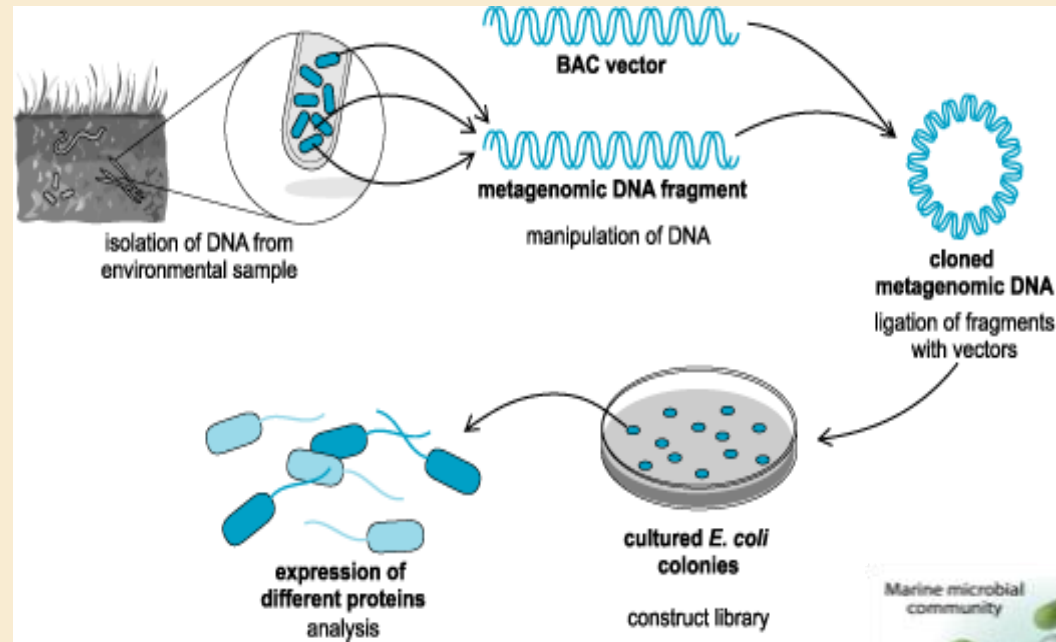


Metagenomics

- **High-throughput Sequencing**
 - **Bioinformatics/Data analysis**

 - **Applications**
 - **Biotechnology**
 - **Disease prevention and treatment**
 - **Agriculture**
 - **Environment**
-

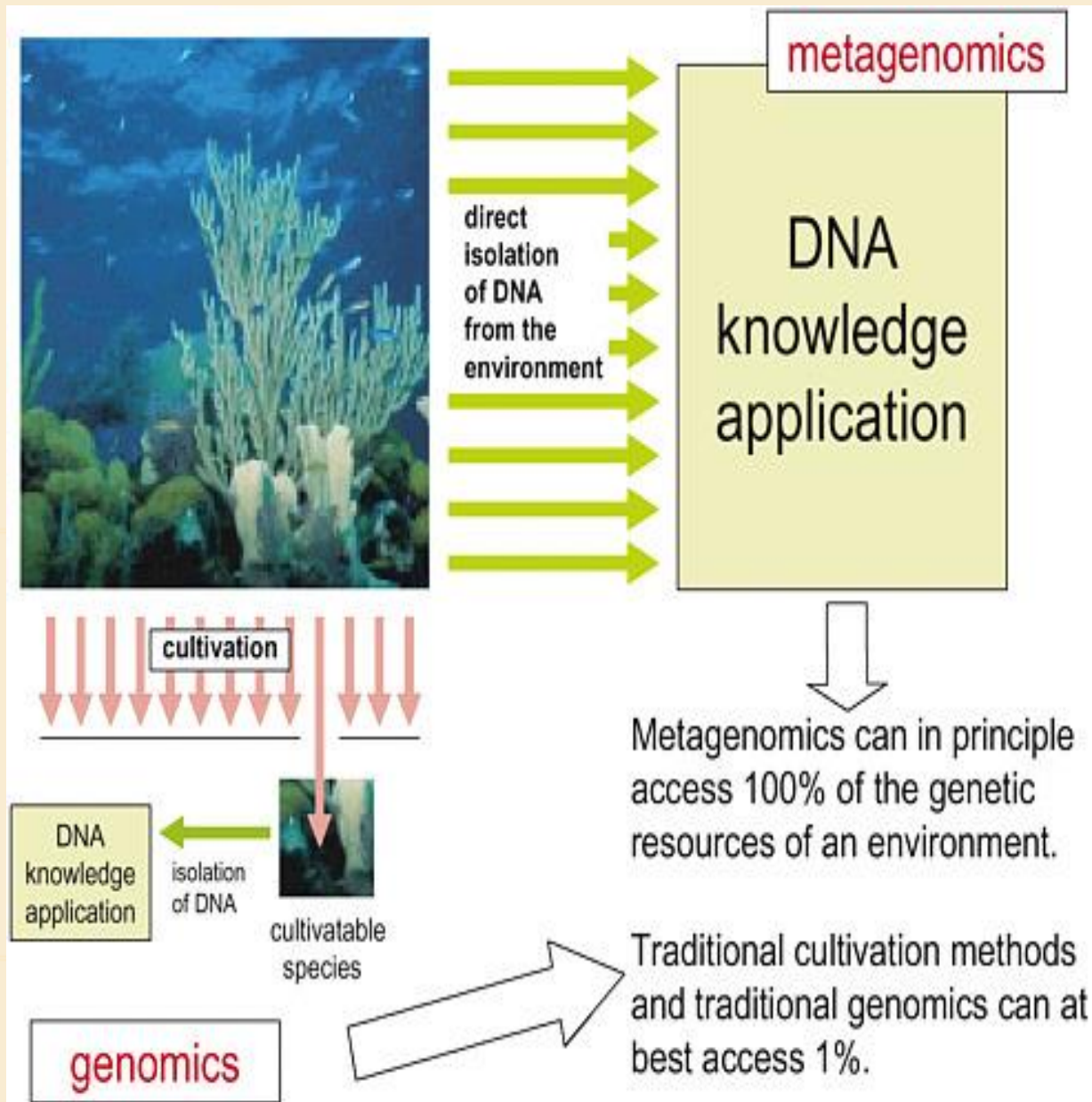
Discovery of Novel Protein from Non-culturable Microorganism



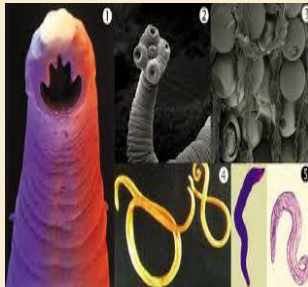
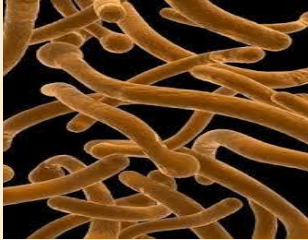
Gilbert JA, Dupont CL. 2011.

Annu. Rev. Mar. Sci. 3:347-71

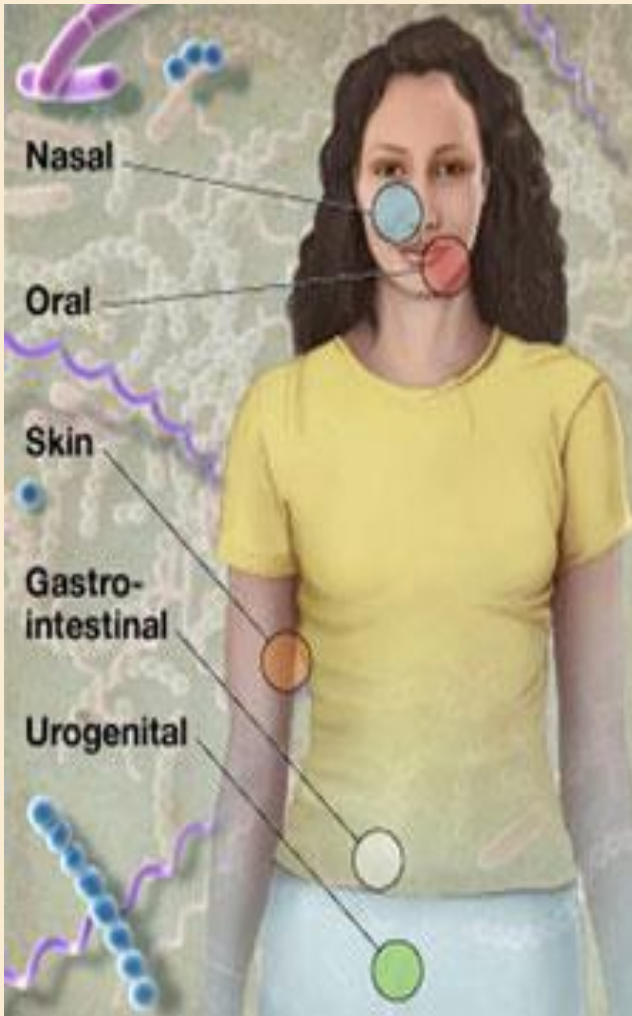
Treasury In The Deep Sea



Diagnosis of Pathogen infection by Genome Sequencing



Human Microbiome Project



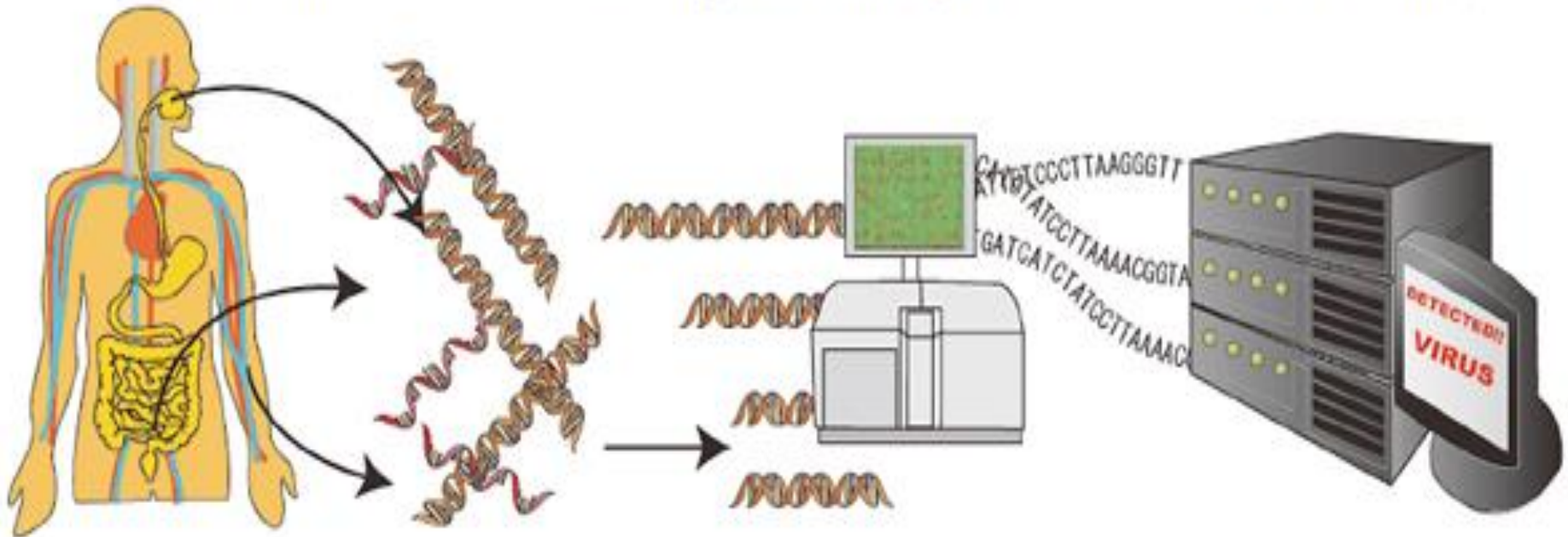
- **Development of a reference set of microbial genome sequences and preliminary characterization of the human microbiome**
- **Elucidation of the relationship between disease and changes in the human microbiome**

Metagenomics for Microorganism Resident in Human

Extract DNA or RNA

Next-generation sequencer

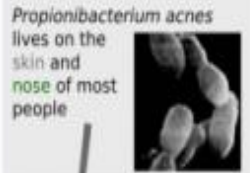
DATA analysis



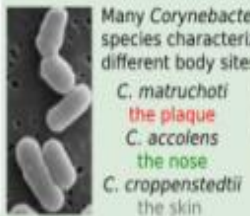
A map of diversity in the human microbiome



Streptococcus dominates the oral cavity with *S. mitis* > 75% in the **cheek**



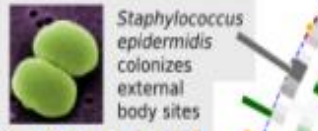
Propionibacterium acnes lives on the skin and nose of most people



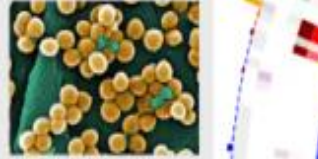
Many *Corynebacterium* species characterize different body sites:
C. matruchoti the **plaque**
C. accolens the **nose**
C. croppenstedtii the **skin**



Lactobacillus species (*L. gasseri*, *L. jensenii*, *L. crispatus*, *L. iners*) are predominant but mutually exclusive in the **vagina**



Staphylococcus epidermidis colonizes external body sites



○ Commensal microbes
 ☆ Potential pathogens

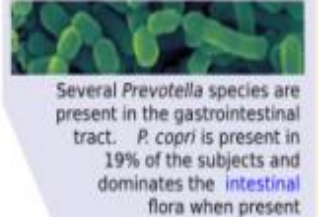
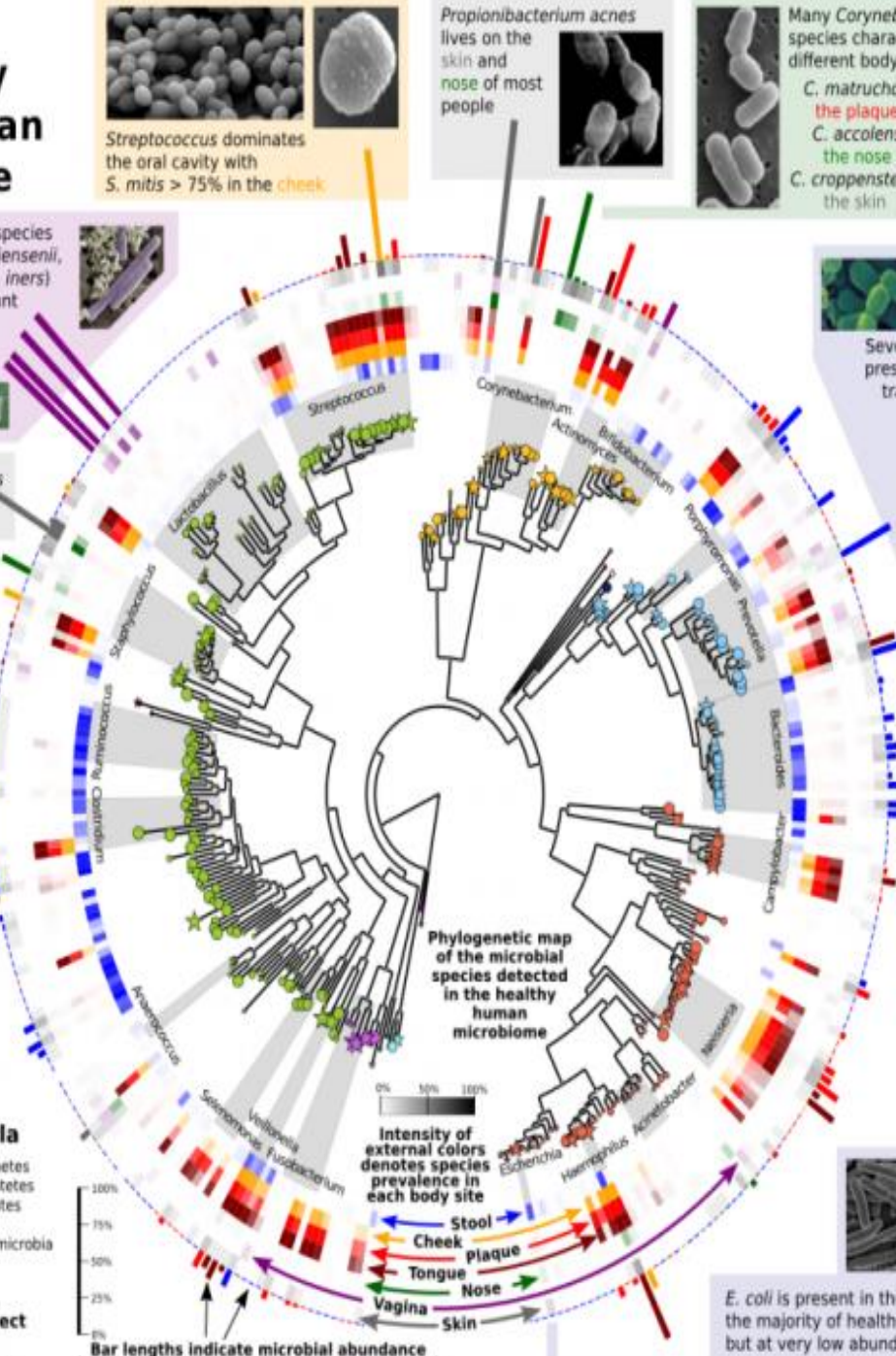
The four most abundant phyla

- Actinobacteria
- Bacteroidetes
- Firmicutes
- Proteobacteria

Low abundance phyla

- Chloroflexi
- Cyanobacteria
- Euryarchaeota
- Fusobacteria
- Lentisphaerae
- Spirochaetes
- Synergistetes
- Tenericutes
- Thermi
- Verrucomicrobia

National Institutes of Health Human Microbiome Project



Several *Prevotella* species are present in the gastrointestinal tract. *P. copri* is present in 19% of the subjects and dominates the **intestinal** flora when present



Bacteroides is the most abundant genus in the **gut** of almost all healthy subjects



Campylobacter includes opportunistic pathogens, but members live in the oral cavities of most healthy people in the cohort



E. coli is present in the **gut** of the majority of healthy subjects but at very low abundance.



Bar lengths indicate microbial abundance (colored by body site of greatest prevalence)

放线杆菌 actinobacteria
 bacteroidetes 拟杆菌
 厚壁菌 firmicute
 Proteobacteria 变形菌门

A healthy adult human harbors:

- about 100 trillion microbes outnumbering our own cells by a factor of ten;
- expanding our own gene repertoire by at least two orders of magnitude.

The whole world is reading
pirated papers pp. 407 & 308

Neurochemistry of sleeping
and waking pp. 517 & 550

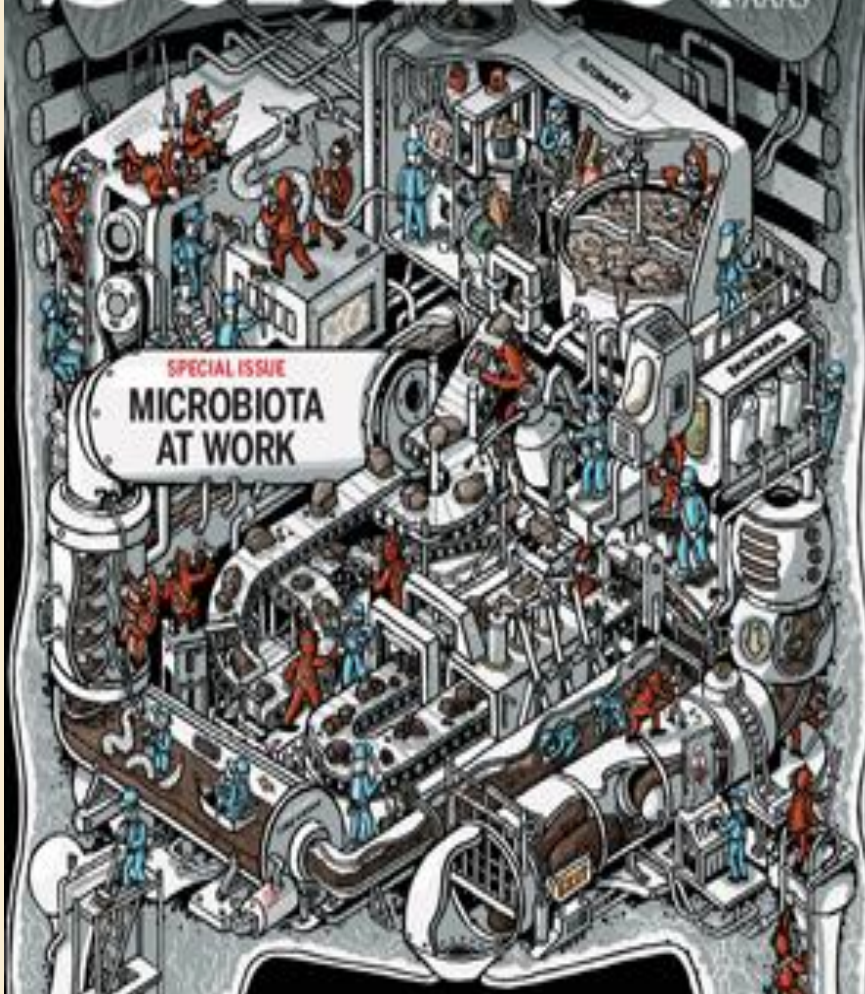
Halogenated olefins
via the E train p. 509

Science

115
29 APRIL 2016
science.sciencemag.org

AAAS

SPECIAL ISSUE
MICROBIOTA
AT WORK

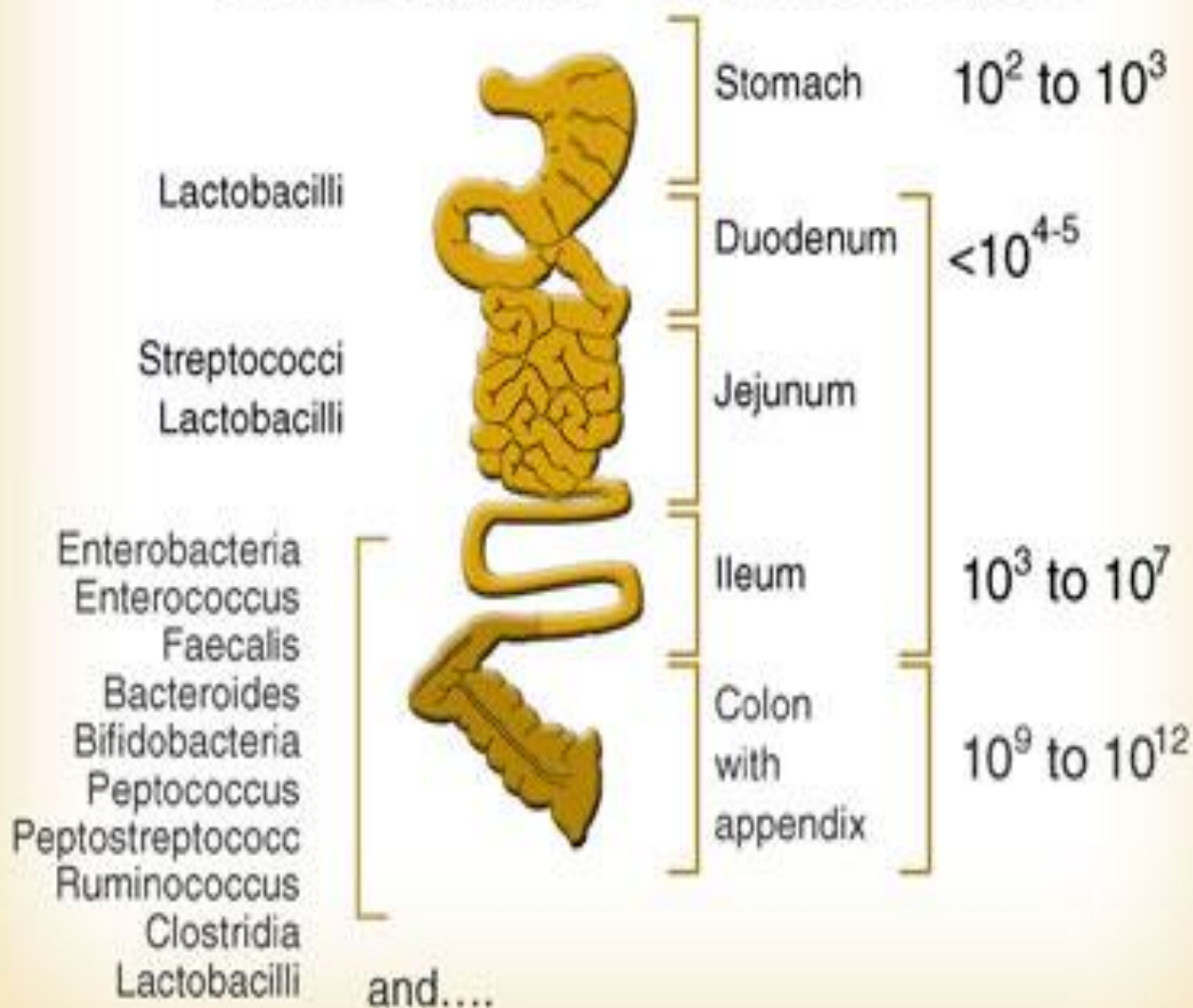


Science AAAS

- Population-level analysis of gut microbiome variation , Science. 2016
- Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity , Science. 2016

INTESTINAL MICROFLORA

10^{14} micro-organisms, >500 differentes species

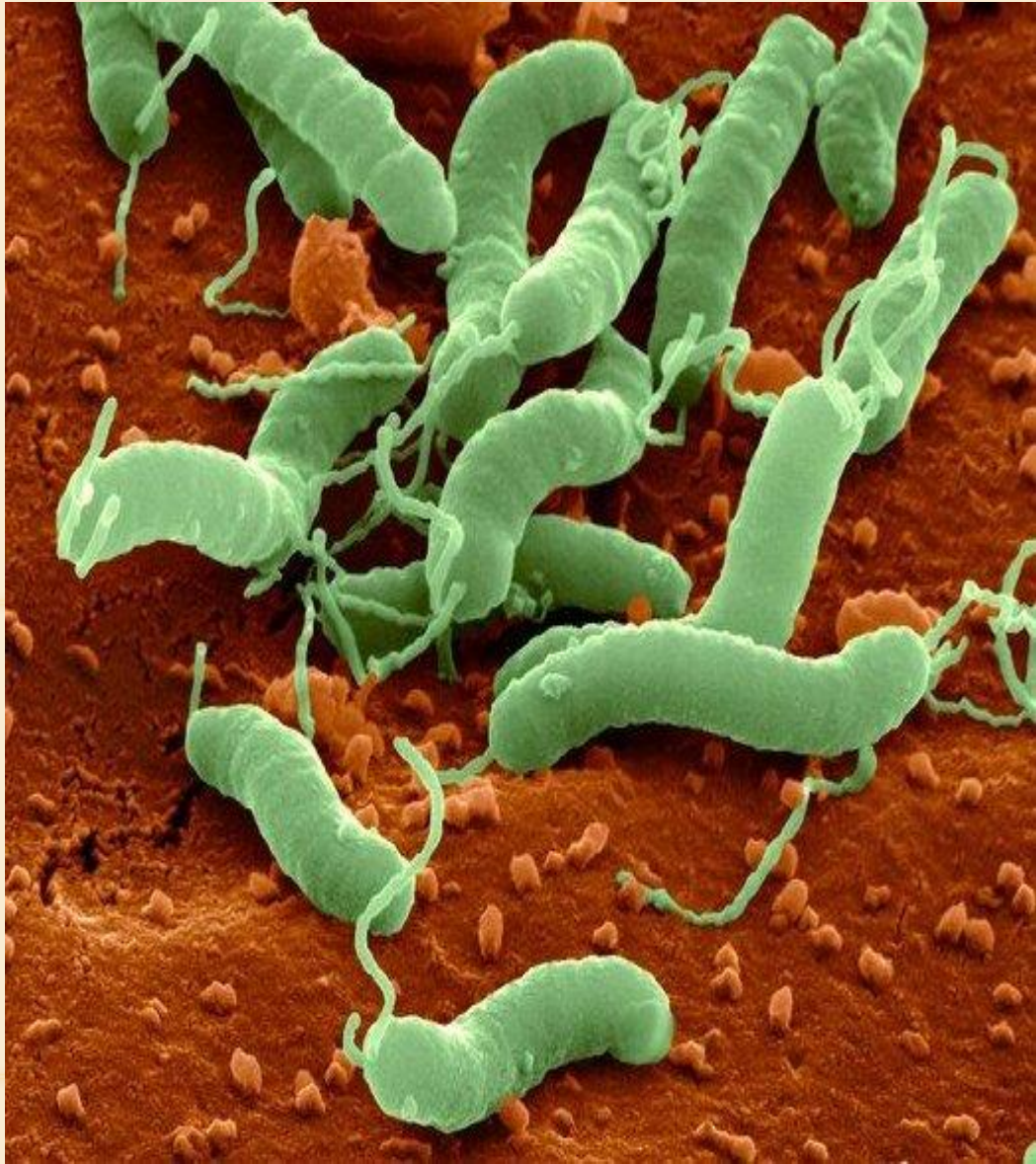




The Nobel Prize in Physiology or Medicine 2005

Barry J. Marshall, J. Robin Warren





Flagella

bacterial mobility & chemotaxis
to colonize under mucosa

Urease

neutralize gastric acid
gastric mucosal injury (by ammonia)

Lipopolysaccharides

adhere to host cells
inflammation

Outer proteins

adhere to host cells

Exotoxin(s)

- **vacuolating toxin (vacA)**
gastric mucosal injury

Secretory enzymes

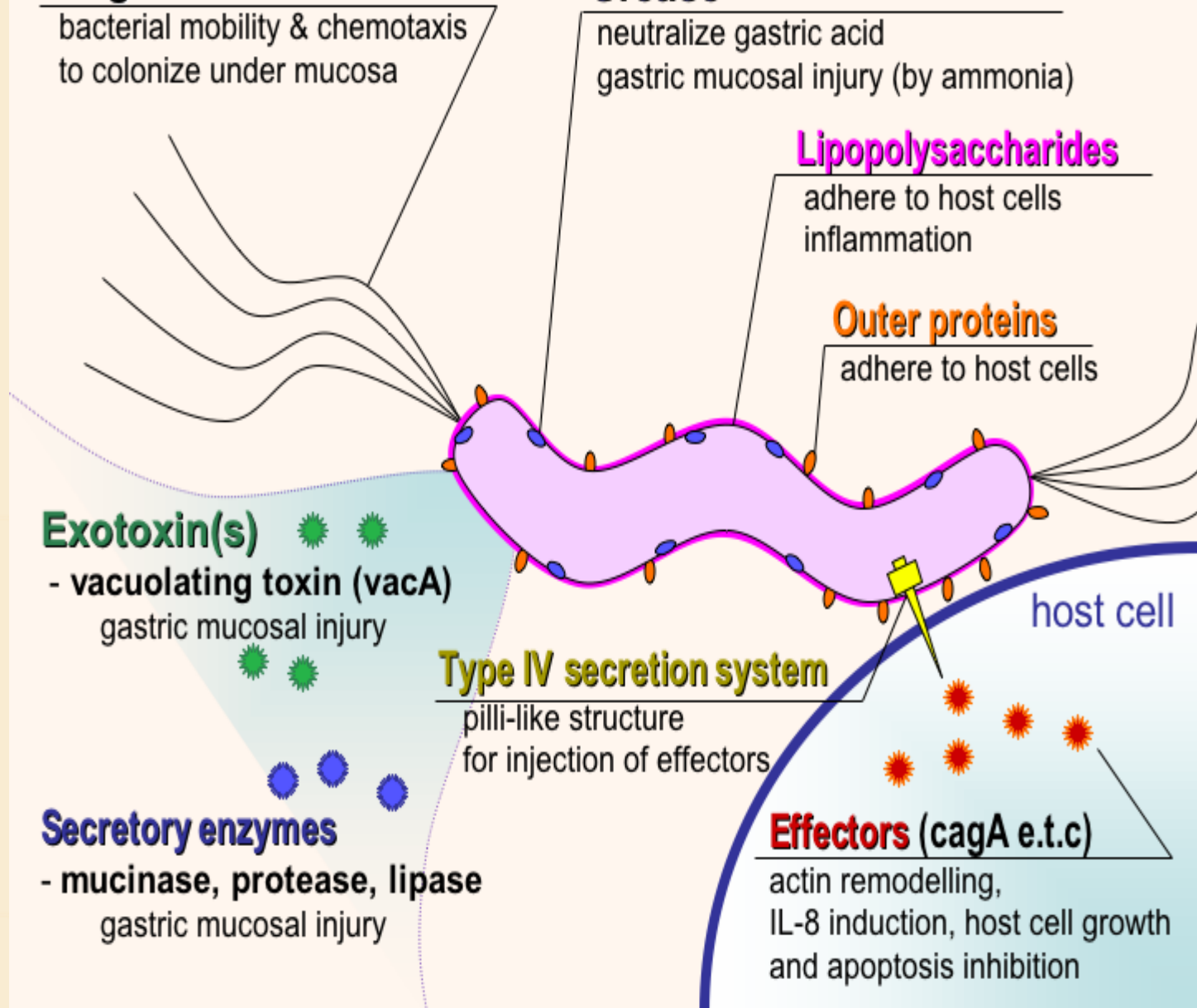
- **mucinase, protease, lipase**
gastric mucosal injury

Type IV secretion system

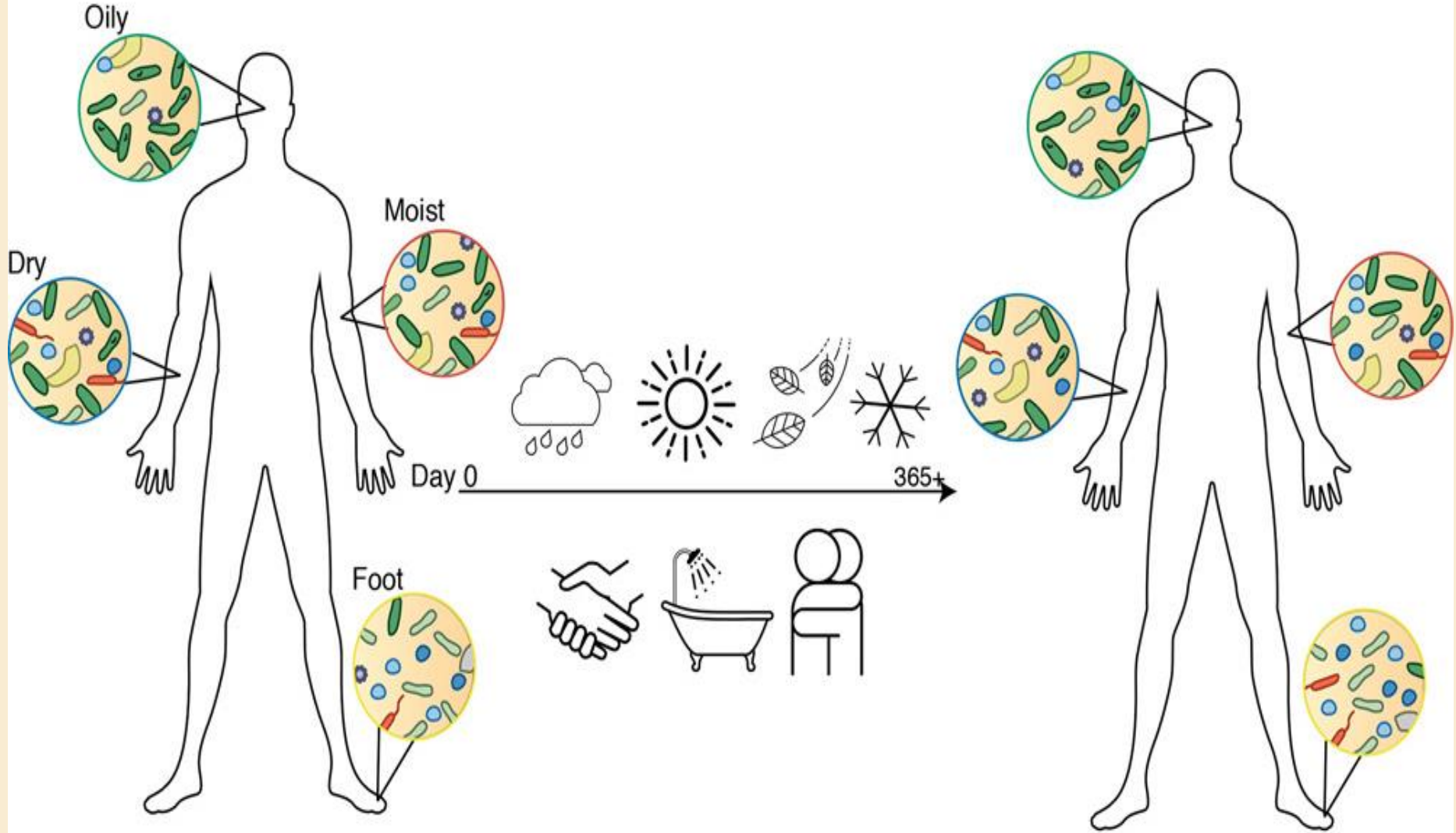
pilli-like structure
for injection of effectors

Effectors (cagA e.t.c)

actin remodelling,
IL-8 induction, host cell growth
and apoptosis inhibition



Temporal Stability of the Human Skin Microbiome



 Bacteria
  Fungi
  Virus
  Phage

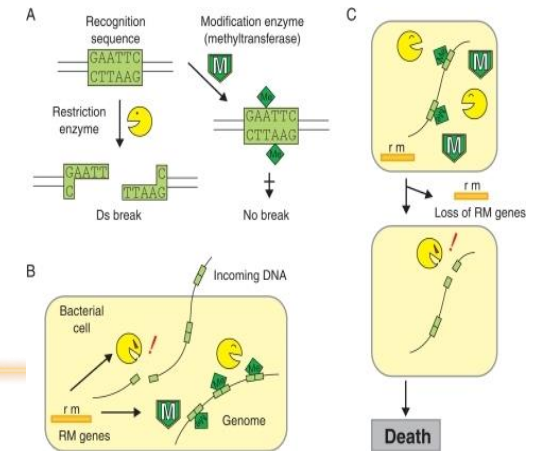
Discussion:

水土不服

夫妻相

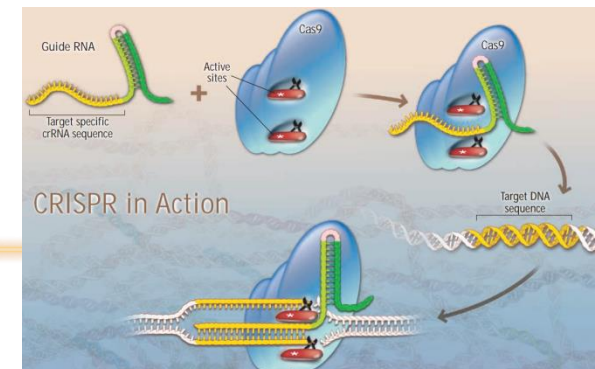
Evolve to distinguish cell own DNA and any foreign DNA entering the cell, and destroy the later.

- **Restriction–Modification Systems in archaea and bacterial**



- **CRISPR/Cas9 system in archaea and bacterial**

bacterial immune system

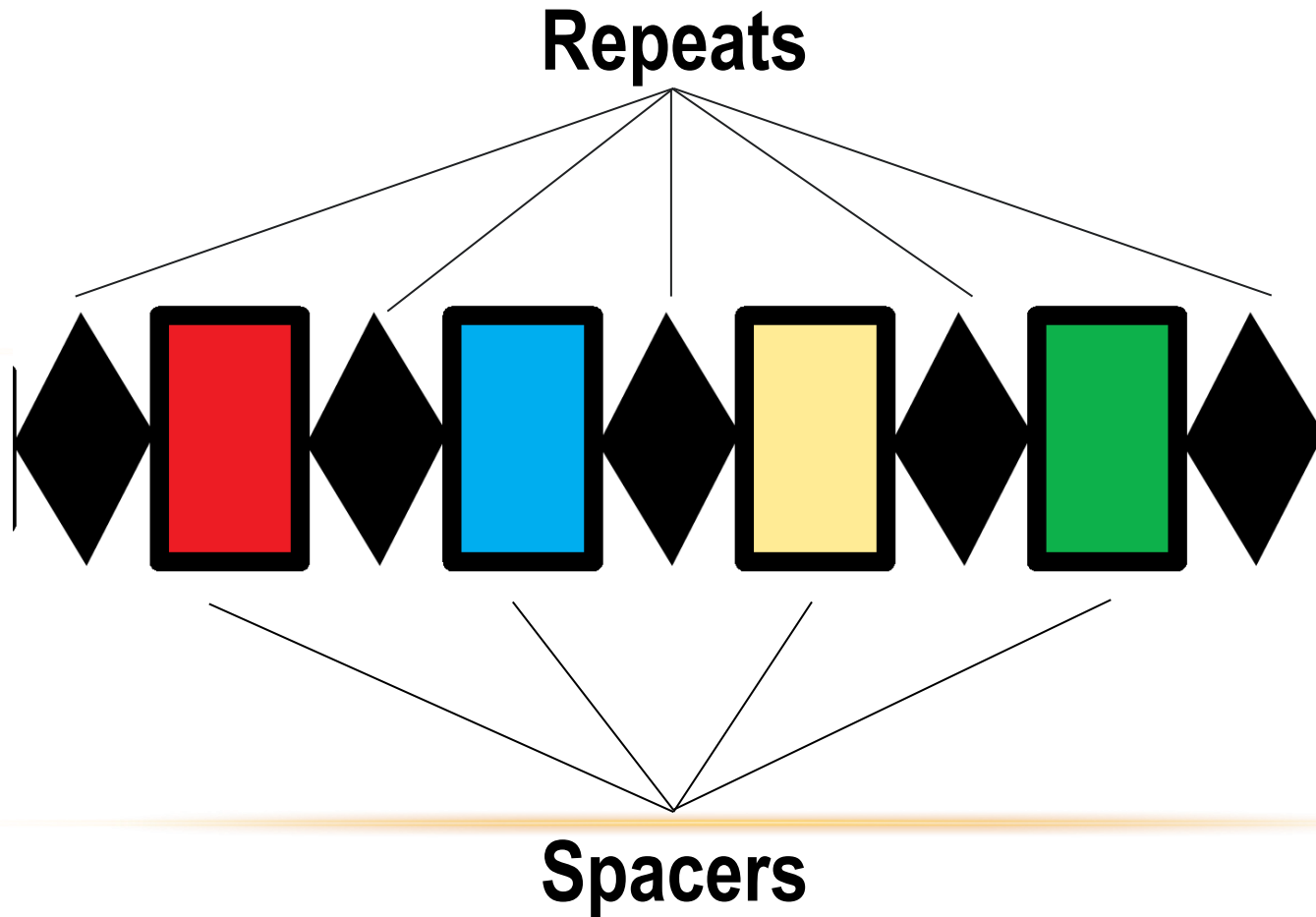


The Biology of CRISPR/Cas9

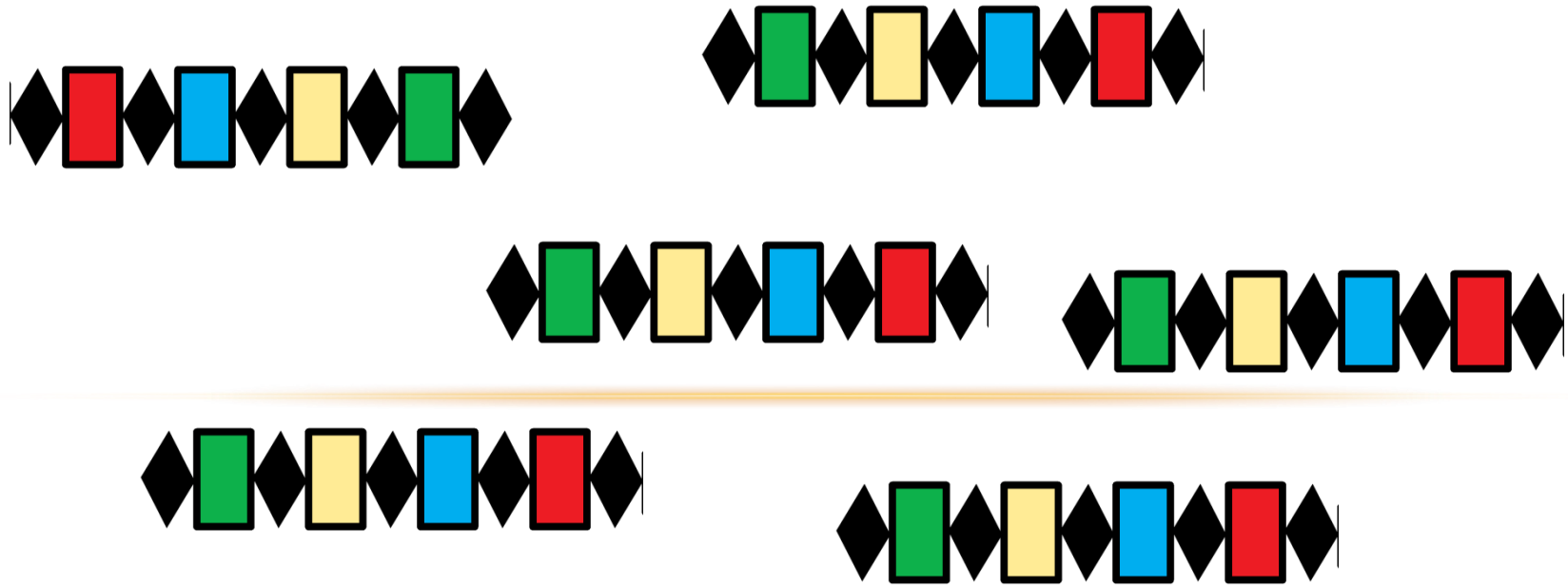
- CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats)
规律成簇间隔短回文重复
- CRISPR-associated (Cas) genes
- essential in adaptive immunity in select bacteria and archaea, enabling the organisms to respond to and eliminate invading genetic material.

An unusual structure was found in the 3'-end flanking region of *iap* (Fig. 5). Five highly homologous sequences of 29 nucleotides were arranged as direct repeats with 32 nucleotides as spacing. The first sequence was included in the putative transcriptional termination site and had less homology than the others. Well-conserved nucleotide sequences containing a dyad symmetry, named REP sequences, have been found in *E. coli* and *Salmonella typhimurium* (28) and may act to stabilize mRNA (18). A dyad symmetry with 14 nucleotide pairs was also found in the middle of these sequences (underlining, Fig. 5), but no homology was found between these sequences and the REP sequence. So far, no sequence homologous to these has been found elsewhere in procaryotes, and the biological significance of these sequences is not known.

Clustered **R**egularly **I**nterspaced **S**hort **P**alindromic **R**epeat sequences,
成簇的、规律间隔的短回文重复序列,即**CRISPR**序列



CRISPR: widespread occurrence in bacteria and archaea genomes




1. Jansen R, Embden JD, Gaastra W, Schouls LM. **Identification of genes that are associated with DNA repeats in prokaryotes.** *Mol Microbiol.* 2002; **43**:1565-75.

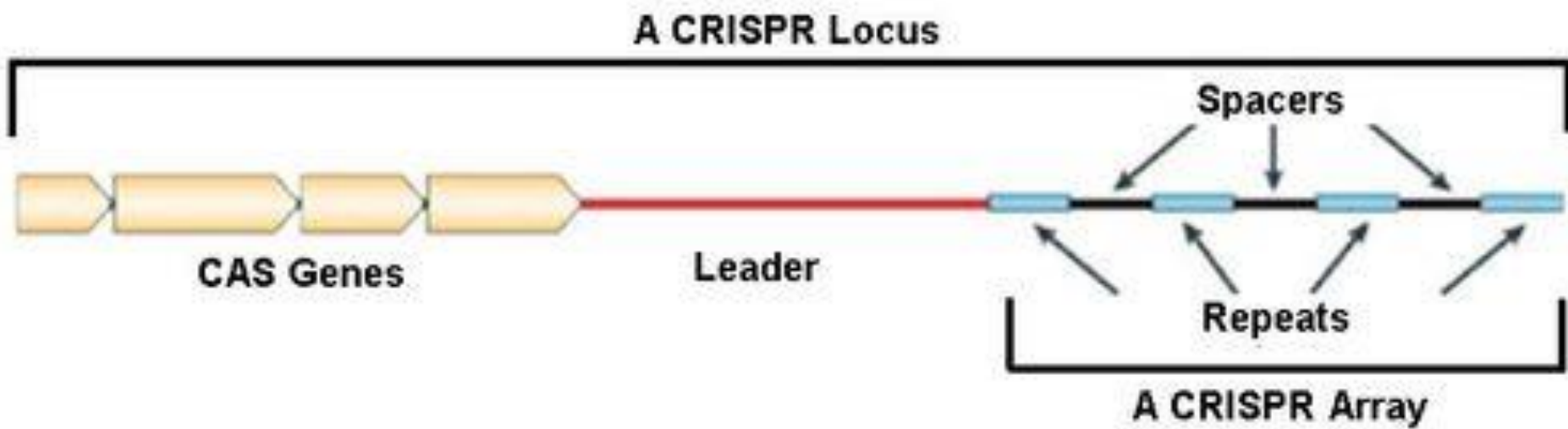
2. Mojica FJ, Diez-Villasenor C, Soria E, Juez G. **Biological significance of a family of regularly spaced repeats in the genomes of Archaea. Bacteria and mitochondria.** *Mol Microbiol.* 2000; **36**:244-6.



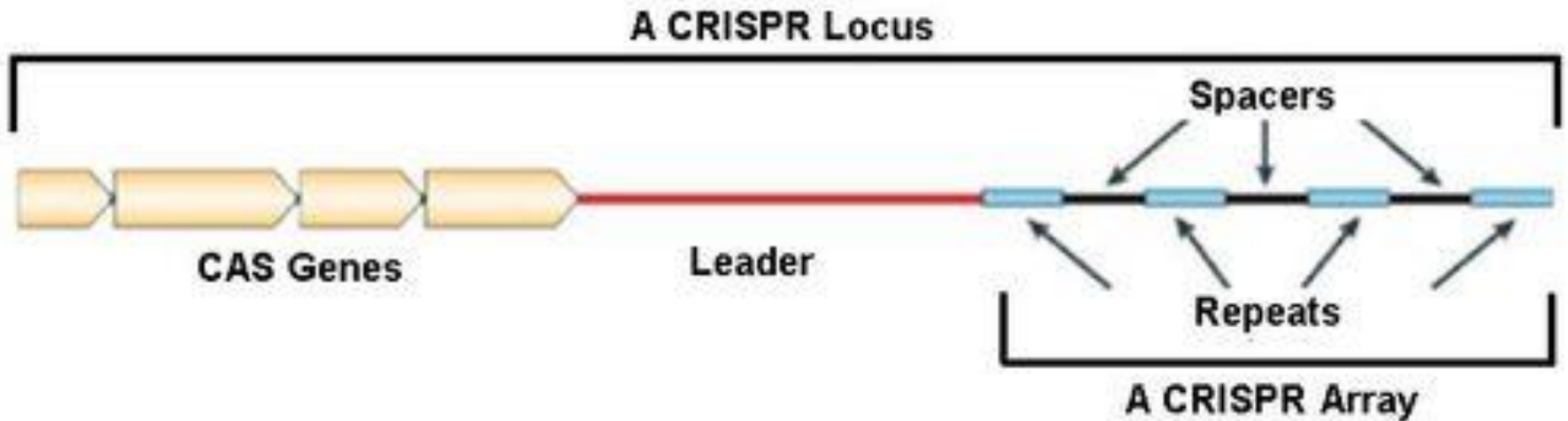
Identification of genes that are associated with DNA repeats in prokaryotes

Ruud. Jansen , Jan. D. A. van Embden, Wim. Gaastra,
Leo. M. Schouls

First published: March 2002 [Full publication history](#)



CRISPR: biological function?



The first functional clue emerged until 2005, with the observation that **CRISPR spacers showed homology (序列同源) to viral sequences.**

1. Pourcel C, Salvignol G, Vergnaud G. **CRISPR elements in *Yersinia pestis* acquire new repeats by preferential uptake of bacteriophage DNA, and provide additional tools for evolutionary studies.** *Microbiology*. 2005; **151**:653-63.

耶尔森氏鼠疫杆菌

2. Mojica FJ, Diez-Villasenor C, Garcia-Martinez J, Soria E. **Intervening sequences of regularly spaced prokaryotic repeats derive from foreign genetic elements.** *J Mol Evol*. 2005; **60**:174-82.

3. Bolotin A, Quinquis B, Sorokin A, Ehrlich SD. **Clustered regularly interspaced short palindrome repeats (CRISPRs) have spacers of extrachromosomal origin.** *Microbiology*. 2005; **151**:2551-61.

耶尔森氏鼠疫杆菌

CRISPR elements in *Yersinia pestis* acquire new repeats by preferential uptake of bacteriophage DNA, and provide additional tools for evolutionary studies

C. Pourcel,¹ G. Salvignol¹ and G. Vergnaud^{1,2}

¹GPMS, Institut de Génétique et Microbiologie, Université Paris XI, 91405 Orsay cedex, France

²Centre d'Etudes du Bouchet, 5 rue Lavoisier, 91710 Vert le Petit, France

Correspondence

G. Vergnaud

Gilles.Vergnaud@igmors.u-psud.fr

2. Intervening sequences of regularly spaced prokaryotic repeats derive from foreign genetic elements. *J Mol Evol.* 2005; **60**:174-82.

Table 2. Distribution of CRISPR-spacer homologs

序列同源

Strain	No. of spacers analyzed	No. of spacers with homologs in	
		Phages ^a	Plasmids
<i>Chlorobium tepidum</i> TLS	62		1
<i>Clostridium tetani</i> Massachusetts E88	62	1	
<i>Corynebacterium efficiens</i> YS-314T	22		1
<i>Escherichia coli</i> ECOR42	14		1
<i>Escherichia coli</i> ECOR44	10	1	
<i>Escherichia coli</i> ECOR47	17	1	
<i>Escherichia coli</i> ECOR49	11		1
<i>Listeria innocua</i> Clip11262	9	3	
<i>Listeria monocytogenes</i> EGD-e	4	1	
<i>Methanothermobacter thermoautotrophicum</i> ΔH	169	9	
<i>Mycoplasma gallisepticum</i> R	71		
<i>Neisseria meningitidis</i> Z2491 (serogroup A)	16		
<i>Photorhabdus luminescens laumondii</i> TT01	65	7	
<i>Porphyromonas gingivalis</i> W83	44		
<i>Pyrobaculum aerophilum</i> IM2	129		
<i>Salmonella typhimurium</i> LT2 SGSC1412	57	1	
<i>Shigella sonnei</i> 53G	3		
<i>Streptococcus agalactiae</i> NEM316	13	1	
<i>Streptococcus agalactiae</i> 2603V/R	25	1	1
<i>Streptococcus pyogenes</i> MI GAS SF370 ←	9	8	
<i>Sulfolobus solfataricus</i> P2	424	6	3
<i>Sulfolobus tokodaii</i> 7	471	2	2
<i>Thermoanaerobacter tengcongensis</i> MB4T	306		
<i>Yersinia pestis</i> CO-92 (Biovar Orientalis)	16	4	
<i>Yersinia pestis</i> KIM5P12 (Biovar Mediaevalis)	10	1	

CRISPR: biological function

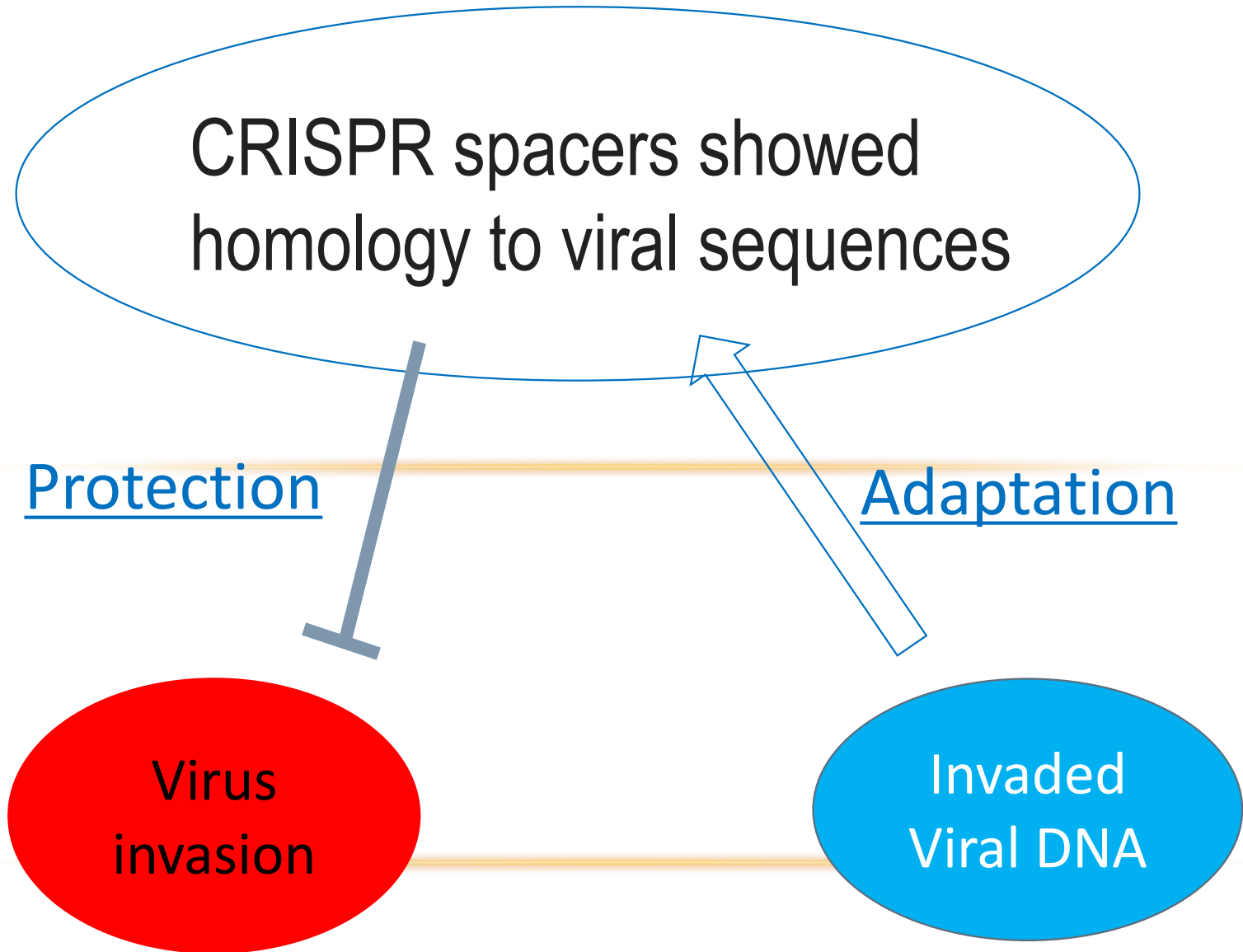
CRISPR spacers showed
homology to viral sequences

Protection

Adaptation

Virus
invasion

Invaded
Viral DNA



Milestone Discovery

www.sciencemag.org

SCIENCE

VOL 315

23 MARCH 2007

CRISPR Provides Acquired Resistance Against Viruses in Prokaryotes

Rodolphe Barrangou,¹ Christophe Fremaux,² H el ene Deveau,³ Melissa Richards,¹ Patrick Boyaval,² Sylvain Moineau,³ Dennis A. Romero,¹ Philippe Horvath^{2*}

Clustered regularly interspaced short palindromic repeats (CRISPR) are a distinctive feature of the genomes of most Bacteria and Archaea and are thought to be involved in resistance to bacteriophages. We found that, after viral challenge, bacteria integrated new spacers derived from phage genomic sequences. Removal or addition of particular spacers modified the phage-resistance phenotype of the cell. Thus, CRISPR, together with associated *cas* genes, provided resistance against phages, and resistance specificity is determined by spacer-phage sequence similarity.

嗜热链球菌-*S. thermophilus*

Abstract

.....

We found that, after viral challenge, bacteria integrated new spacers derived from phage genomic sequences.

Removal or addition of particular spacers modified the phage-resistance phenotype of the cell.

Thus, CRISPR, together with associated cas genes, provided resistance against phages, and resistance specificity is determined by spacer-phage sequence similarity.

CRISPR: biological function

CRISPR spacers showed
homology to viral sequences

Protection

Adaptation

Virus
invasion

Invaded
Viral DNA

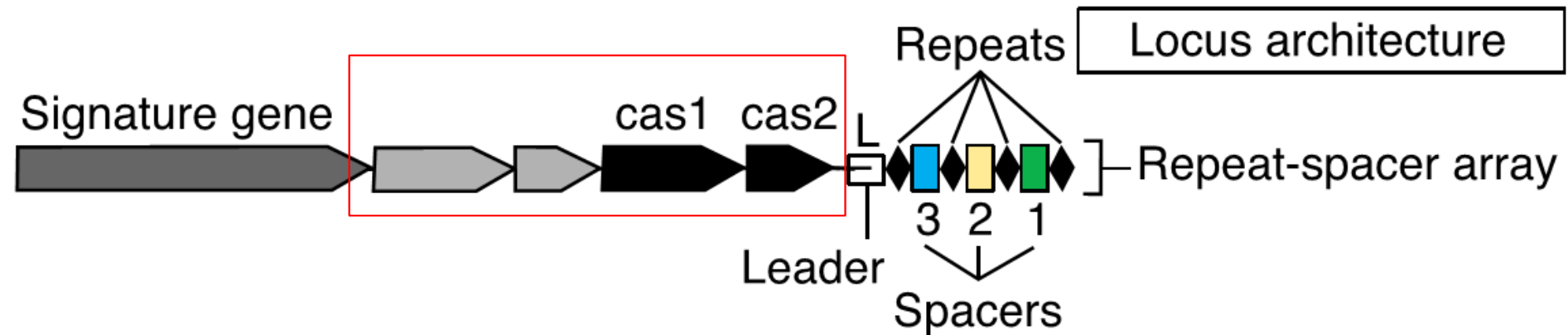
CRISPR: Molecular mechanism

1. Brouns SJ, Jore MM, Lundgren M, Westra ER, Slijkhuis RJ, Snijders AP et al.. **Small CRISPR RNAs guide antiviral defense in prokaryotes.** *Science*. 2008; **321**:960-4.

2. Marraffini LA, Sontheimer EJ. **CRISPR interference limits horizontal gene transfer in staphylococci by targeting DNA.** *Science*. 2008; **322**:1843-5.葡萄状球菌

3. Hale CR, Zhao P, Olson S, Duff MO, Graveley BR, Wells L et al.. **RNA-guided RNA cleavage by a CRISPR RNA-Cas protein complex.** *Cell*. 2009; **139**:945-56.

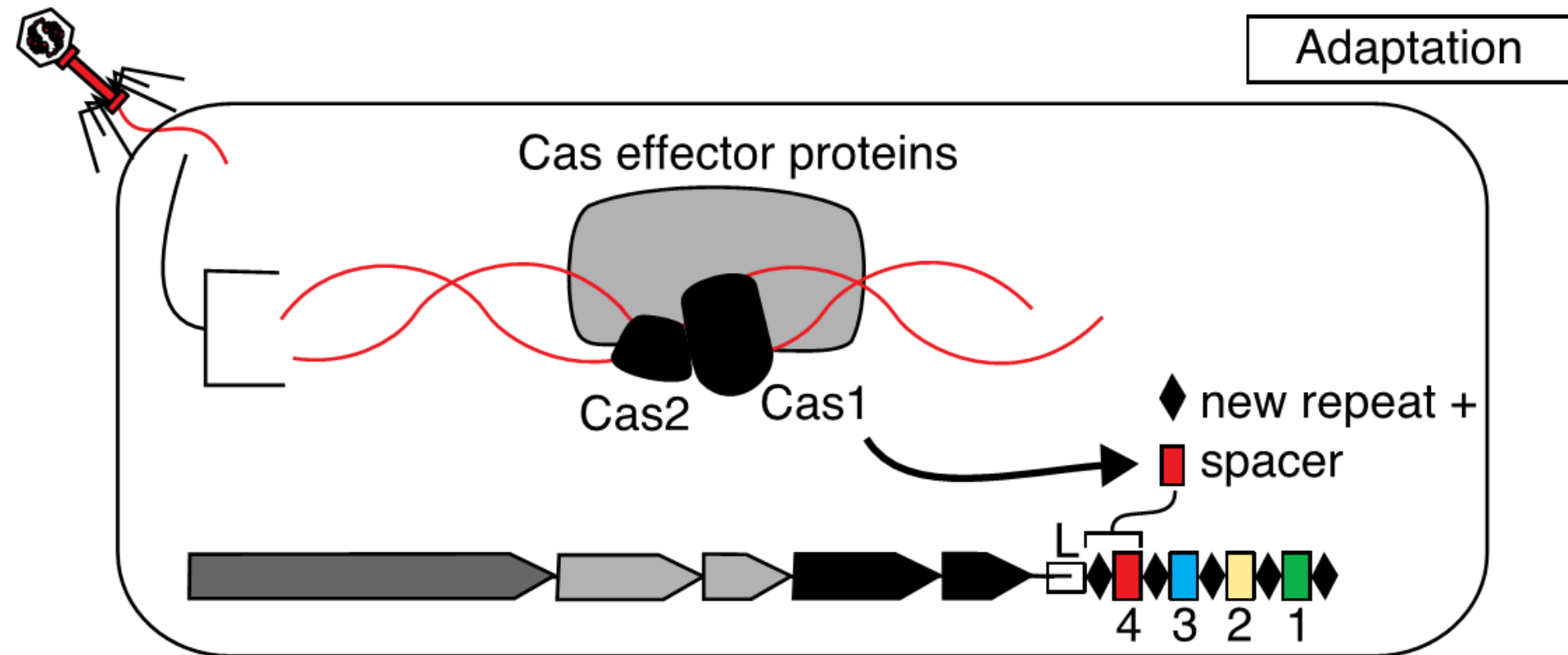
CRISPR associated (**Cas**) protein



Cas encodes proteins involved in the three stages of CRISPR-encoded immunity, namely adaptation, expression and interference

CRISPR-Cas systems and adaptive immunity

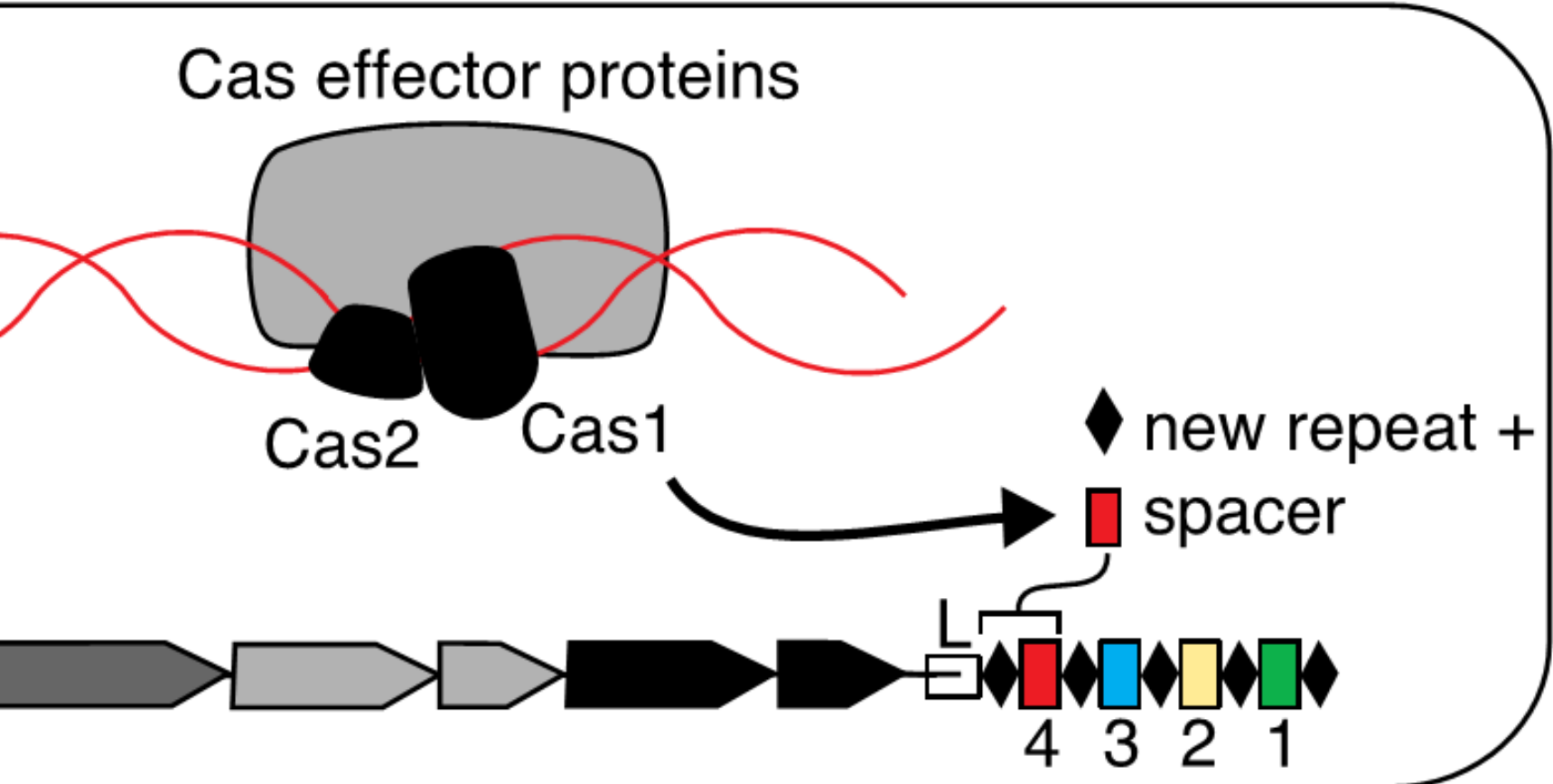
1. Adaptation



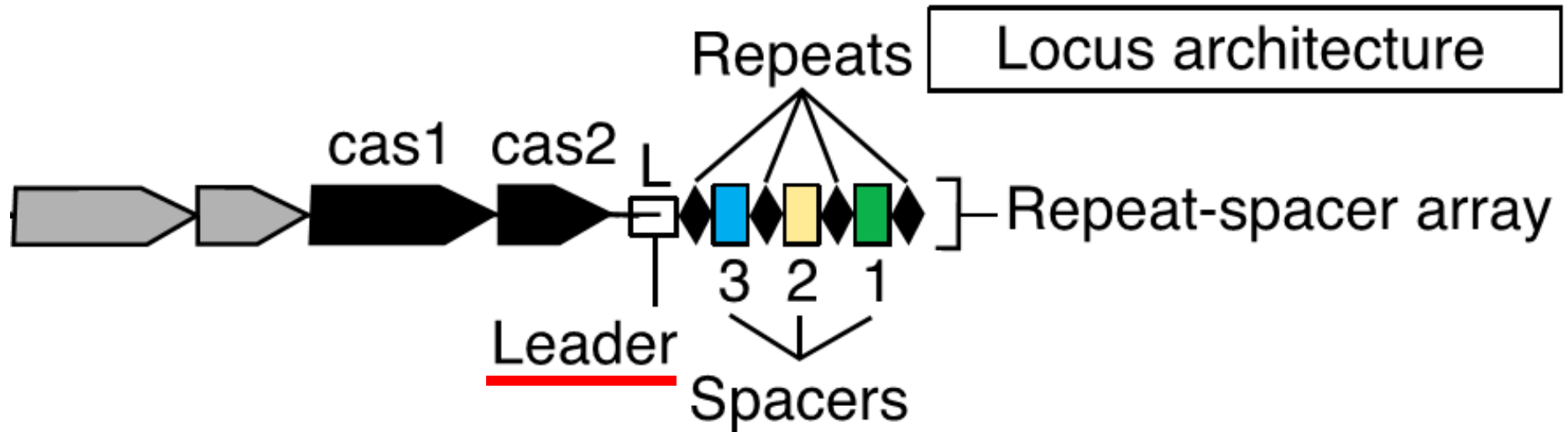
1. Adaptation

CRISPR-Cas protein machinery involved process-identification and integrating the short segments of foreign invading DNA (termed spacers) into the CRISPR array locus in host cell genome, forming an immunological memory to invading DNA molecules.

Adaptation



2. Expression

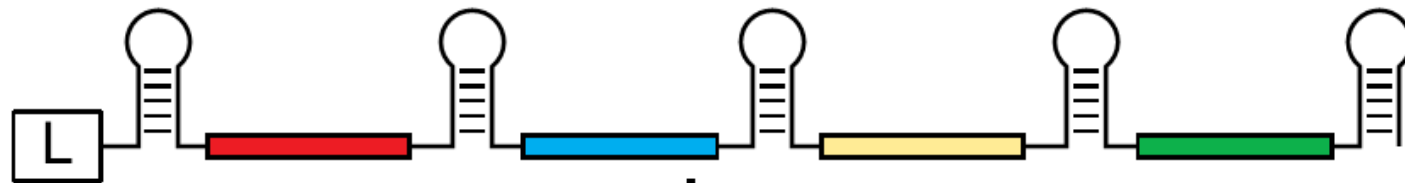


2. Expression

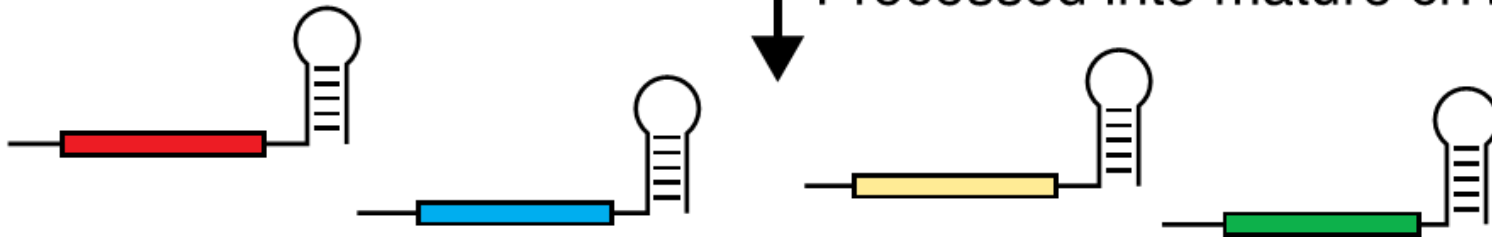
Expression



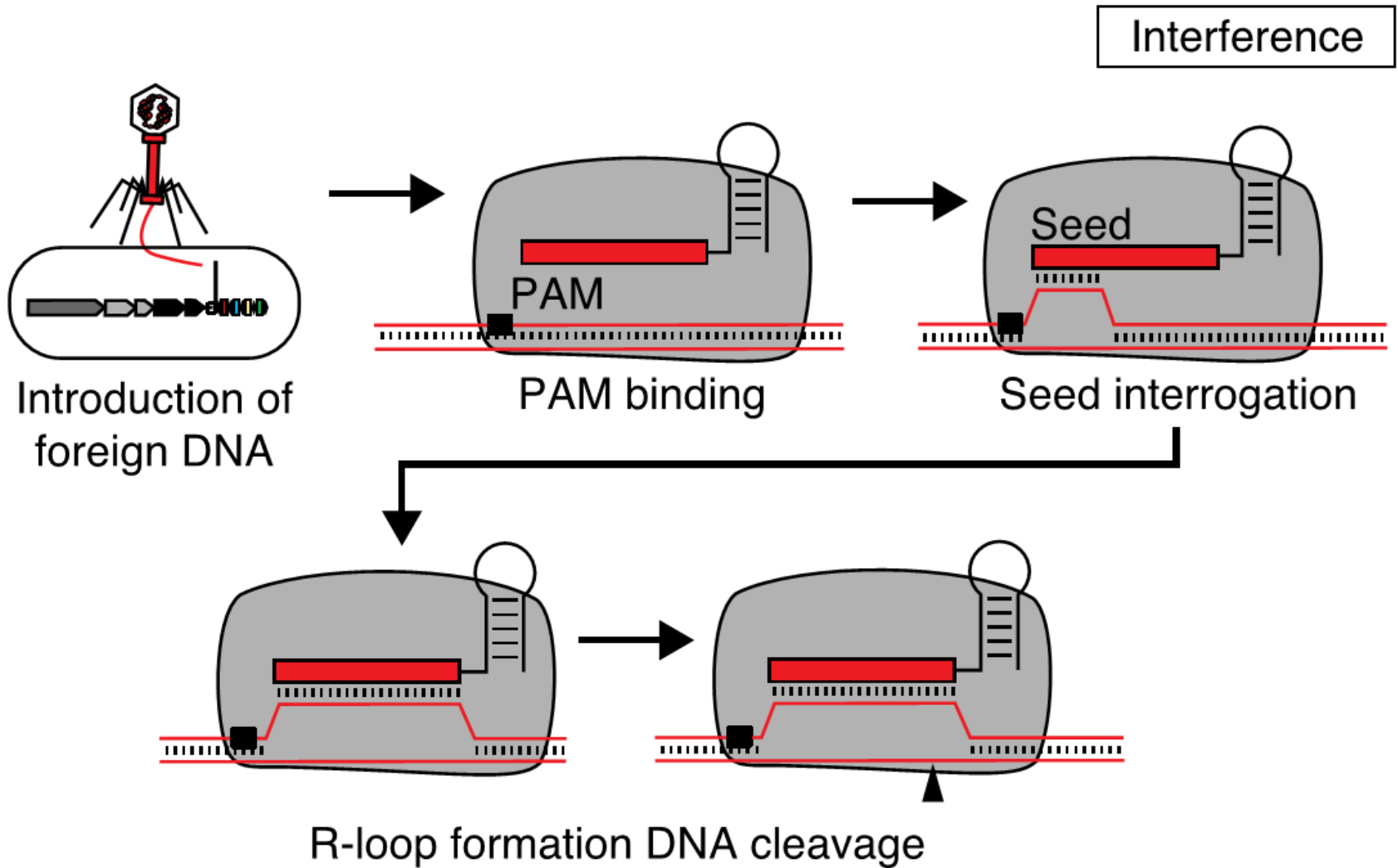
↓
Transcribed into pre-crRNA



↓
Processed into mature crRNAs



3. Interference



CRISPR-Cas systems and adaptive immunity. CRISPR repeats, together with CRISPR spacers, constitute repeat-spacer arrays that define clustered regularly interspaced short palindromic repeats (CRISPRs). These CRISPR arrays are typically flanked by CRISPR associated sequences (cas) that encode Cas proteins involved in the three stages of CRISPR-encoded immunity, namely **adaptation**, **expression** and **interference**.

During **adaptation**, Cas proteins sample invasive DNA, leading to the genesis of a new repeat-spacer unit that is inserted in a polarized manner in the CRISPR array.

During the second stage — **expression** — the CRISPR array is transcribed into a full pre-crRNA transcript that is processed into small, mature, interfering CRISPR RNAs (crRNAs).

In the third stage: **interference**, crRNAs guide Cas effector proteins towards complementary nucleic acids for sequence-specific targeting. Interaction between the interference complex and the target nucleic acid is typically initiated by binding to the protospacer adjacent motif (PAM), which triggers interrogation of flanking DNA by the loaded crRNA. If complementarity extends beyond the seed sequence, an R-loop is formed, and nickase domains within Cas effector proteins cleave the target DNA.

CRISPR/Cas *in vivo*:

Bacterial Adaptive Immunity

Evolutionary significance

Found in 90% of archaea and 40% of bacteria tested so far

CRISPR/Cas

Developed as Molecular Biology Tools
used for genome editing in Eukaryotic

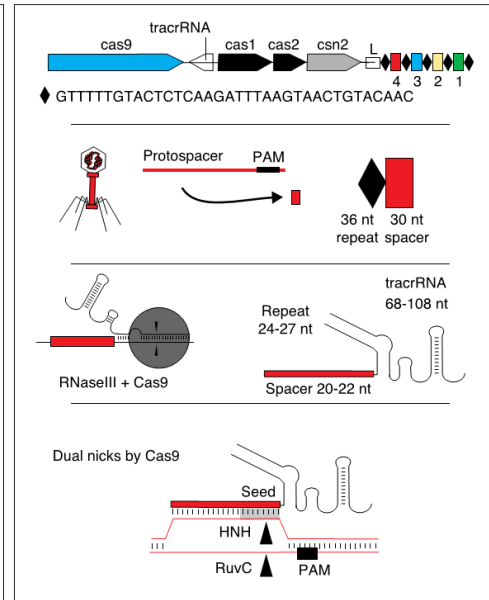
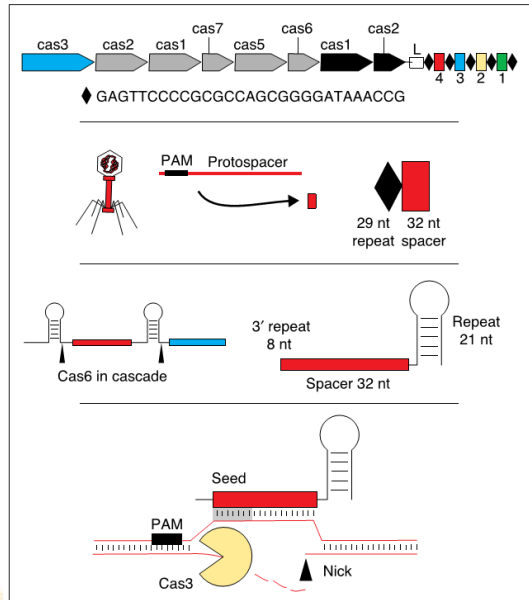
Diversity of CRISPR-Cas Machines in Prokaryotes

Class I

Class II

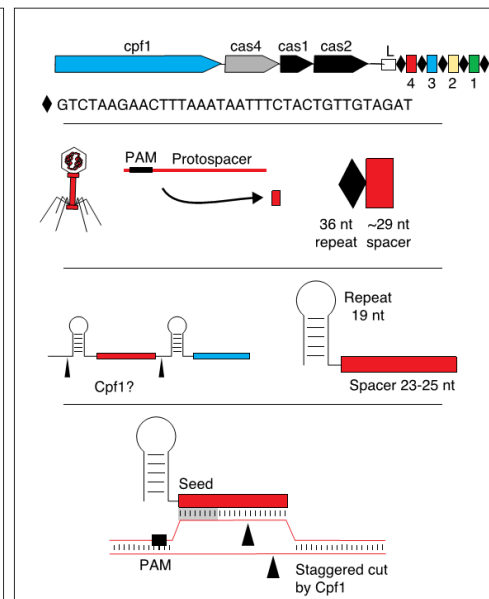
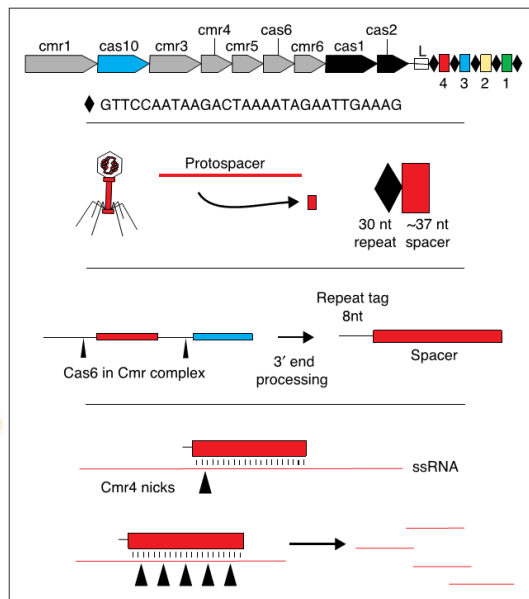
Type I-E *E. coli* K12

Type II-A *S. thermophilus* LMD-9



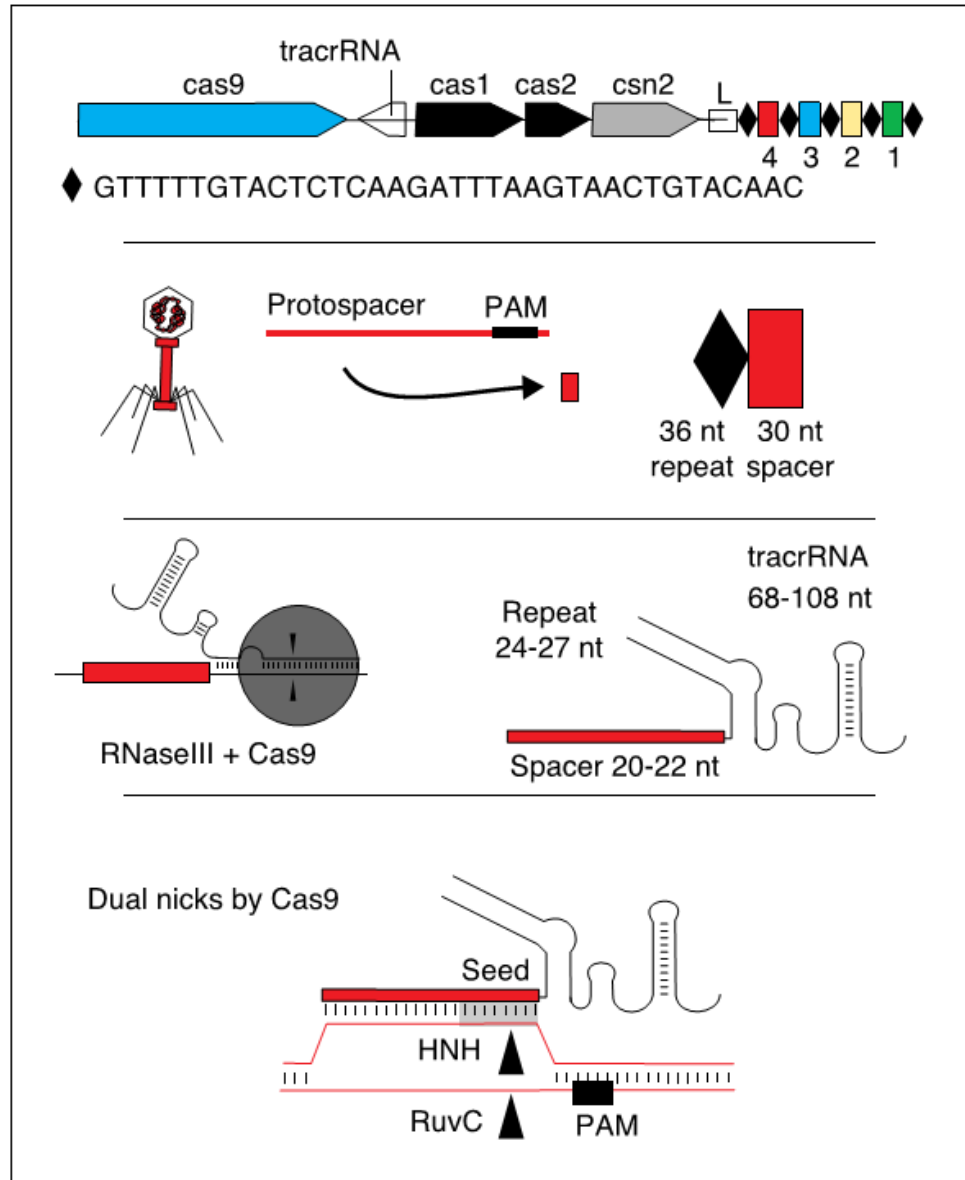
Type III-B *P. furiosus* DSM 3638

Type V *F. novicida* U112



Class II type Cas system

Type II-A *S. thermophilus* LMD-9



Engineering Type II CRISPR/Cas system

Three-component System

- Cas9
- trans-activating
CRISPR RNA
(tracrRNA)
- crRNA

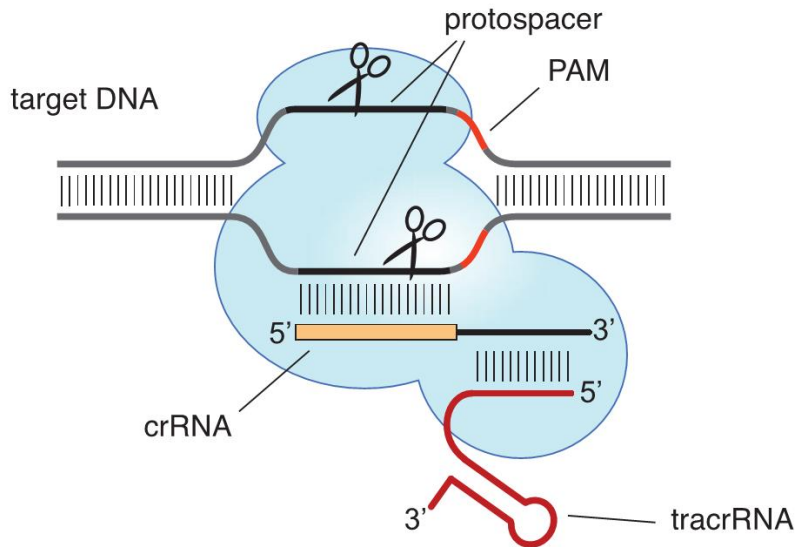


A Programmable Dual-RNA–Guided DNA Endonuclease in Adaptive Bacterial Immunity

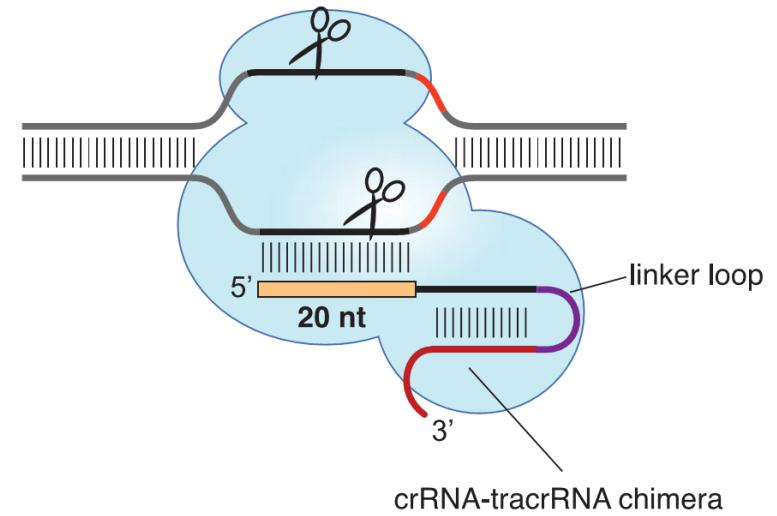
Martin Jinek,^{1,2*} Krzysztof Chylinski,^{3,4*} Ines Fonfara,⁴ Michael Hauer,^{2†}
Jennifer A. Doudna,^{1,2,5,6‡} Emmanuelle Charpentier^{4‡}

Clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated (Cas) systems provide bacteria and archaea with adaptive immunity against viruses and plasmids by using CRISPR RNAs (crRNAs) to guide the silencing of invading nucleic acids. We show here that in a subset of these systems, the mature crRNA that is base-paired to trans-activating crRNA (tracrRNA) forms a two-RNA structure that directs the CRISPR-associated protein Cas9 to introduce double-stranded (ds) breaks in target DNA. At sites complementary to the crRNA-guide sequence, the Cas9 HNH nuclease domain cleaves the complementary strand, whereas the Cas9 RuvC-like domain cleaves the noncomplementary strand. The dual-tracrRNA:crRNA, when engineered as a single RNA chimera, also directs sequence-specific Cas9 dsDNA cleavage. Our study reveals a family of endonucleases that use dual-RNAs for site-specific DNA cleavage and highlights the potential to exploit the system for RNA-programmable genome editing.

Cas9 programmed by crRNA:tracrRNA duplex



Cas9 programmed by single chimeric RNA



single guide RNA (sgRNA)

Streptococcus pyogenes
酿脓链球菌

Engineering Type II CRISPR/Cas system

Three-component
System

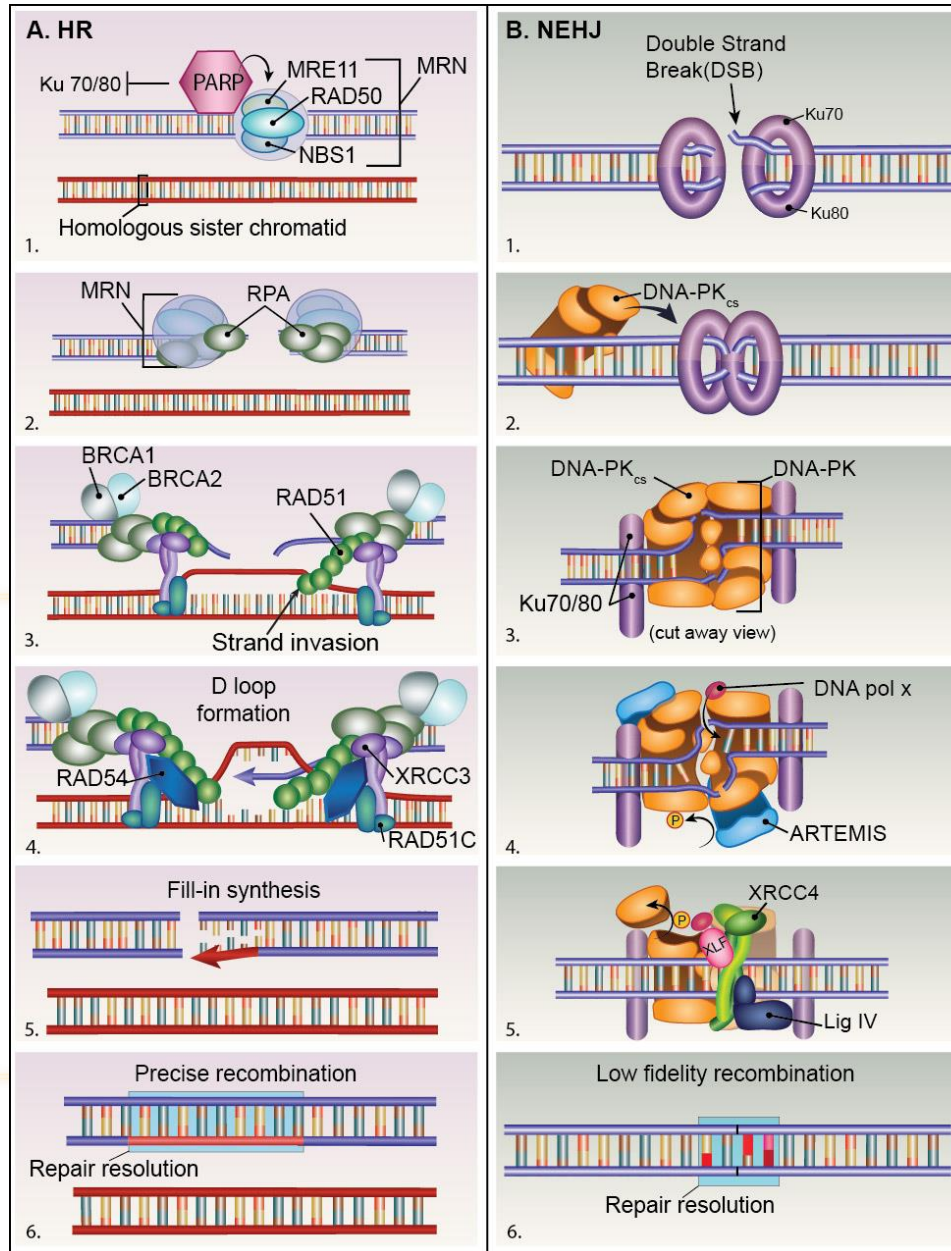
- Cas9
- trans-activating
CRISPR RNA
(tracrRNA)
- crRNA



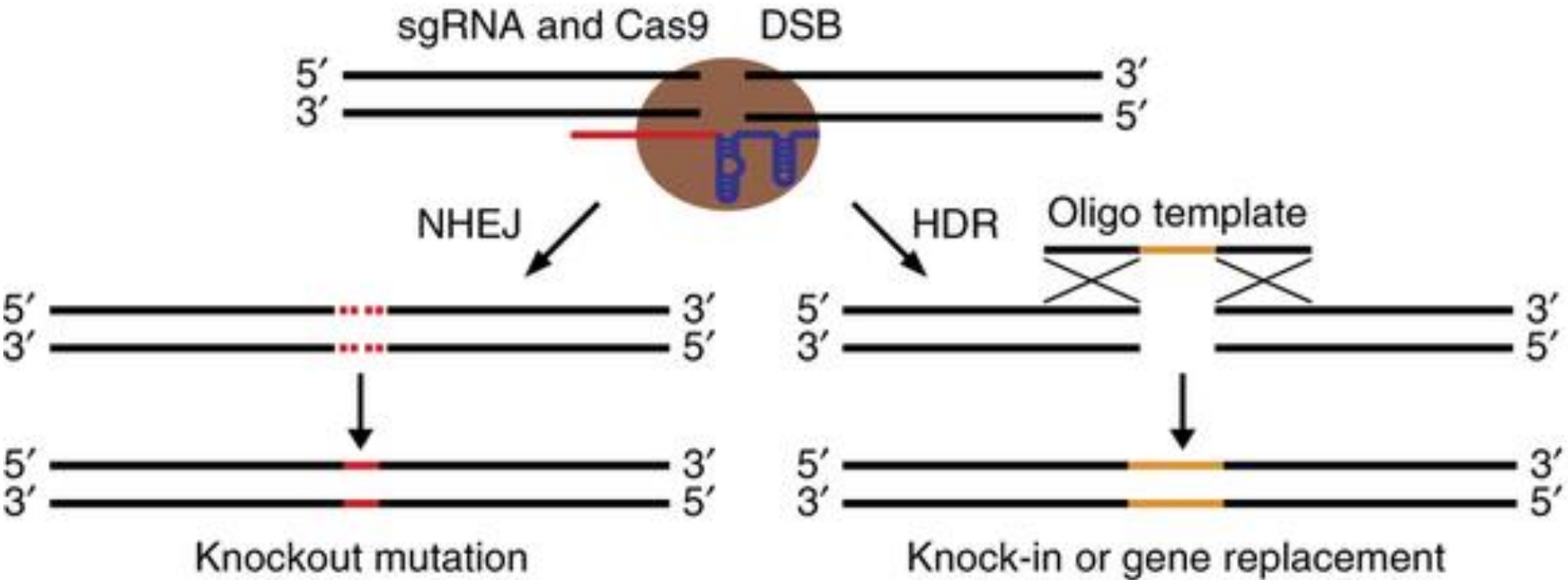
Two-component
System

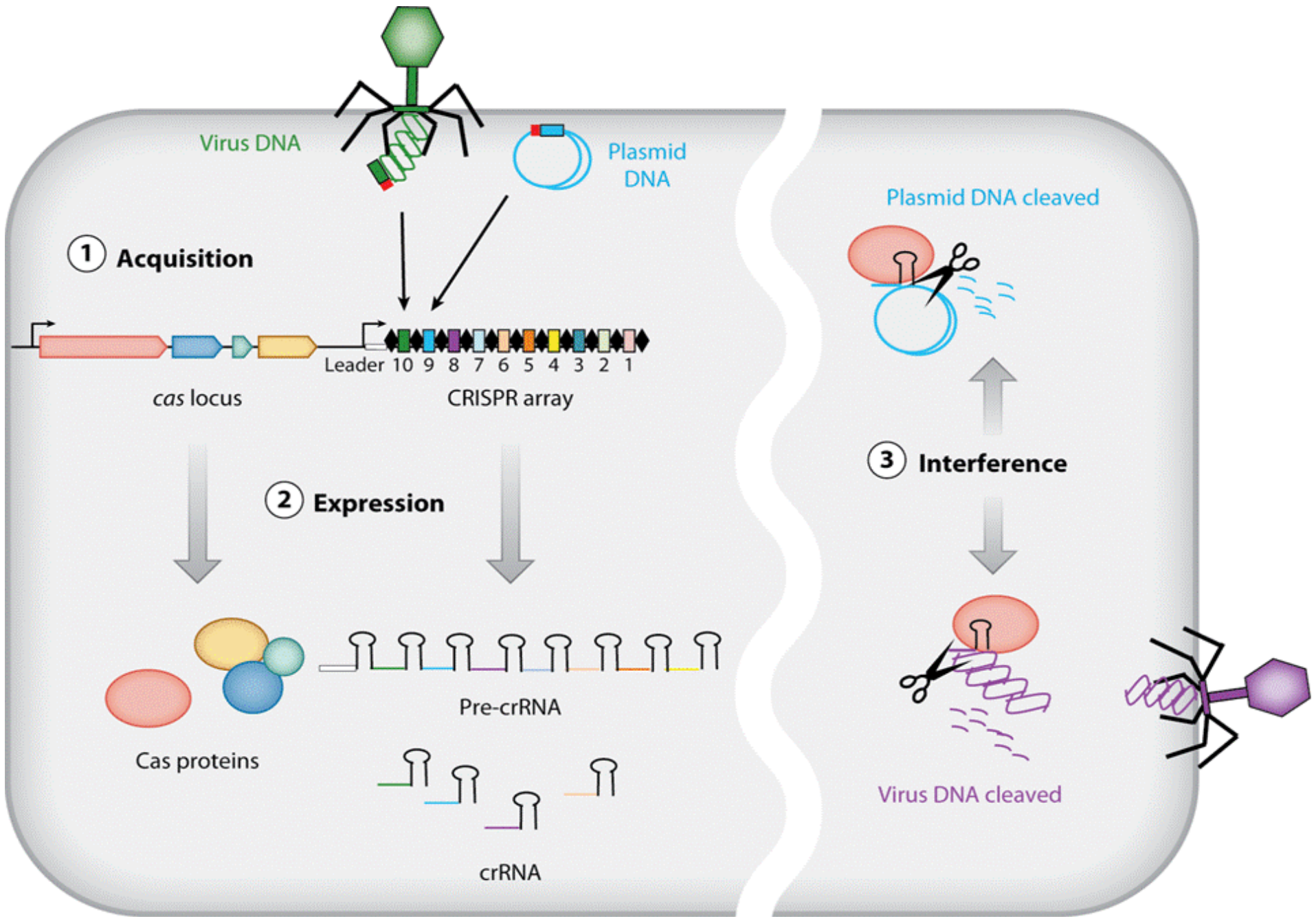
- Cas9
- sgRNA

DNA repairing machine in eukaryote



Mutation introduced in DNA repair





CRISPR-Cas systems and adaptive immunity. CRISPR repeats, together with CRISPR spacers, constitute repeat-spacer arrays that define clustered regularly interspaced short palindromic repeats (CRISPRs). These CRISPR arrays are typically flanked by CRISPR associated sequences (cas) that encode Cas proteins involved in the three stages of CRISPR-encoded immunity, namely **adaptation**, **expression** and **interference**.

During **adaptation**, Cas proteins sample invasive DNA, leading to the genesis of a new repeat-spacer unit that is inserted in a polarized manner in the CRISPR array.

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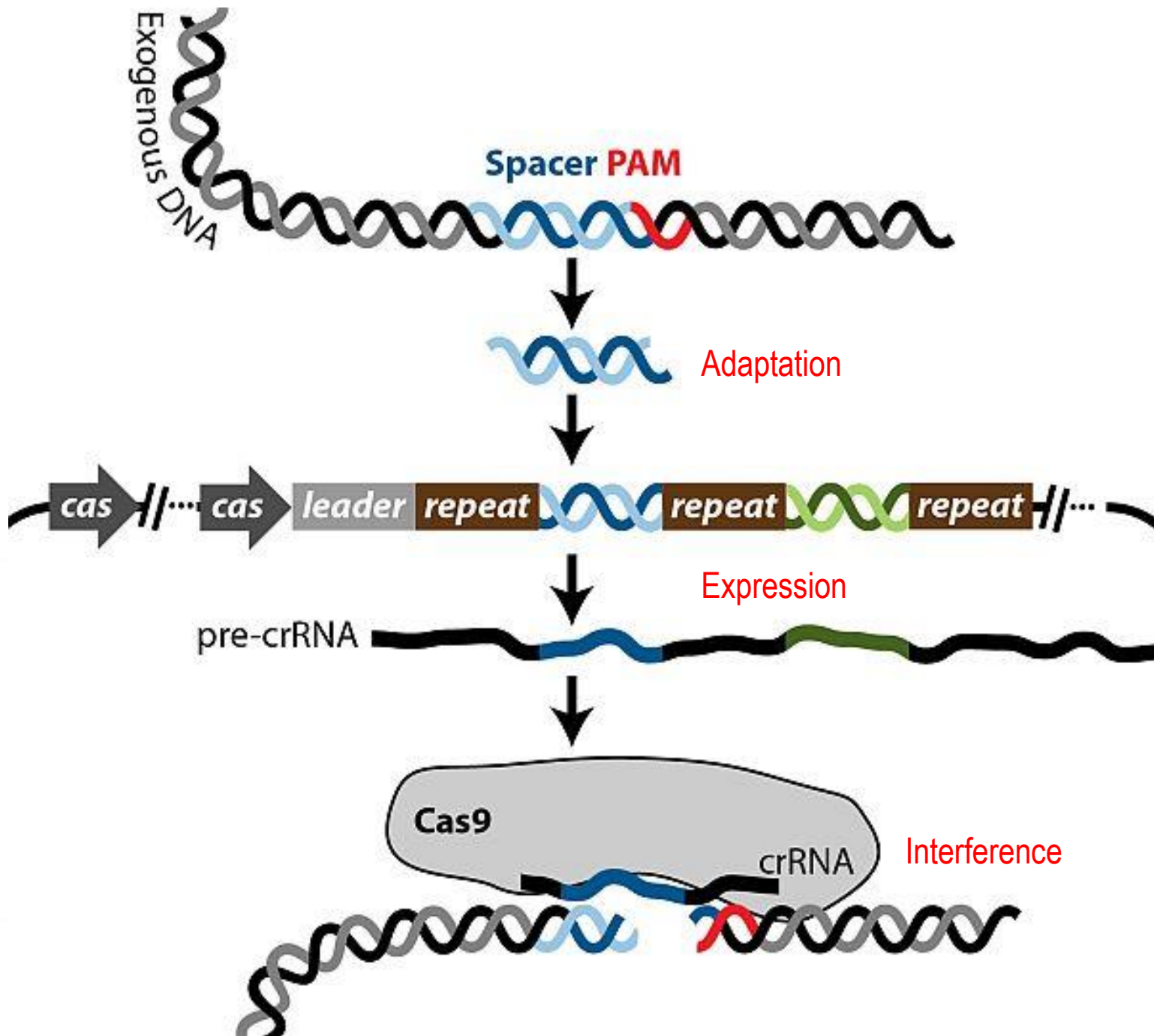
In the third stage: **interference**, crRNAs guide Cas effector proteins towards complementary nucleic acids for sequence-specific targeting. Interaction between the interference complex and the target nucleic acid is typically initiated by binding to the protospacer adjacent motif (PAM), which triggers interrogation of flanking DNA by the loaded crRNA. If complementarity extends beyond the seed sequence, an R-loop is formed, and nickase domains within Cas effector proteins cleave the target DNA.

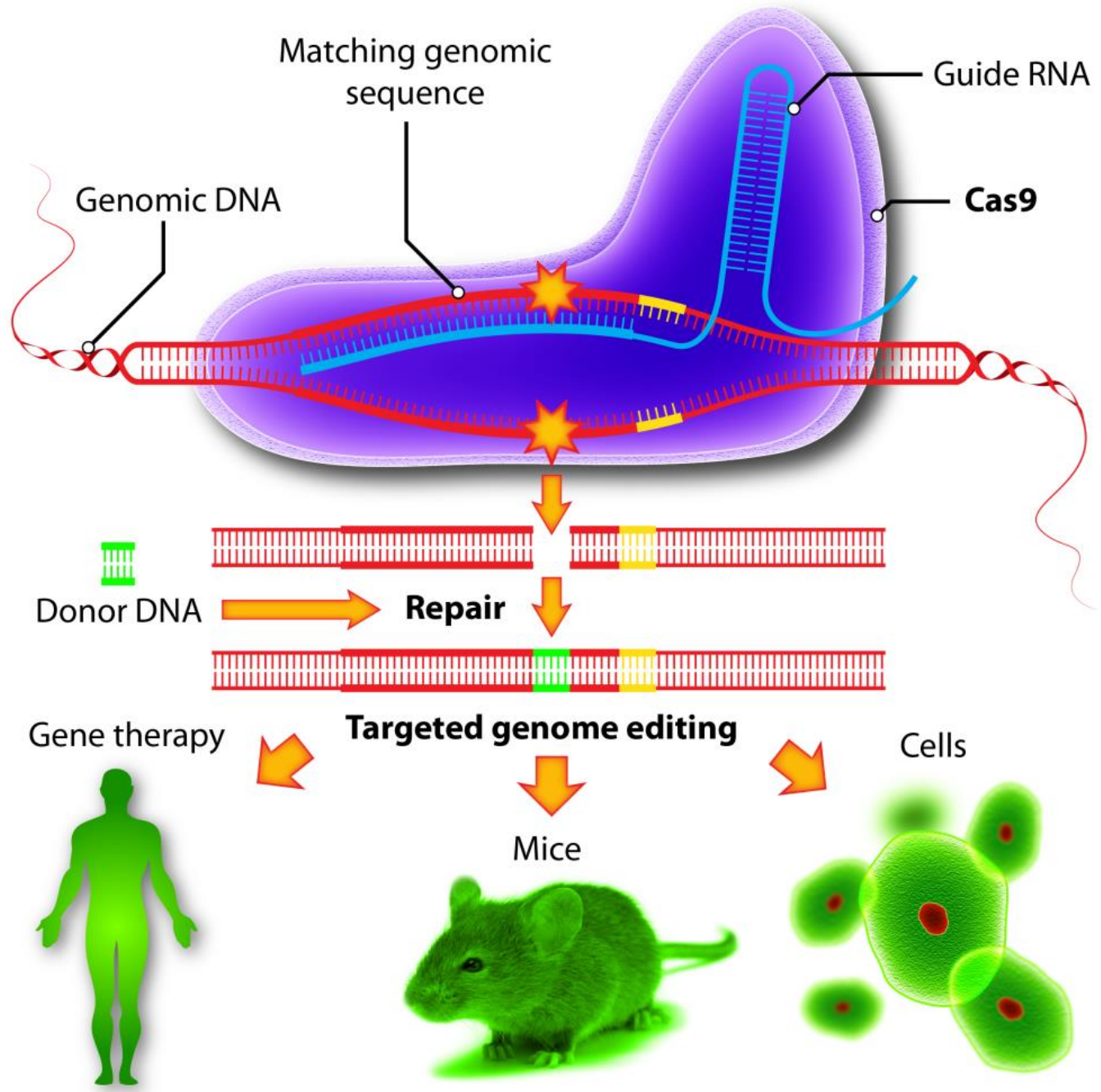
In the **acquisition/adaptation** phase, foreign DNA is incorporated into the bacterial genome at the CRISPR loci.

CRISPR loci is then transcribed and processed into crRNA during crRNA biogenesis. **Expression**

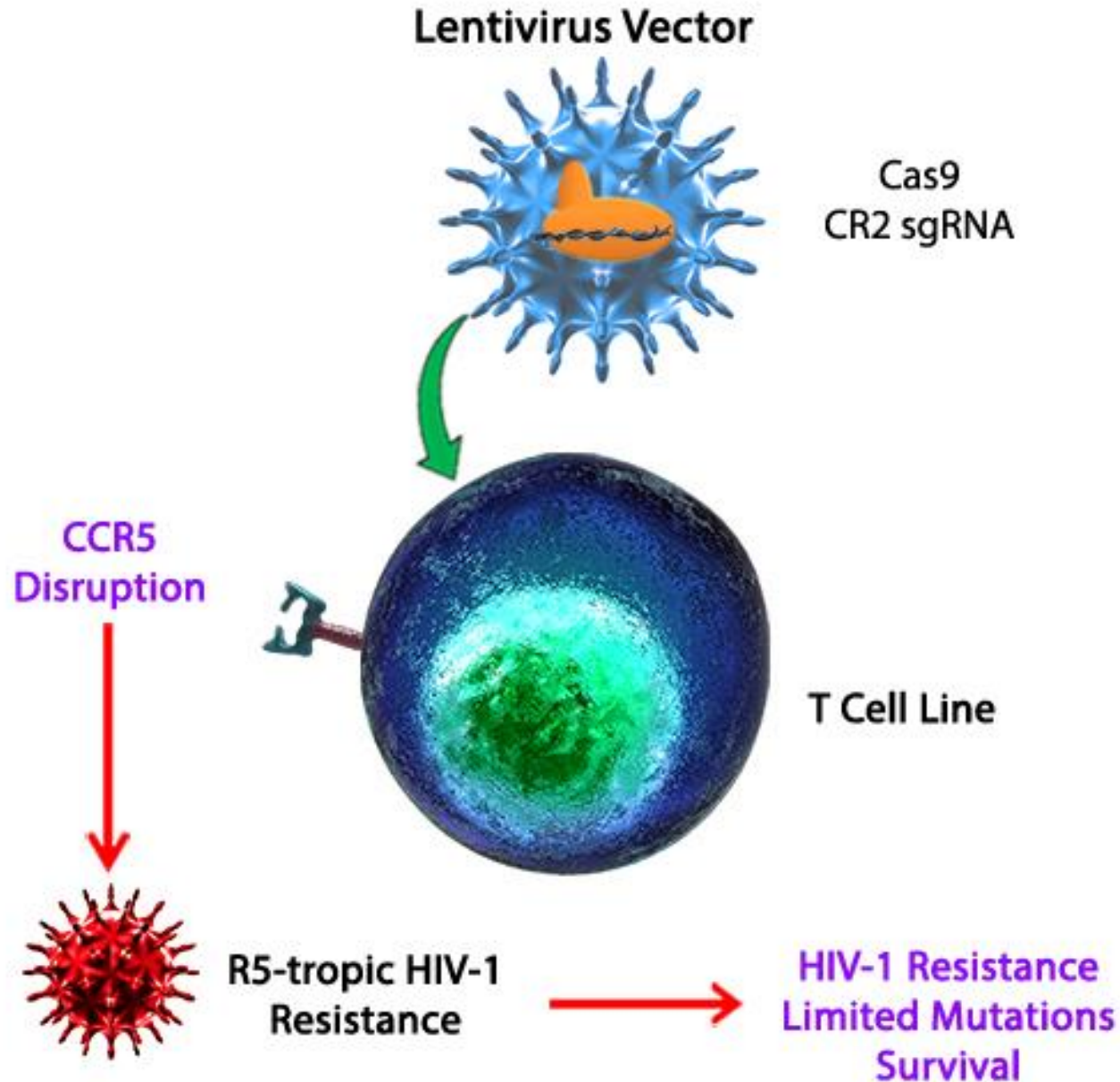
During **interference**, Cas9 endonuclease complexed with a crRNA and separate tracrRNA cleaves foreign DNA containing a 20-nucleotide crRNA complementary sequence adjacent to the PAM sequence.

Key

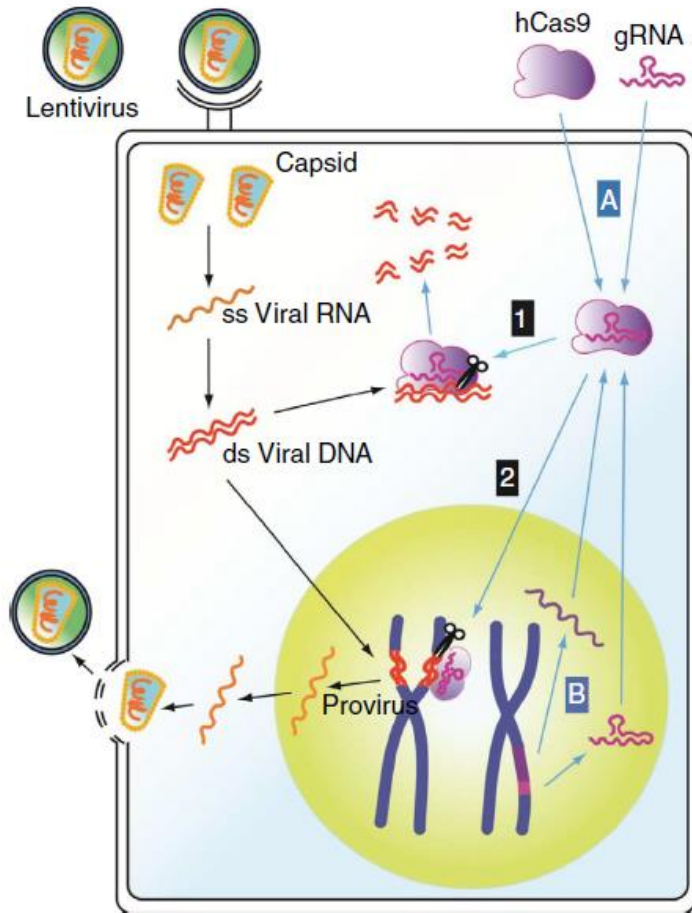




Modified cell resistant to HIV



Genome Editing Cuts Out HIV



Researchers use the **CRISPR/Cas9** method to remove the virus from the host genome in human cell lines.

W. Hu et al., "RNA-directed gene editing specifically eradicates latent and prevents new HIV-1 infection," *PNAS*, doi:10.1073/pnas.1405186111, 2014.

CRISPR Takes the PERV Out of Pig Organs

GENOME EDITING

Genome-wide inactivation of porcine endogenous retroviruses (PERVs)

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The shortage of organs for transplantation is a major barrier to the treatment of organ failure. Although porcine organs are considered promising, their use has been checked by concerns about the transmission of porcine endogenous retroviruses (PERVs) to humans. Here we describe the eradication of all PERVs in a porcine kidney epithelial cell line (PK15). We first determined the PK15 PERV copy number to be 62. Using CRISPR-Cas9, we disrupted all copies of the PERV *pol* gene and demonstrated a >1000-fold reduction in PERV transmission to human cells, using our engineered cells. Our study shows that CRISPR-Cas9 multiplexability can be as high as 62 and demonstrates the possibility that PERVs can be inactivated for clinical application of porcine-to-human xenotransplantation.

“一段重复序列的故事”
